



Utrecht University

Writing Assignment Master Drug Innovation

**Anti-cancer mechanisms of the most used drugs
worldwide: old drugs, new insights**

By

Gijs van Slobbe

6907903

Daily supervisor and 1st accessor

Dr. E.E (Ed) Moret

Department of Pharmaceutical Sciences
Chemical Biology and Drug Discovery group

e.e.moret@uu.nl

2st accessor

Prof. Dr. A.F.M. (Maarten) Altelaar
Department of Pharmaceutical Sciences
Biomolecular Mass Spectrometry and
Proteomics Group

m.altelaar@uu.nl

1 Abstract

Despite recent therapeutical improvements, cancer remains the second leading cause of death. Therefore there is an emerging need for novel cancer treatments. Development of novel anti-cancer drugs to fulfill the need for novel treatments is expensive, takes a long time and goes along with a low change of gaining marketing authorization. An alternative cheaper and faster approach to fulfill the need of new anti-cancer drugs is drug repurposing. The worldwide most used drugs could be potential candidates for drug repurposing since their intensive use provides extensive safety data. Therefore, mechanisms that clarify anti-cancer activity of eight of the most used drugs worldwide and the drug classes where they belong to were investigated in this study. Literature review showed that the drug gabapentin and the drug classes statins, renin angiotensin system (RAS) inhibitors, selective betablockers (BBs), dihydropyridine calcium channel blockers (CCBs), biguanides and protein pump inhibitors (PPIs) showed potential anti-tumorigenic effects *in vitro* and *in vivo*. The anti-cancer effects were established by targeting of several cancer hallmarks including: sustaining proliferation, induction of invasion and metastasis, avoiding immune destruction, induction of angiogenesis, deregulation of cellular energetics, genome instability and resisting cell death. Beside the drug effects on cancer hallmarks, it was shown that some drugs induced the delivery of chemotherapeutic drugs. In addition, the drug classes statins, RAS inhibitors, selective BBs, biguanides and PPIs showed combinational effects with conventional anti-cancer drugs, which increases the changes of successful drug repurposing since drugs have higher changes to get authorized as repurposed drugs if they will be combined. Overall, anti-cancer mechanisms were identified for all the investigated drugs. These mechanisms support suggested anti-cancer activity of the investigated drugs and therefore it was concluded that the drug gabapentin and the drug classes statins, RAS inhibitors, selective betablockers, dihydropyridine CCBs, biguanides and PPIs have the potential to be used for cancer drug repurposing. Since the drug classes statins, RAS inhibitors, selective BBs, biguanides and PPIs showed combinational effects with conventional treatments, these drug classes were especially considered as potential agents for cancer drug repurposing.

2 Layman's summary

Ondanks jaren onderzoek is kanker nog steeds een moeilijk te behandelen en dodelijke ziekte. Om deze reden zijn er nieuwe kankertherapieën nodig die de behandeling van kanker kunnen verbeteren. Voor deze nieuwe kankertherapieën zouden nieuwe kankermedicijnen ontwikkeld kunnen worden, maar de ontwikkeling van nieuwe kankermedicijnen is duur, kost veel tijd en deze nieuwe medicijnen hebben vaak maar een kleine kans om uiteindelijk op de markt te komen. Een alternatieve methode om nieuwe kankermedicijnen te ontdekken, is het toepassen van medicijnen die al gebruikt worden voor de behandeling van andere ziektes, ook wel bekend als drug repurposing. Drug repurposing is een goedkopere en snellere manier om nieuwe medicijnen te ontdekken doordat er onder anderen geen tijd en geld nodig is om het medicijn te ontwikkelen. De beste kandidaten voor drug repurposing zijn medicijnen die veel gebruikt worden, doordat het vele gebruik goed inzicht geeft over de veiligheid en de bijwerkingen van deze medicijnen. Omdat veel gebruikte medicijnen veelbelovende zijn voor drug repurposing, is er in dit onderzoek literatuur onderzoek gedaan of de wereldwijd meest gebruikte medicijnen anti-kanker effecten hebben en of er mechanismes zijn beschreven die deze anti-kanker effecten kunnen verklaren.

In literatuur was gevonden dat het medicijn gabapentine en de drugklassen statines, renine angiotensine systeem (RAS) remmers, selectieve bètablokkers, dihydropyridine calcium kanaal blokkers (CKBs), biguaniden en proton pomp remmers (PPR) anti-kanker effecten lieten zien in zowel laboratorium experimenten als in dierstudies. De drugs lieten anti-kanker effecten zien door verschillende kenmerken van kanker te beïnvloeden. De kankerkenmerken die werden beïnvloed door de medicijnen waren: het ontregelde energiemetabolisme van de kankercellen, ongeremde deling, het voorkomen van celdood, het induceren van invasie en metastase, het induceren van bloedvaatontwikkeling, het ontwijking van de immuunrespons tegen de kanker en het aangepaste herstel van DNA schade in kankercellen. Naast deze directe effecten op kankercellen, lieten sommige medicijnen zien dat ze er voor zorgden dat conventionele kankermedicijnen beter in de tumor terechtkwamen en daardoor de effectiviteit van deze medicijnen verbeterden. Ook lieten de drugklassen statines, RAS remmers, selectieve bètablokkers, biguaniden en PPRs combinatie effecten zien met conventionele medicijnen, wat de kans op succesvolle drug repurposing vergroot omdat medicijnen een grotere kans hebben om ge-repurposed te worden indien ze als combinatie gebruikt gaan worden.

Samengevat werden er voor het medicijn gabapentine en de drugklassen statines, RAS remmers, selectieve bètablokkers, dihydropyridine CKBs, biguaniden en PPRs anti-kanker mechanismes geïdentificeerd. Daarom werd er geconcludeerd dat deze drugklassen de potentie hebben om gebruikt te worden voor kanker drug repurposing. Omdat statines, RAS remmers, selectieve bètablokkers, biguaniden en PPRs combinatie effecten lieten zien met conventionele therapieën werd er geconcludeerd dat deze drug klassen de grootste potentie hebben om gebruikt te worden voor kanker drug repurposing.

3 Table of contents

1	Abstract	2
2	Layman’s summary.....	3
3	Table of contents.....	4
4	Introduction.....	5
5	Main text	8
5.1	Statins.....	8
5.2	Renin angiotensin system inhibitors	9
5.3	Selective betablockers.....	11
5.4	Dihydropyridine calcium channel blockers	12
5.5	Gabapentin	13
5.6	Metformin	14
5.7	Protein pump inhibitors	16
6	Discussion	18
7	Bibliography.....	20

4 Introduction

Cancer is the second leading cause of death worldwide¹. In most cases, cancer is still incurable and therefore, there is an emerging need for novel treatments. Development of novel drugs to fulfill the need for novel treatments is expensive, takes a long time and has only a limited change to gain marketing authorization². For example, 95% of the anti-cancer drugs that are tested in phase I trials will not gain marketing authorization³. This low change of successful drug development can partly be declared by the flexibility of cancer cells to adapt themselves upon treatment. As shown in Figure 1, tumor growth is dependent on ten different biological processes which are known as the hallmarks of cancer⁴. These hallmarks can be targeted therapeutically, but a notable number of these targeted therapies only showed transiently effects since cancer cells showed to have the flexibility to upregulate other hallmarks when a certain hallmark was targeted with targeted therapies⁴. For example, inhibition of the hallmark angiogenesis initially showed promising effects in some preclinical cancer models^{5,6}, but these studies showed over time upregulation of the cancer hallmark “activation of invasion and metastasis” which resulted in severe metastasis and treatment relapse. Since it is challenging to develop effective new anti-cancer drugs, it is understandable that novel drugs are not always as effective as hoped. Indeed there is skepticism whether recently approved anti-cancer drugs really have beneficial effects compared to conventional drugs⁷⁻¹⁰. In summary, it is challenging, expensive and it takes a long time to develop effective new anti-cancer drugs. However, the earlier new treatments will be available, the earlier cancer patients would benefit from it. Therefore it could be helpful to find alternative faster and cheaper approaches to develop novel cancer treatments.

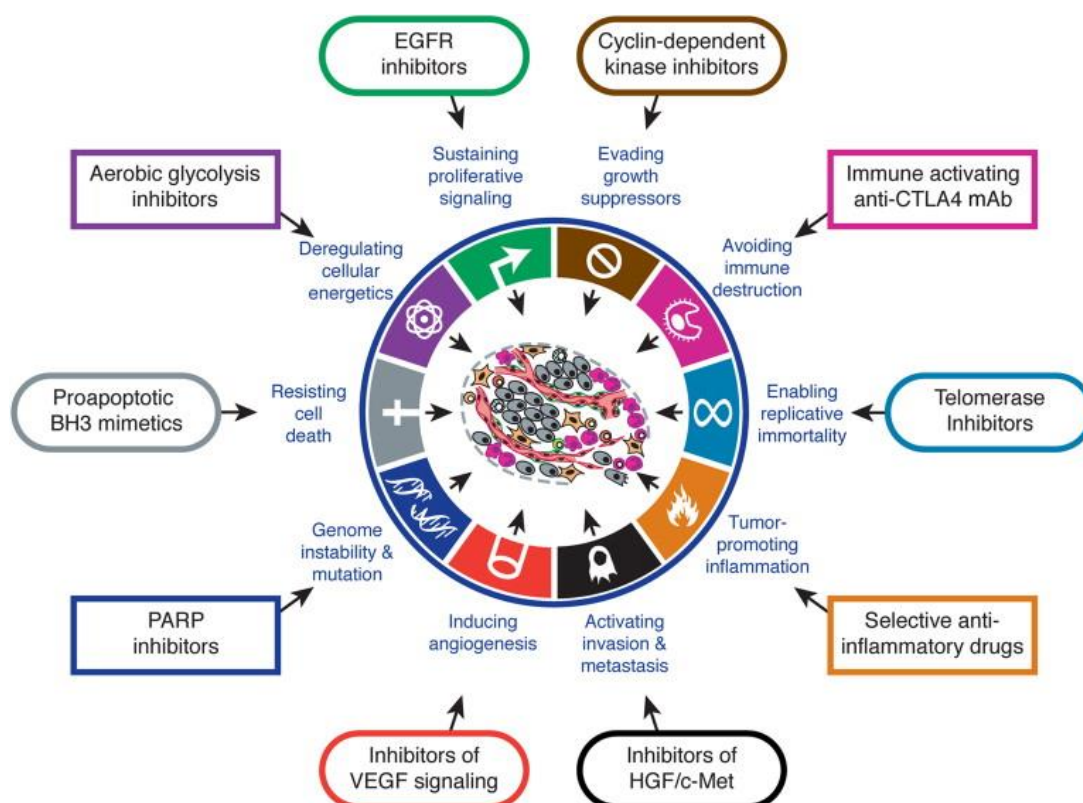


Figure 1. The ten hallmarks of cancer and examples of therapeutic agents to target them⁴.

An alternative approach to develop novel cancer treatments is drug repositioning/repurposing of approved drugs^{11,12}. Repositioning of approved drugs enables quick entry to clinical trials since these drugs already went through extensive toxicity and safety profiling¹³. In addition, the authorization

process of approved drugs is estimated to be 50-60 percent cheaper compared with novel compounds¹⁴. Therefore, repositioning of approved drugs is a promising and cost-effective approach to develop novel cancer therapies. Often prescribed drugs are good candidates for drug repositioning since high drug utilization provides extensive safety data. The most used drugs worldwide, see table 1, could therefore be promising agents for drug repurposing. Interestingly, anti-cancer activity had been suggested for eight of the ten drug classes that are involved in this list^{12,15-23}. It is therefore interesting to further investigate the potential of these drugs for cancer drug repurposing. It is especially interesting to identify mechanisms that clarify anti-cancer activity. These mechanisms will provide evidence of anti-cancer activity and will help to estimate whether the drugs are candidates for cancer drug repurposing. Therefore, mechanisms that support anti-cancer effects of the drug classes: statins, renin angiotensin system (RAS) inhibitors, dihydropyridine calcium channel blockers (CCBs), selective betablockers (BBs), biguanides PPIs and the drug gabapentin were further investigated in this literature review.

In order to clarify suggested anti-cancer mechanisms of each drug, it was questioned (1) which hallmark of cancer is affected by exposure to the drug and (2) what is the proposed mechanism that clarifies this effect?

Relevant literature was searched using the literature databases pubmed and scopus. In the first place, review articles were searched in pubmed using key words like “cancer”, “statins” (and all other drug classes) “metoprolol” (and all other drugs) etc. Papers were partly selected for further investigation based on the number of citations according to scopus. For some papers, it was decided to look for follow-up studies or more recent findings by checking which papers cited the respectable paper using scopus.

Table 1. Characteristics of the top 10 most prescribed drugs in 2019²⁴.

Drug name	Prescriptions in 2019	Indication	Drug class	Mode of action	Effect
Atorvastatin	112,104,359	Hyperlipidemia	Statins, Lipid lowering drugs	Inhibition of HMG-CoA reductase	Lowering of cholesterol synthesis
Levothyroxine	102,595,103	Hypothyroidism	Synthetic hormone	Molecule with the same effect as natural occurring T4	Increases the amount of bioavailable T4
Lisinopril	91,862,708	acute myocardial infarction, hypertension and as an adjunct therapy for heart failure.	ACE inhibitor, RAS inhibitors	Inhibition of angiotensin converting enzyme	Decreased conversion of angiotensin I to angiotensin II resulting in downstream lowering effect on blood pressure
Metformin	85,739,443	Diabetes type II	Biguanides, antihyperglycemic drugs	Inhibition of mitochondrial complex I	Increased ADP:ATP and AMP:ATP ratios, resulting in AMPK activation, which regulates glucose metabolism
Metoprolol	74,578,817	angina, heart failure, myocardial infarction, atrial fibrillation, atrial flutter and hypertension	Selective betablocker, Betablockers	Antagonist of the β 1-adrenergic receptor and negligible antagonism of	Receptor inhibition results in lowered cardiac output

				β 2-adrenergic receptor	
Amlodipine	73,542,114	Hypertension, Coronary artery disease, Chronic stable angina, Vasospastic angina and Angiographically documented coronary artery disease	dihydropyridine calcium channel blockers, calcium channel blockers	Antagonist of calcium channels in vascular smooth muscle and cardiac muscle	Lowered contraction of vascular smooth muscle and cardiac muscle, resulting in lowered blood pressure
Albuterol	60,679,987	Prevention of bronchospasm due to bronchial asthma, chronic bronchitis, reversible obstructive airway disease, and other chronic bronchopulmonary disorders in which bronchospasm is a complicating factor	β 2 adrenergic receptor agonist, Anti-asthmatic Agents	Agonist of the β 2-adrenergic receptor.	Relaxation of airway smooth muscle.
Omeprazole	52,546,641	gastroesophageal reflux disease (GERD) and drug-induced peptic ulcers	Proton pump inhibitors, Acid secretion inhibitors	Inhibition H ⁺ /K ⁺ ATPase of parietal cells	Inhibition of acid secretion
Losartan	51,773,869	Hypertension, diabetic nephropathy and hypertension with left ventricular hypertrophy,	Angiotensin receptor type I antagonist, RAS inhibitors	Antagonist of angiotensin II receptor type I	prevention of angiotensin II binding causes vascular smooth muscle relaxation, lowering blood pressure
Gabapentin	47,149,505	Epilepsy, neuropathic pain	Anticonvulsant	Inhibition of α 2 δ -1 subunit of voltage-gated calcium channels	Inhibition α 2 δ -1 which results in less dense pre-synaptic voltage-gated calcium channels.

5 Main text

5.1 Statins

Statins are usually prescribed as lipid lowering drugs for patients with hyperlipidemia and/or with a high risk for atherosclerosis²⁵. Statins inhibit 3-hydroxy-3-methylglutaryl CoA reductase (HMGCR), the rate limiting enzyme of the mevalonate pathway. This pathway is responsible for cholesterol biosynthesis and therefore, inhibition of this pathway results in lowered cholesterol synthesis accompanied with lowered intracellular cholesterol levels, which in turn activates a negative feedback loop to lower circulating lipid concentrations²⁵. Beside its lipid lowering effect, statins also have an effect on other end-products of the mevalonate pathway like farnesyl-diphosphate (FPP) and geranylgeranyl-diphosphate (GGPP) isoprenoids, ubiquinone (coenzyme Q10, CoQ10) and dolichol²⁶. In cancer cells, it was shown that inhibition of the mevalonate pathway majorly affected the production of these end-products rather than the production of cholesterol²⁷⁻²⁹. Statin-induced inhibition of the production of isoprenoids, CoQ10 and dolichol has been suggested to be the mechanism underlying anti-cancer activity of statins. The mechanisms of the proposed anti-cancer activity of statins are described below and visualized in Figure 2. These mechanisms suggest that statins affect the cancer hallmarks sustaining proliferative signaling, deregulating of cellular energetics and invasion & metastasis.

Firstly, statins showed to induce apoptosis and reduce invasiveness by targeting isoprenoid synthesis. The isoprenoids FFP and GGP are commonly used for prenylation, a post-translational modification where FFP and GGP are added to cysteine residues of proteins³⁰. FFP and GGP function as lipophilic anchors of prenylated proteins, which enables proper localization of the proteins to cell membranes³¹⁻³³. An important class of prenylated proteins is the RAS GTPases superfamily, which includes tumor driver proteins like Ras and Rho GTPases^{31,32}. Statins showed previously to reduce protein prenylation of GTPases via inhibition of FFP and GGP^{26,34}, leading to apoptosis in some cancer cells³⁵⁻³⁸. Cytotoxic effects of statins were rescued by supplementation with GGPP and sometimes with FFP³⁵⁻³⁸, providing evidence that anti-cancer effects were established via inhibition of isoprenoid production. Beside cytotoxic effects, statins also showed to decrease invasion via inhibition of prenylation. In aggressive breast cancer cells, it was observed that statin treatment decreased invasion by reduced prenylation of RhoA and RAS^{39,40}.

In addition, statins have been shown to reduce proliferation, induce apoptosis and deregulate cellular energetics by inhibition of CoQ10 synthesis. CoQ10 is involved as electron carrier in the electron transport chain (ETC) and functions as a cofactor of dihydroorotate dehydrogenase (DHDOH), a rate limiting enzyme in pyrimidine synthesis. Statins have shown to specifically inhibit CoQ10 synthesis in pancreatic ductal adenocarcinoma, multiple myeloma and p53 deficient colon cancer²⁷⁻²⁹. Statin-induced inhibition of CoQ10 resulted in decreased pyrimidine synthesis by inhibition of DHDOH²⁷ and induced oxidative stress by ETC-mediated inhibition of oxidative phosphorylation (OXPHOS)^{27,28}. These processes decreased proliferation and induced apoptosis in the respectable cancer cells.

Finally, statins showed to induce apoptosis and affect the hallmark resisting cell death by targeting dolichol synthesis. Dolichol is involved in N-linked protein glycosylation as a carrier of oligosaccharides⁴¹. Inhibition of dolichol synthesis by statins can therefore affect N-linked protein glycosylation⁴². Statins showed anti-glioblastoma effects by affecting glycosylation of multi-drug resistance protein (MDR-1)⁴³, which made the tumor cells more sensitive to irinotecan. A comparable mechanism was found in FMS-like tyrosine kinase 3/ internal tandem duplication ((FLT3)/(ITD))

positive AML cells. These cells are hard to treat, but were sensitive for statins which was accompanied with decreased glycosylation of FLT3⁴⁴.

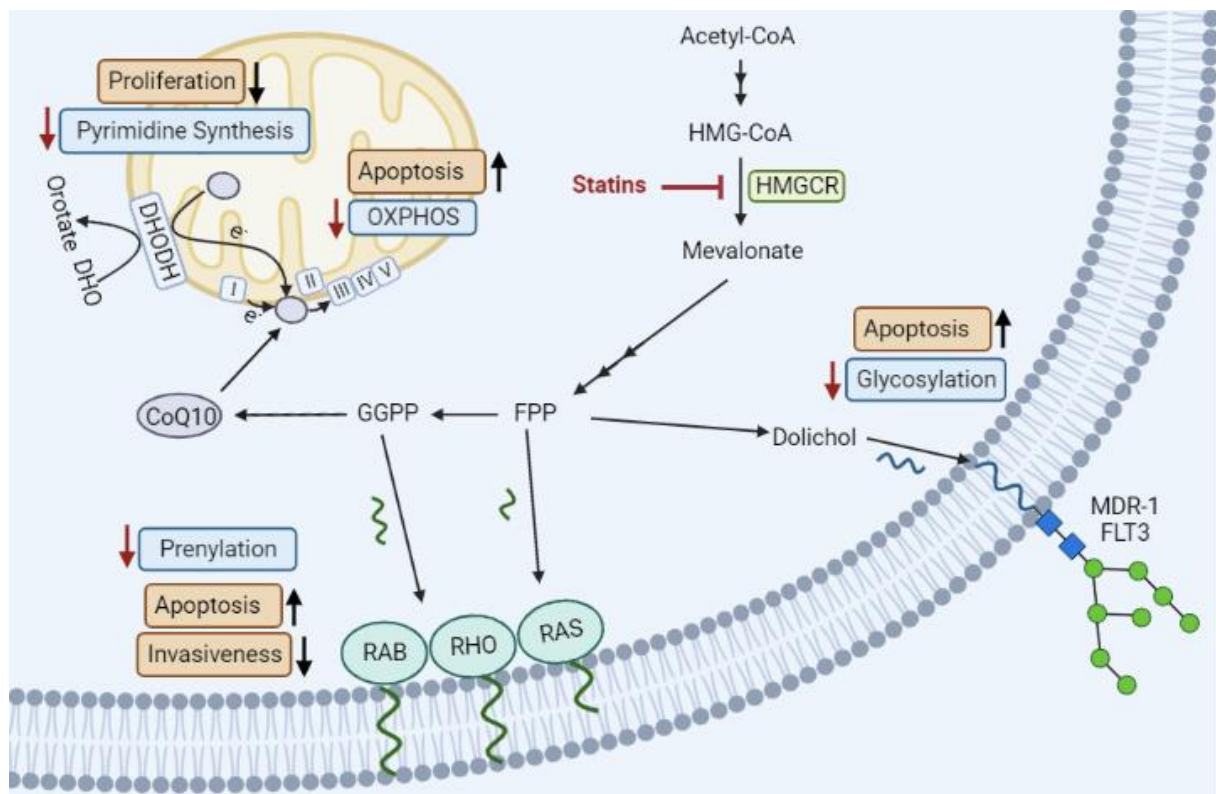


Figure 2. Proposed anti-cancer mechanisms of statins. Statins inhibit the mevalonate pathway by inhibition of HMGCR. This results in a decrease in the synthesis of isoprenoids, CoQ10 and dolichol, which affects prenylation, pyrimidine synthesis, OXPHOS and glycosylation. The following cancer hallmarks are targeted by statin treatment: sustaining proliferative signaling by decreased pyrimidine synthesis, deregulation of cellular energetics by inhibition of OXPHOS, invasion & metastasis by reduced prenylation of GTPases and resisting cell death by decreased glycosylation of MDR-1 and FLT3. Square boxes on the mitochondrion represent the complexes (I-V) of the electron transport chain (ETC). Blue boxes represent biological key processes that are affected upon statin treatment. Orange boxes represent effects of statins on relevant anti-cancer endpoints.

5.2 Renin angiotensin system inhibitors

The renin angiotensin system (RAS) regulates blood pressure homeostasis⁴⁵, which makes it a frequently used target to treat hypertension. The two most used RAS inhibitors are angiotensin converting enzyme inhibitors (ACEIs) and angiotensin receptor blockers (ARBs). ACEIs inhibit angiotensin converting enzyme (ACE) which reduces the conversion of biological inactive angiotensin I to biological active angiotensin II. ARBs are antagonists of the angiotensin II receptor type I (AT1R) and therefore evade binding of angiotensin II to this receptor. Components of the RAS system like AT1R and ACE are expressed on tumor cells and on several cells of the tumor microenvironment (TME)⁴⁶. Therefore, it is suggested that RAS signaling is involved in tumor progression and that RAS inhibitors could reverse these effects^{19,20,47,48}. Indeed, RAS inhibition showed to affect the cancer hallmarks activating invasion & metastasis, inducing angiogenesis and avoiding immune destruction. In addition, it was shown that RAS inhibitors increased the drug delivery of chemotherapeutics. The mechanisms that clarify these effects are discussed below and visualized in Figure 3.

RAS signaling influences invasion and metastasis via multiple mechanisms. One suggested mechanism is by supporting epithelial to mesenchymal transition (EMT), which is a known driver of invasiveness of cancer cells⁴⁹. In colorectal cancer, it was observed that induction of AT1R signaling using angiotensin II resulted in increased migration and increased expression of the EMT marker

ZEB1. Treatment with the ARB irbesartan reversed both migration and ZEB1 expression⁵⁰, showing that RAS inhibitors successfully can be used to target migration and EMT in colorectal cancer. Beside EMT, metastasis can be modulated by increasing invading capabilities. RAS signaling was suggested to increase invasiveness in breast cancer cells by inducing the expression of metalloproteinase (MMP)-2, MMP-9 and intercellular adhesion molecule 1 (ICAM-1)⁵¹. Especially MMP-2 and MMP-9 play a role in invasion and migration by enzymatically degrading extracellular matrix (ECM)⁵².

Angiogenesis is as well modulated by RAS signaling. In AT1R positive ovarian carcinoma cell lines, it was observed that angiotensin II resulted in increased vascular endothelial growth factor (VEGF) excretion and that this effect was successfully reversed by AT1R blockade with the ARB candesartan⁵³. When these cells were grown in a mice model, it was again observed that angiotensin II induced angiogenesis. In addition, increased invasiveness was observed upon angiotensin II exposure. Candesartan treatment reversed both angiogenesis and invasiveness in the mice model, which provided extra evidence that RAS signaling modulates angiogenesis and invasiveness.

Beside the effects of RAS inhibitors on cancer hallmarks, it was also found that RAS inhibitors affected drug delivery. Losartan showed to increase the delivery of chemotherapeutic drugs to cancer cells by decompression of the tumor vasculature in the TME and by reducing solid stress (i.e., accumulation of solid structural components in the TME⁵⁴)⁵⁵. Losartan reduced solid stress by decreasing collagen I production in cancer associated fibroblast (CAFs)⁵⁵. In a study of the same authors, it was shown that this collagen I production was a result of AT1R-mediated secretion of transforming growth factor β 1 (TGF- β 1) activators like thrombospondin-1 (TSP-1)⁵⁶, which shows that cross-talk between AT1-R and TGF- β 1 signaling is responsible for the effect on solid stress.

Finally, RAS inhibitors showed to affect the cancer hallmark avoiding immune destruction. Immune cells can either be pro-tumorigenic by suppressing the immune system or anti-tumorigenic by increasing the anti-tumor immune response. Examples of pro-tumorigenic immune cells include regulatory T-cells and M2 macrophages which both suppress the anti-tumor immune response. Anti-tumorigenic immune cells include pro-inflammatory M1 macrophages and cytotoxic T-cells. Nanoconjugated valsartan showed a synergistic effect with anti-PD1 and anti-CTLA4 checkpoint inhibitors⁵⁷. This effect was associated with increased cytotoxic T-cells to Treg ratio and an increased M1 macrophage to M2 macrophage ratio. These effects can be declared by the previously indicated effects of RAS inhibitors on solid stress, TGF- β and VEGF since these factors are known mediators of immune invasion⁴⁸.

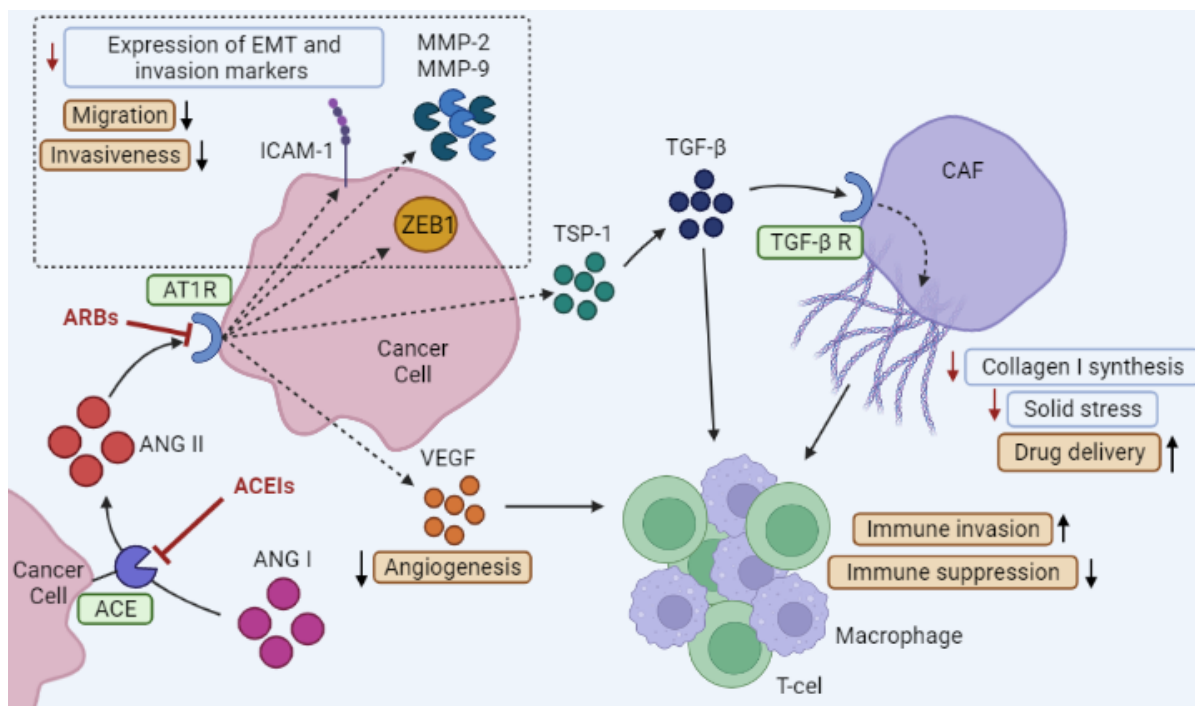


Figure 3. Anti-cancer mechanisms of RAS inhibitors. RAS inhibitors directly or indirectly antagonize AT1R resulting in decreased expression of EMT marker ZEB1, decreased expression of invasion markers MMP-2, MMP-9 and ICAM-1, decreased secretion of VEGF and decreased secretion of TSP-1. Reduced secretion of TSP-1 results in a reduction of available TGF- β 1 levels which decreases collagen I synthesis by cancer associated fibroblast (CAFs). Ras inhibitors affected the cancer hallmarks invasion & metastasis by decreased expression of EMT marker ZEB1 and decreased expression of MMP-2, MMP-9 and ICAM-1, angiogenesis by reduced expression of VEGF and avoiding immune destruction by decreasing VEGF and TGF- β levels and by decreased collagen I synthesis accompanied with reduced solid stress. In addition, RAS inhibitors showed to induce drug delivery by decreasing collagen I synthesis which reduced solid stress. Blue boxes represent biological key processes that are affected by RAS inhibitors. Orange boxes represent effects of RAS inhibitors on relevant anti-cancer endpoints. Green boxes represent important receptors and enzymes.

5.3 Selective betablockers

There is growing evidence that chronic stress, inflammation and accumulation of catecholamines stimulates cancer progression⁵⁸. In line with this, it was shown that accumulation of catecholamines and increased density of β -adrenergic receptors (β -AR) promoted carcinogenesis of breast, pancreas and ovary cancers¹⁸. β -ARs can be targeted with betablockers (BBs) and showed anti-tumor activity previously⁵⁸. BBs can be subdivided in non-selective and selective BBs. Non-selective BBs target both the β 1-AR and the β 2-AR while selective BBs solely target the β 1-AR. As reviewed elsewhere, anti-cancer effects of BBs are majorly established by targeting β 2-AR using non-selective BBs^{15,47,59,60}. However, the goal of this review was to declare anti-cancer effects of the most used drugs. Since the prescription of selective BBs is higher than non-selective BBs²⁴, the potential anti-cancer effect of selective BBs was further investigated.

Some studies suggest that the selective BB nebivolol induces apoptosis in cancer cells and affects the cancer hallmarks deregulation of cellular energetics and angiogenesis^{61,62}. The mechanisms that clarify these effects are discussed below and visualized in Figure 4.

Nebivolol was suggested to induce apoptosis by β 1-AR dependent targeting of the mitochondria. It was shown that β 1-AR inhibition using Nebivolol resulted in upregulation of ATPase inhibitory factor I (IF1), which inhibited ATP synthase (also known as complex V of the ETC)⁶¹. In addition, nebivolol prevented phosphorylation of complex I of the ETC. As a result, OXPHOS was impaired resulting in apoptosis.

The effect of nebivolol on angiogenesis was established by targeting of human umbilical vein endothelial cells (HUVEC). It is known that decreased proliferation of these cells results in decreased formation of new blood vessels⁶³. Nebivolol showed anti-angiogenic activity by decreasing proliferation of HUVECs⁶¹. Mechanistically, nebivolol prevented the phosphorylation of ERK, which subsequently reduced glycolysis and finally lead to cell cycle arrest.

Together, the anti-proliferative and anti-angiogenic effects of nebivolol resulted in decreased cancer growth in colon and breast cancer mice models⁶¹. Furthermore, nebivolol showed to reduce angiogenesis and proliferation *in vitro* and synergized with vincristine *in vivo* in neuroblastoma⁶².

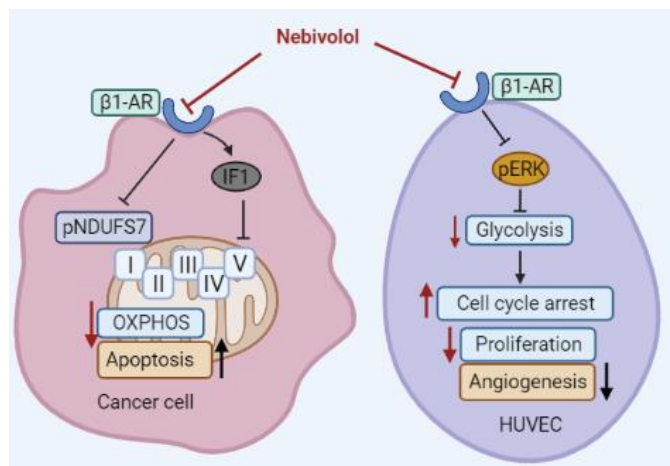


Figure 4. The proposed anti-cancer mechanism of nebivolol. Nebivolol targets the cancer hallmark deregulation of cellular energetics by β 1-AR-dependent inhibition of complex I and complex V of the ETC resulting in decreased OXPHOS and induction of apoptosis. Nebivolol also targets angiogenesis by β 1-AR dependent inhibition of the proliferation of human umbilical vein endothelial cells (HUVECs). Square boxes on the mitochondrion represent the complexes (I-V) of the ETC. Blue boxes represent biological key processes that are affected upon nebivolol treatment. Orange boxes represent effects of nebivolol on relevant anti-cancer endpoints.

5.4 Dihydropyridine calcium channel blockers

Calcium signaling is a complex signaling network that plays a key-role in multiple cellular processes⁶⁴. Disruption of calcium homeostasis has been shown to affect multiple functions including proliferation, gene expression, cell death, and protein phosphorylation and dephosphorylation⁶⁵. Interestingly, expression of calcium channels/pumps and calcium regulating proteins is altered in cancer⁶⁶, which makes targeting of calcium signaling a potential cancer drug target. Since dihydropyridine calcium channel blockers (CCBs) block L-type voltage gated calcium channels, these drugs might have an anti-cancer effect by modulating calcium signaling. Indeed, some studies suggest anti-cancer activity of CCBs by affecting the cancer hallmark sustaining proliferation^{67,68}. The proposed anti-cancer mechanisms of CCBs are discussed below and visualized in Figure 5A.

Disruption of calcium regulation by the dihydropyridine CCB Amlodipine showed anti-proliferative effects on uveal melanoma and epidermoid carcinoma cell lines^{67,68}. For both uveal melanoma and epidermoid carcinoma, it was observed that anti-proliferative effects were established by induction of cell-cycle arrest. This effect could be clarified by changed intracellular calcium signaling. Intracellular calcium will bind to calmodulin and calcineurin, which subsequently upregulate p21 expression and cyclin D1 synthesis^{23,69}. These both processes are involved in cell cycle progression and it is known that inhibition of calmodulin results in cell cycle arrest⁷⁰. Therefore, the proposed anti-cancer mechanism of amlodipine in uveal melanoma and epidermoid carcinoma is decreasing of calcium influx, which subsequently down regulates calcium signaling and results in cell cycle arrest.

Conflicting data were found for dihydropyridine CCBs. While the dihydropyridine CCB amlodipine showed anti-proliferative effects in uveal melanoma and epidermoid carcinoma, nifedipine, another

dihydropyridine CCB, showed to increase proliferation in breast cancer cells^{71,72}. Therefore, it is debatable whether all dihydropyridine CCBs have anti-cancer activity.

A clarification of these opposing results might be that the anti-cancer effect of dihydropyridine CCBs is dependent on the expressed calcium channel isoform. Anti-cancer activity of dihydropyridine CCBs was observed for uveal melanoma and epidermoid carcinoma which both arise from the skin. In melanoma, another form of skin cancer, it was observed that majorly the Cav1.3 calcium channel isoform was expressed⁷³. Interestingly, it was shown that silencing of Cav1.3 resulted in a decrease in proliferation in breast cancer cells⁷⁴. This study is in contrast with the study where targeting of calcium channels with nifedipine in the same form of cancer resulted in an increase in proliferation⁷¹. Therefore, it might be possible that the anti-cancer effect of dihydropyridine CCBs is dependent on expression of specific calcium channel isoforms e.g. Cav1.3.

5.5 Gabapentin

Gabapentin is a structural analogue of gamma-aminobutyric acid (GABA), which is an inhibitory neurotransmitter. The drug was originally developed as an anti-epileptic drug to treat certain types of seizures, but nowadays it is as well used for treatment of neuropathic pain including cancer-induced pain^{75,76}. Interestingly, two studies suggest that gabapentin also has anti-cancer activity by affecting the cancer hallmark sustaining proliferative signaling^{22,77}. The mechanism behind this proposed effect is discussed below and visualized in Figure 5B.

Likely to CCBs, gabapentin could modulate calcium signaling. Gabapentin affects calcium signaling as it is a ligand of $\alpha 2\delta 1$ and $\alpha 2\delta 2$ subunits of voltage gated calcium channels⁷⁸. Both studies that discovered anti-proliferative activity in cancer, suggested that this effect was established by inhibition of the $\alpha 2\delta 2$ subunit of calcium channels^{22,77}. In prostate cancer cells, it was shown that up- and downregulation of the $\alpha 2\delta 2$ subunit resulted in in- or decreased cell proliferation by in- or reducing cell cycle arrest²². In this study, it was shown that gabapentin reduced cell proliferation both *in vitro* and *in vivo*. The anti-proliferative effect was suggested to be caused by targeting of the calcineurin/nuclear factor of the activated T cell (NFAT) pathway. It was shown that $\alpha 2\delta 2$ overexpressing cells had an increased calcium influx. The subsequently increased cytosolic calcium levels can activate calcineurin, which in turn increases NFAT activity resulting in increased proliferation. Due to increased calcium influx, $\alpha 2\delta 2$ overexpressing cells indeed showed increased NFAT activity. In addition, calcineurin inhibitors showed to decrease proliferation in these $\alpha 2\delta 2$ overexpressing cells. Together these data show that $\alpha 2\delta 2$ regulates proliferation using the calcineurin/NFAT pathway. In melanoma, gabapentin also reduced proliferation *in vitro* and *in vivo*⁷⁷. In this study it was shown that gabapentin reduced calcium influx, which also links the anti-proliferative effect of gabapentin to calcium signaling.

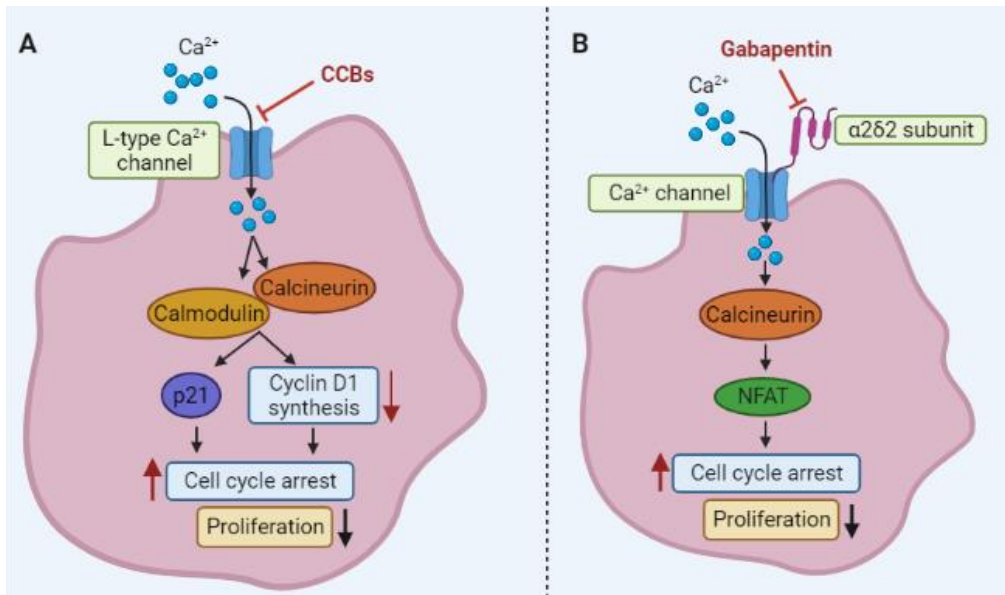


Figure 5. The proposed anti-cancer effects of dihydropyridine CCBs and gabapentin. A) CCBs affect the cancer hallmark sustaining proliferative signaling by reducing the calcium influx resulting in decreased activation and downstream synthesis of cyclin D1 which eventually induces cell cycle arrest. B) Gabapentin affects the cancer hallmark sustaining proliferative signaling by inducing cell cycle arrest. Gabapentin reduces calcium influx by inhibition of the $\alpha 2\delta 2$ subunit of calcium channels which results in decreased binding to calcineurin and downstream activation of NFAT which eventually induces cell cycle arrest. Blue boxes represent biological key processes that are affected by the drugs. Orange boxes represent effects of the drugs on relevant anti-cancer endpoints. Green boxes represent important channels or subunits of channels.

5.6 Metformin

Metformin is a biguanide that is typically prescribed as a first line therapy for type II diabetes⁷⁹. Metformin decreases circulating glucose levels by inhibition of hepatic gluconeogenesis⁸⁰. In addition, metformin is suggested to induce insulin sensitivity by increased expression of the insulin receptor⁸¹. Beside this, some studies suggest that metformin also has anti-cancer activity. Metformin is suggested to have anti-cancer activity by targeting the cancer hallmarks invasion & metastasis, sustaining proliferative signaling and deregulation of cellular energetics. The mechanisms that declare these anti-cancer effects are discussed below and visualized in Figure 6.

Direct and indirect anti-cancer mechanisms of metformin are described¹². Direct anti-cancer mechanisms are direct effects of metformin on cancer cells. Indirect effects are effects that are established via the insulin lowering effect of metformin. The direct anti-cancer mechanisms will be discussed first.

Firstly Metformin affects the cancer hallmarks sustaining proliferative signaling and deregulating cellular energetics. Metformin inhibits complex I of the ETC and subsequently reduces OXPHOS, resulting in decreased ATP production and relative high AMP levels. These elevated AMP levels induce AMP-activated protein kinase (AMPK), which is an important mediator of (cancer) metabolism⁸². Beside AMPK activation via AMP, it was shown that low doses of metformin activates AMPK by binding to the lysosomal protein PEN2⁸³. Metformin-induced AMPK activation showed to inhibit mTOR which resulted in cell cycle arrest in multiple myeloma cells⁸⁴. This caused inhibition of proliferation, but did not initiate apoptosis. This is in line with other studies that as well showed that metformin alone does not induce apoptosis in tumor cell lines, but does decrease proliferation by inducing cell cycle arrest⁸⁵⁻⁸⁷.

In another study, metformin did show an apoptotic effect. It was shown that metformin induced apoptosis in p53 deficient colon cancer cell lines, but not in cells with functioning p53⁸⁸. In a later study, it was found that the apoptotic effect in p53 deficient colon cancer cells was established by

the inhibition of complex I and subsequent impaired OXPHOS⁸⁹. However, conflicting data were found in another type of cancer. In breast cancer, cells with functioning p53 showed metformin-induced apoptosis while mutated p53 cells were resistant to metformin.⁹⁰ Therefore, it might be possible that the apoptotic effect on p53 deficient cells is specific for colon cancer cells.

Beside the effect of metformin on proliferation and apoptosis, it has also been shown that metformin reduces invasion and migration in some cancers⁹⁰⁻⁹². The effect of metformin on invasion in multiple myeloma was dependent on AMPK activation and subsequent activation of p53⁹³. In addition, this study showed that metformin effectively reduced metastasis of multiple myeloma cells in a mice model. In hepatocellular carcinoma cell lines, metformin reduced both migration and invasion in a p53 dependent manner, which supports the anti-metastatic potential of metformin⁹¹. Finally, it was shown in cholangiocarcinoma that metformin reduced invasion and synergized the anti-migratory effect of cisplatin⁹². This effect was again accompanied with increased activation of p53 which provided additional evidence that metformin affects migration and invasion by regulating p53 activation.

Indirect anti-cancer effects of metformin are related to insulin-modulating activity of metformin. Metformin decreases insulin-like growth factor (IGF) levels in hyperinsulinemic patients, by decreasing the IGF binding protein 1 (IGFBP-1) levels⁹⁴. The indirectly lowering effect on circulating insulin and insulin growth factor is proposed as an anti-neoplastic effect of metformin⁹⁵. It was observed in a lung cancer mice model that metformin indeed decreased circulating IGF levels with subsequent decreased phosphorylation of the IGF receptor (IGFR)⁹⁶. This affected downstream AKT signaling which resulted in inhibition of mTOR. This inhibition of mTOR was not accompanied with increased activation of AMPK, which confirmed an alternative mechanism of metformin to inhibit mTOR. Again it was shown that inhibition of mTOR resulted in decreased cell proliferation, but did not result in apoptosis.

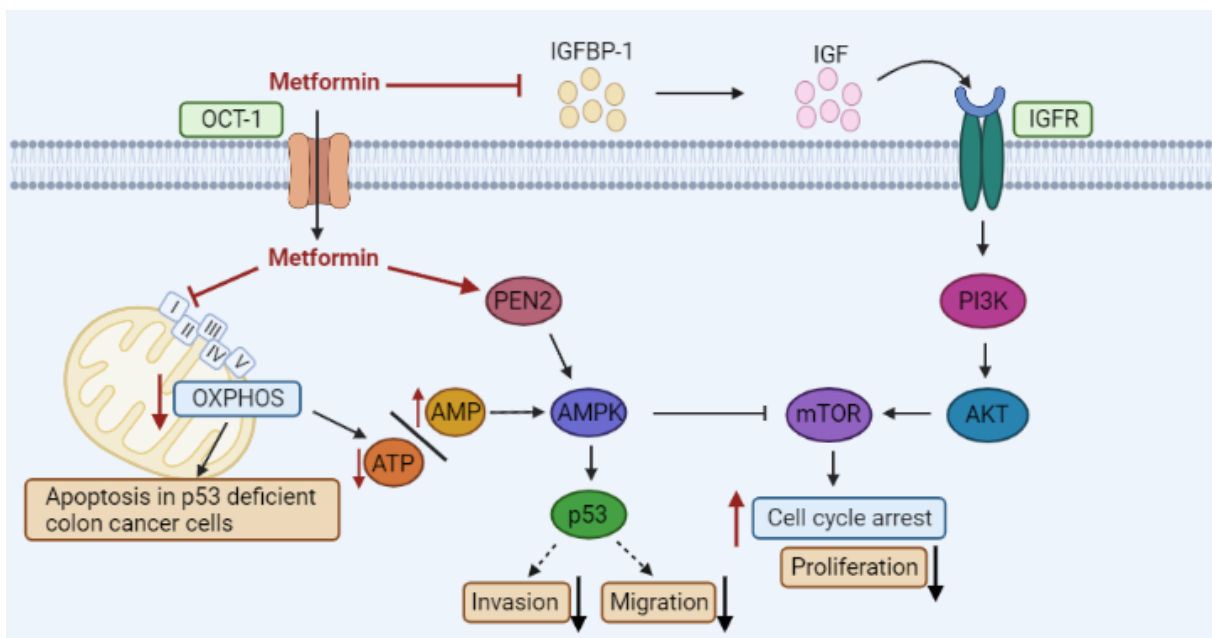


Figure 6. Proposed anti-cancer mechanisms of Metformin. Metformin directly targets the cancer cell by inhibition of complex I of the ETC or by binding to PEN2 and subsequent downstream effects. In addition, metformin targets cancer cells indirectly by decreasing the circulating IGF levels. The affected cancer hallmarks are deregulating cellular energetics by inhibition of complex I of the ETC and subsequent decreased OXPHOS, invasion & metastasis by activation of p53 and sustaining proliferative signaling by inducing cell cycle arrest. Blue boxes represent biological key processes that are affected by metformin. Orange boxes represent effects of metformin on relevant anti-cancer endpoints. Green boxes represent important receptors or transporters.

5.7 Protein pump inhibitors

Proton pump inhibitors (PPIs) reduce acidification of the stomach by inhibiting H⁺/K⁺ ATPase of parietal cells⁹⁷. Because of its anti-acid effect, PPIs are prescribed for treatment of gastroesophageal reflux disease (GERD) and drug-induced peptic ulcers. Beside its effect on H⁺/K⁺ ATPase, PPIs can inhibit the vacuolar-type ATPase (V-ATPase)⁹⁸ and fatty acid synthase (FASN)⁹⁹. Some studies suggest anti-cancer activity by PPI-induced inhibition of these two targets^{100–104}. PPIs were suggested to affect the cancer hallmarks migration & invasion and genome instability. In addition, it was shown that PPIs induced the delivery of weakly basic drugs^{105,106}. The mechanisms that clarify the suggested anti-cancer effects of PPIs are discussed below and visualized in Figure 7.

Firstly, PPIs induce apoptosis in cancer cells and increase drug delivery by inhibition of V-ATPase and subsequent pH modulation. V-ATPase is expressed on cancer cells and is involved in regulation of the intra- and extracellular pH¹⁰⁷. The extracellular pH is acidic while the intracellular pH is neutral to alkaline¹⁰⁸. PPIs successfully induced apoptosis in melanoma *in vitro* and *in vivo* by inhibition of V-ATPase, which was related with elevated extracellular pH levels and decreased intracellular pH levels¹⁰⁰. This pH modulation can also affect the delivery of chemotherapeutic drugs. It is known that the activity of weakly basic chemotherapeutic drugs is decreased by the low pH in the TME¹⁰⁹. PPIs induce the extracellular pH by targeting V-ATPase on tumor cells and therefore could reverse this effect¹⁰⁰. In line with this, it was shown that pre-treatment with omeprazole and esomeprazole improved the drug response of the weakly basic chemotherapeutics cisplatin, 5-fluorouracil and vinblastine in multidrug resistant cells¹⁰⁶. In addition, it was shown that omeprazole and lansoprazole induced the drug delivery of the weakly basic anticancer drug doxorubicin in 3D breast cancer spheroids¹⁰⁵.

Secondly, several studies showed that PPIs target the cancer hallmark migration and invasion in gastric cancer, breast cancer and glioma^{101,102,104}. The effect of PPIs in these studies was accompanied with decreased expression of EMT markers like vimentin, n-cadherin and snail. However, these studies did not provide a direct link between the mechanism of action of PPIs and the observed effects on EMT markers. Potentially, the effects on migration and invasion were established by PPI-induced inhibition of FASN. Knockdown of FASN showed decreased migration in gastric cancer cells accompanied with decreased expression of the EMT marker vimentin¹¹⁰, which was as well affected in PPI-treated cells. Therefore, it is hypothesized that the effect of PPIs on migration, invasion and EMT is established by inhibition of FASN.

Finally, PPI-induced inhibition of FASN is suggested to affect the hallmark genomic instability. It is mechanistically known that FASN can regulate nonhomologous end-joining (NHEJ) pathways¹¹¹. These pathways are involved in DNA repair and are therefore useful to protect the cancer cells from chemotherapeutics and ionizing radiation¹¹². FASN regulates NHEJ by suppression of NF-κβ and induction of specificity protein 1 (SP1) expression. Both suppression of NF-κβ and increased expression of SP1 result in induction of the poly(ADP-ribose) polymerase 1 (PARP-1) promoter. Finally, PARP-1 recruits Ku proteins to induce chromatin and DNA repair resulting in increased DNA repair by NHEJ¹¹¹. Palmitate, the fat molecule which is produced by FASN, modulated the activation of PARP1 and SP1. This showed that the effect on NHEJ is dependent on the production of palmitate by FASN. Inhibition of FASN using PPIs showed to induce apoptosis in breast cancer cells¹⁰³. In this study, it was observed that FASN inhibition was accompanied with reduced PARP1 expression and decreased DNA repair, which is in line with the effect of FASN on NHEJ. In addition, it was found that PPIs synergized with doxorubicin. In a follow up study, PPIs were tested for their potential to synergize with chemotherapeutics in a phase II clinical trial with patients with triple negative breast cancer (TNBC)²¹. This study population was selected since FASN is overexpressed in 70% of the TNBC

cases. In the clinical trial, it was observed that addition of high dose omeprazole improved the pathologic complete response of neoadjuvant chemotherapy²¹.

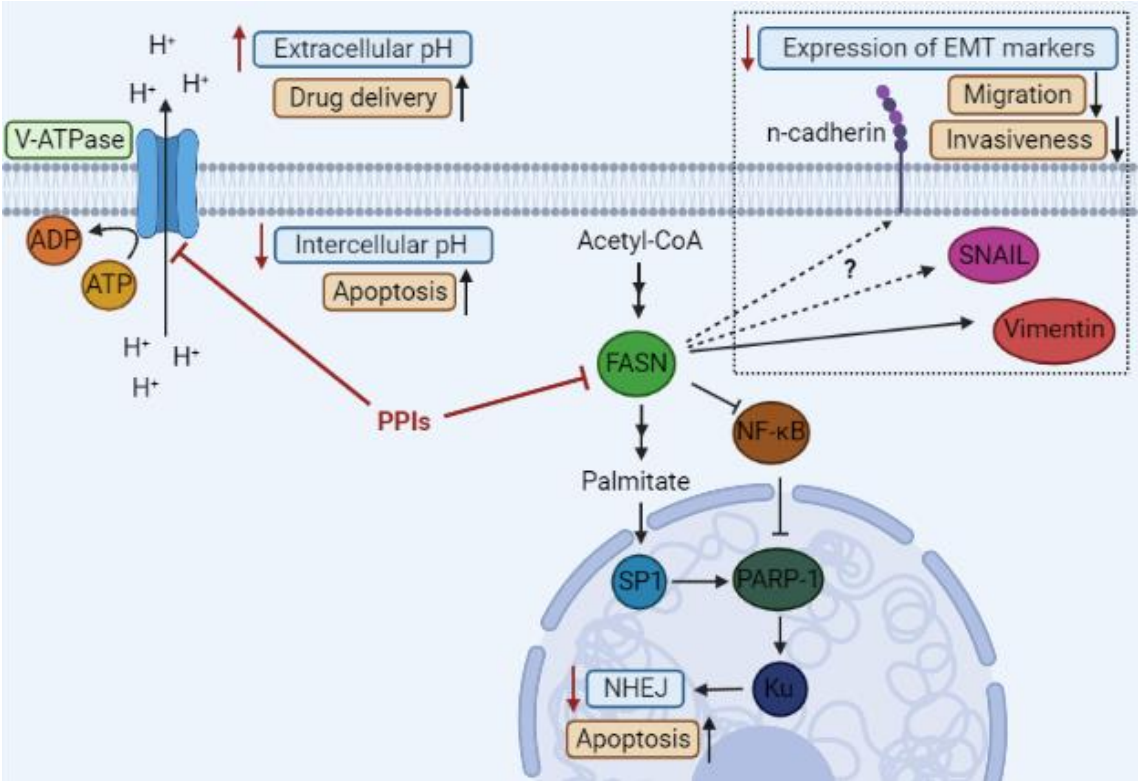


Figure 7. Proposed anti-cancer mechanisms of PPIs. PPIs target V-ATPase which affects the inter- and extracellular pH homeostasis and results in increased delivery of drugs and/or cancer cell apoptosis. In addition, PPIs target FASN which is involved in NHEJ and is hypothesized to be involved in the expression of EMT markers. The affected cancer hallmarks are invasion & metastasis by inhibition of the expression of EMT markers and genomic instability by decreasing NHEJ. Blue boxes represent biological key processes that are affected by metformin. Orange boxes represent effects of metformin on relevant anti-cancer endpoints. Green boxes represent important transporters.

6 Discussion

Drug repurposing is an attractive approach to develop novel cancer treatments since it is less time consuming, has lower costs and has higher chances to get marketing authorization compared to development of new drugs^{13,14}. Potential candidates for drug repurposing are extensively used drugs since their high utilization provides a lot of safety data and insight in potential side effects. Therefore, mechanistical evidence that supports anti-cancer activity of (the drug classes of) eight of the ten most used drugs worldwide was investigated in this literature review. Data from experimental *in vitro* and *in vivo* studies suggested anti-cancer activity for all of the investigated drugs. The anti-cancer effects were established by targeting of several cancer hallmarks including: sustaining proliferation, induction of invasion and metastasis, avoiding immune destruction, induction of angiogenesis, deregulation of cellular energetics, genome instability and resisting cell death. Mechanistical evidence that clarified anti-cancer effects were found for all drugs. These mechanisms support suggested anti-cancer activity of the investigated drugs.

Targeting of cancer with single compounds is challenging since hallmarks of cancer are clearly not regulated by single signaling pathway^{4,113}. Drug combinations that target multiple hallmarks can therefore be superior compared to monotreatment since they may target supporting pathways that are not targeted by single treatments, ultimately resulting in reversion of drug resistance and increased effect of the drug^{4,114}. Therefore, combining of drugs is an effective approach to develop novel cancer treatments. In addition, combining of drugs is an attractive approach for drug repurposing since drugs are more likely to get authorized as repurposed drugs if they are combined with other drugs¹¹⁵. Since combining of drugs is an attractive and effective approach for cancer drug repurposing, the drug classes statins⁴³, RAS inhibitors⁵⁵, selective BBs⁶², biguanides⁹² and PPIs¹⁰³ which showed combinational effects with conventional treatments in this literature review, were considered as potential candidates for drug repurposing.

Repositioning of RAS inhibitors, selective BBs and CCBs could be challenging as these drugs decrease systemic blood pressure. Therefore, there is a risk of hypotension when these drugs are administered to patients with normotension. Because of this, clinical trials to test the efficacy of these drugs might be restricted to patients with existing hypertension. A novel approach to avoid unwanted hypotension is targeted delivery of the drugs to the TME. For ARBs, several approaches were already tested. Encapsulation of losartan in liposomes effectively improved the efficacy of liposomal paclitaxel without affecting the blood pressure¹¹⁶. In addition, the ARB valsartan conjugated to a pH sensitive polymer showed effective delivery of valsartan to the TME and synergism with immune-checkpoint inhibitors in a breast cancer mice model⁵⁷. These data show that targeted delivery could avoid side effects of blood pressure lowering drugs. Furthermore, it was shown that the investigated drugs might have the potential to be used as active agents for novel nanomedicines.

Interestingly, the compounds pitavastatin and amlodipine were identified as potential anti-cancer drugs by high-throughput screening^{43,67}. The drugs showed effects on uveal melanoma and glioblastoma, which are both orphan diseases with limited treatment options¹¹⁷⁻¹¹⁹. Drug repurposing holds a great promise for these orphan diseases, since the low incidence of these diseases makes it relatively expensive to develop novel drugs for such a small population¹²⁰. Especially the results of pitavastatin on glioblastoma were promising since this drug showed anti-cancer effects both *in vitro* and *in vivo*. Therefore, these two studies show that high-throughput screening could be an effective approach to identify compounds for drug repurposing in orphan diseases.

Overall, anti-cancer mechanisms were identified for all the investigated drugs. These mechanisms support suggested anti-cancer activity of the investigated drugs and therefore it was concluded that

the drug gabapentin and the drug classes statins, RAS inhibitors, selective betablockers, dihydropyridine CCBs, biguanides and PPIs have the potential to be used for cancer drug repurposing. Since the drug classes statins, RAS inhibitors, selective BBs, biguanides and PPIs showed combinational effects with conventional treatments, these drug classes were especially considered as potential agents for cancer drug repurposing.

7 Bibliography

1. Sung H, Ferlay J, Siegel RL, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin*. 2021;71(3):209-249. doi:10.3322/caac.21660
2. Kunnumakkara AB, Bordoloi D, Sailo BL, et al. Cancer drug development: The missing links. *Exp Biol Med*. 2019;244(8):663-689. doi:10.1177/1535370219839163
3. Moreno L, Pearson AD. How can attrition rates be reduced in cancer drug discovery? *Expert Opin Drug Discov*. 2013;8(4):363-368. doi:10.1517/17460441.2013.768984
4. Hanahan D, Weinberg RA. Hallmarks of Cancer: The Next Generation. *Cell*. 2011;144(5):646-674. doi:10.1016/j.cell.2011.02.013
5. Bergers G, Hanahan D. Modes of resistance to anti-angiogenic therapy. *Nat Rev Cancer*. 2008;8(8):592-603. doi:10.1038/nrc2442
6. Ebos JML, Lee CR, Kerbel RS. Tumor and Host-Mediated Pathways of Resistance and Disease Progression in Response to Antiangiogenic Therapy: Fig. 1. *Clin Cancer Res*. 2009;15(16):5020-5025. doi:10.1158/1078-0432.CCR-09-0095
7. Davis C, Naci H, Gurpinar E, Poplavska E, Pinto A, Aggarwal A. Availability of evidence of benefits on overall survival and quality of life of cancer drugs approved by European Medicines Agency: retrospective cohort study of drug approvals 2009-13. *BMJ*. 2017;359:j4530. doi:10.1136/bmj.j4530
8. Wieseler B, McGauran N, Kaiser T. New drugs: where did we go wrong and what can we do better? *BMJ*. 2019;366:l4340. doi:10.1136/bmj.l4340
9. Pease AM, Krumholz HM, Downing NS, Aminawung JA, Shah ND, Ross JS. Postapproval studies of drugs initially approved by the FDA on the basis of limited evidence: systematic review. *BMJ*. 2017;357:j1680. doi:10.1136/bmj.j1680
10. Gyawali B, Hey SP, Kesselheim AS. Assessment of the Clinical Benefit of Cancer Drugs Receiving Accelerated Approval. *JAMA Intern Med*. 2019;179(7):906-913. doi:10.1001/jamainternmed.2019.0462
11. Verbaanderd C, Meheus L, Huys I, Pantziarka P. Repurposing Drugs in Oncology: Next Steps. *Trends in Cancer*. 2017;3(8):543-546. doi:10.1016/j.trecan.2017.06.007
12. Kirtonia A, Gala K, Fernandes SG, et al. Repurposing of drugs: An attractive pharmacological strategy for cancer therapeutics. *Semin Cancer Biol*. 2021;68:258-278. doi:10.1016/j.semcancer.2020.04.006
13. Shah K, Gupta S, Ghosh J, Baipai J, Maheshwari A. Acute non-ST elevation myocardial infarction following paclitaxel administration for ovarian carcinoma: A case report and review of literature. *J Cancer Res Ther*. 2012;8(3):442-444. doi:10.4103/0973-1482.103530.
14. Chong CR, Sullivan DJ. New uses for old drugs. *Nature*. 2007;448(7154):645-646. doi:10.1038/448645a
15. Gelosa P, Castiglioni L, Camera M, Sironi L. Repurposing of drugs approved for cardiovascular diseases: Opportunity or mirage? *Biochem Pharmacol*. 2020;177:113895. doi:10.1016/j.bcp.2020.113895
16. Tatokoro M, Fujii Y, Kawakami S, et al. Phase-II trial of combination treatment of interferon- α , cimetidine, cyclooxygenase-2 inhibitor and renin-angiotensin-system inhibitor (I-CCA therapy)

- for advanced renal cell carcinoma. *Cancer Sci.* 2011;102(1):137-143. doi:10.1111/j.1349-7006.2010.01756.x
17. O'Rawe M, Wickremesekera AC, Pandey R, et al. Treatment of glioblastoma with re-purposed renin-angiotensin system modulators: Results of a phase I clinical trial. *J Clin Neurosci.* 2022;95:48-54. doi:10.1016/j.jocn.2021.11.023
 18. Peixoto R, Pereira M de L, Oliveira M. Beta-Blockers and Cancer: Where Are We? *Pharmaceuticals.* 2020;13(6):105. doi:10.3390/ph13060105
 19. Ishida J, Konishi M, Ebner N, Springer J. Repurposing of approved cardiovascular drugs. *J Transl Med.* 2016;14(1):269. doi:10.1186/s12967-016-1031-5
 20. Perini M V., Dmello RS, Nero TL, Chand AL. Evaluating the benefits of renin-angiotensin system inhibitors as cancer treatments. *Pharmacol Ther.* 2020;211:107527. doi:10.1016/j.pharmthera.2020.107527
 21. Sardesai SD, Thomas A, Gallagher C, et al. Inhibiting Fatty Acid Synthase with Omeprazole to Improve Efficacy of Neoadjuvant Chemotherapy in Patients with Operable TNBC. *Clin Cancer Res.* 2021;27(21):5810-5817. doi:10.1158/1078-0432.CCR-21-0493
 22. Warnier M, Roudbaraki M, Derouiche S, et al. CACNA2D2 promotes tumorigenesis by stimulating cell proliferation and angiogenesis. *Oncogene.* 2015;34(42):5383-5394. doi:10.1038/onc.2014.467
 23. Yoshida J, Ishibashi T, Nishio M. G1 cell cycle arrest by amlodipine, a dihydropyridine Ca²⁺ channel blocker, in human epidermoid carcinoma A431 cells. *Biochem Pharmacol.* 2007;73(7):943-953. doi:10.1016/j.bcp.2006.12.011
 24. Fuentes A, Pineda M, Venkata K. Comprehension of Top 200 Prescribed Drugs in the US as a Resource for Pharmacy Teaching, Training and Practice. *Pharmacy.* 2018;6(2):43. doi:10.3390/pharmacy6020043
 25. Sirtori CR. The pharmacology of statins. *Pharmacol Res.* 2014;88:3-11. doi:10.1016/j.phrs.2014.03.002
 26. Wong WW-L, Dimitroulakos J, Minden M, Penn L. HMG-CoA reductase inhibitors and the malignant cell: the statin family of drugs as triggers of tumor-specific apoptosis. *Leukemia.* 2002;16(4):508-519. doi:10.1038/sj.leu.2402476
 27. McGregor GH, Campbell AD, Fey SK, et al. Targeting the Metabolic Response to Statin-Mediated Oxidative Stress Produces a Synergistic Antitumor Response. *Cancer Res.* 2020;80(2):175-188. doi:10.1158/0008-5472.CAN-19-0644
 28. Kaymak I, Maier CR, Schmitz W, et al. Mevalonate Pathway Provides Ubiquinone to Maintain Pyrimidine Synthesis and Survival in p53-Deficient Cancer Cells Exposed to Metabolic Stress. *Cancer Res.* 2020;80(2):189-203. doi:10.1158/0008-5472.CAN-19-0650
 29. Zaal EA, de Grooth H-J, Oudaert I, et al. Targeting coenzyme Q10 synthesis overcomes bortezomib resistance in multiple myeloma. *Mol Omi.* 2021. doi:10.1039/D1MO00106J
 30. Palsuledesai CC, Distefano MD. Protein Prenylation: Enzymes, Therapeutics, and Biotechnology Applications. *ACS Chem Biol.* 2015;10(1):51-62. doi:10.1021/cb500791f
 31. Berndt N, Hamilton AD, Sebti SM. Targeting protein prenylation for cancer therapy. *Nat Rev Cancer.* 2011;11(11):775-791. doi:10.1038/nrc3151
 32. Likus W, Siemianowicz K, Bieńk K, et al. Could drugs inhibiting the mevalonate pathway also

- target cancer stem cells? *Drug Resist Updat.* 2016;25:13-25. doi:10.1016/j.drug.2016.02.001
33. Wang M, Casey PJ. Protein prenylation: unique fats make their mark on biology. *Nat Rev Mol Cell Biol.* 2016;17(2):110-122. doi:10.1038/nrm.2015.11
 34. Pandya A, Mullen PJ, Kalkat M, et al. Immediate Utility of Two Approved Agents to Target Both the Metabolic Mevalonate Pathway and Its Restorative Feedback Loop. *Cancer Res.* 2014;74(17):4772-4782. doi:10.1158/0008-5472.CAN-14-0130
 35. Jiang Z, Zheng X, Lytle RA, Higashikubo R, Rich KM. Lovastatin-induced up-regulation of the BH3-only protein, Bim, and cell death in glioblastoma cells. *J Neurochem.* 2004;89(1):168-178. doi:10.1111/j.1471-4159.2004.02319.x
 36. Shellman YG, Ribble D, Miller L, et al. Lovastatin-induced apoptosis in human melanoma cell lines. *Melanoma Res.* 2005;15(2):83-89. doi:10.1097/00008390-200504000-00001
 37. Xia Z, Tan M, Wei-Lynn Wong W, Dimitroulakos J, Minden M, Penn L. Blocking protein geranylgeranylation is essential for lovastatin-induced apoptosis of human acute myeloid leukemia cells. *Leukemia.* 2001;15(9):1398-1407. doi:10.1038/sj.leu.2402196
 38. Agarwal B, Halmos B, Feoktistov AS, et al. Mechanism of lovastatin-induced apoptosis in intestinal epithelial cells. *Carcinogenesis.* 2002;23(3):521-528. doi:10.1093/carcin/23.3.521
 39. Denoyelle C, Vasse M, Körner M, et al. Cerivastatin, an inhibitor of HMG-CoA reductase, inhibits the signaling pathways involved in the invasiveness and metastatic properties of highly invasive breast cancer cell lines: an in vitro study. *Carcinogenesis.* 2001;22(8):1139-1148. doi:10.1093/carcin/22.8.1139
 40. Kang S, Kim E-S, Moon A. Simvastatin and lovastatin inhibit breast cell invasion induced by H-Ras. *Oncol Rep.* 2009;21(5):1317-1322. doi:10.3892/or_00000357
 41. Burda P, Aebi M. The dolichol pathway of N-linked glycosylation. *Biochim Biophys Acta - Gen Subj.* 1999;1426(2):239-257. doi:10.1016/S0304-4165(98)00127-5
 42. Forbes K, Shah VK, Siddals K, Gibson JM, Aplin JD, Westwood M. Statins inhibit insulin-like growth factor action in first trimester placenta by altering insulin-like growth factor 1 receptor glycosylation. *Mol Hum Reprod.* 2015;21(1):105-114. doi:10.1093/molehr/gau093
 43. Jiang P, Mukthavavam R, Chao Y, et al. Novel anti-glioblastoma agents and therapeutic combinations identified from a collection of FDA approved drugs. *J Transl Med.* 2014;12(1):13. doi:10.1186/1479-5876-12-13
 44. Williams AB, Li L, Nguyen B, Brown P, Levis M, Small D. Fluvastatin inhibits FLT3 glycosylation in human and murine cells and prolongs survival of mice with FLT3/ITD leukemia. *Blood.* 2012;120(15):3069-3079. doi:10.1182/blood-2012-01-403493
 45. Patel S, Rauf A, Khan H, Abu-Izneid T. Renin-angiotensin-aldosterone (RAAS): The ubiquitous system for homeostasis and pathologies. *Biomed Pharmacother.* 2017;94:317-325. doi:10.1016/j.biopha.2017.07.091
 46. Deshayes F, Nahmias C. Angiotensin receptors: a new role in cancer? *Trends Endocrinol Metab.* 2005;16(7):293-299. doi:10.1016/j.tem.2005.07.009
 47. Regulska K, Regulski M, Karolak B, Murias M, Stanisz B. Can cardiovascular drugs support cancer treatment? The rationale for drug repurposing. *Drug Discov Today.* 2019;24(4):1059-1065. doi:10.1016/j.drudis.2019.03.010
 48. Pinter M, Jain RK. Targeting the renin-angiotensin system to improve cancer treatment:

- Implications for immunotherapy. *Sci Transl Med*. 2017;9(410). doi:10.1126/scitranslmed.aan5616
49. Mimeault M, Batra SK. Interplay of distinct growth factors during epithelial mesenchymal transition of cancer progenitor cells and molecular targeting as novel cancer therapies. *Ann Oncol Off J Eur Soc Med Oncol*. 2007;18(10):1605-1619. doi:10.1093/annonc/mdm070
 50. Nguyen L, Ager EI, Neo J, Christophi C. Regulation of colorectal cancer cell epithelial to mesenchymal transition by the renin angiotensin system. *J Gastroenterol Hepatol*. 2016;31(10):1773-1782. doi:10.1111/jgh.13307
 51. Rodrigues-Ferreira S, Abdelkarim M, Dillenburger-Pilla P, et al. Angiotensin II facilitates breast cancer cell migration and metastasis. *PLoS One*. 2012;7(4):e35667. doi:10.1371/journal.pone.0035667
 52. Zitka O, Kukacka J, Krizkov S, et al. Matrix Metalloproteinases. *Curr Med Chem*. 2010;17(31):3751-3768. doi:10.2174/092986710793213724
 53. Suganuma T, Ino K, Shibata K, et al. Functional Expression of the Angiotensin II Type1 Receptor in Human Ovarian Carcinoma Cells and Its Blockade Therapy Resulting in Suppression of Tumor Invasion, Angiogenesis, and Peritoneal Dissemination. *Clin Cancer Res*. 2005;11(7):2686-2694. doi:10.1158/1078-0432.CCR-04-1946
 54. Levayer R. Solid stress, competition for space and cancer: The opposing roles of mechanical cell competition in tumour initiation and growth. *Semin Cancer Biol*. 2020;63:69-80. doi:10.1016/j.semcancer.2019.05.004
 55. Chauhan VP, Martin JD, Liu H, et al. Angiotensin inhibition enhances drug delivery and potentiates chemotherapy by decompressing tumour blood vessels. *Nat Commun*. 2013;4(1):2516. doi:10.1038/ncomms3516
 56. Diop-Frimpong B, Chauhan VP, Krane S, Boucher Y, Jain RK. Losartan inhibits collagen I synthesis and improves the distribution and efficacy of nanotherapeutics in tumors. *Proc Natl Acad Sci*. 2011;108(7):2909-2914. doi:10.1073/pnas.1018892108
 57. Chauhan VP, Chen IX, Tong R, et al. Reprogramming the microenvironment with tumor-selective angiotensin blockers enhances cancer immunotherapy. *Proc Natl Acad Sci*. 2019;116(22):10674-10680. doi:10.1073/pnas.1819889116
 58. Tang J, Li Z, Lu L, Cho CH. β -Adrenergic system, a backstage manipulator regulating tumour progression and drug target in cancer therapy. *Semin Cancer Biol*. 2013;23(6):533-542. doi:10.1016/j.semcancer.2013.08.009
 59. Zhang X, Zhang Y, He Z, et al. Chronic stress promotes gastric cancer progression and metastasis: an essential role for ADRB2. *Cell Death Dis*. 2019;10(11):788. doi:10.1038/s41419-019-2030-2
 60. Baker FL, Bigley AB, Agha NH, et al. Systemic β -Adrenergic Receptor Activation Augments the ex vivo Expansion and Anti-Tumor Activity of V γ 9V δ 2 T-Cells. *Front Immunol*. 2019;10:3082. doi:10.3389/fimmu.2019.03082
 61. Nuevo-Tapioles C, Santacatterina F, Stamatakis K, et al. Coordinate β -adrenergic inhibition of mitochondrial activity and angiogenesis arrest tumor growth. *Nat Commun*. 2020;11(1):3606. doi:10.1038/s41467-020-17384-1
 62. Pasquier E, Street J, Pouchy C, et al. β -blockers increase response to chemotherapy via direct antitumour and anti-angiogenic mechanisms in neuroblastoma. *Br J Cancer*.

- 2013;108(12):2485-2494. doi:10.1038/bjc.2013.205
63. Lamy S, Lachambre M-P, Lord-Dufour S, Béliveau R. Propranolol suppresses angiogenesis in vitro: inhibition of proliferation, migration, and differentiation of endothelial cells. *Vascul Pharmacol.* 53(5-6):200-208. doi:10.1016/j.vph.2010.08.002
 64. Patergnani S, Danese A, Bouhamida E, et al. Various Aspects of Calcium Signaling in the Regulation of Apoptosis, Autophagy, Cell Proliferation, and Cancer. *Int J Mol Sci.* 2020;21(21). doi:10.3390/ijms21218323
 65. Giorgi C, Danese A, Missiroli S, Patergnani S, Pinton P. Calcium Dynamics as a Machine for Decoding Signals. *Trends Cell Biol.* 2018;28(4):258-273. doi:10.1016/j.tcb.2018.01.002
 66. Marchi S, Giorgi C, Galluzzi L, Pinton P. Ca²⁺ Fluxes and Cancer. *Mol Cell.* 2020;78(6):1055-1069. doi:10.1016/j.molcel.2020.04.017
 67. Shaughnessy M, Lamuraglia G, Klebanov N, et al. Selective uveal melanoma inhibition with calcium channel blockade. *Int J Oncol.* 2019;55(5):1090-1096. doi:10.3892/ijo.2019.4873
 68. Yoshida J, Ishibashi T, Nishio M. Antiproliferative effect of Ca²⁺ channel blockers on human epidermoid carcinoma A431 cells. *Eur J Pharmacol.* 2003;472(1-2):23-31. doi:10.1016/s0014-2999(03)01831-4
 69. Kahl CR, Means AR. Calcineurin regulates cyclin D1 accumulation in growth-stimulated fibroblasts. *Mol Biol Cell.* 2004;15(4):1833-1842. doi:10.1091/mbc.e03-10-0730
 70. Yokokura S, Yurimoto S, Matsuoka A, et al. Calmodulin antagonists induce cell cycle arrest and apoptosis in vitro and inhibit tumor growth in vivo in human multiple myeloma. *BMC Cancer.* 2014;14:882. doi:10.1186/1471-2407-14-882
 71. Chovancova B, Liskova V, Miklikova S, et al. Calcium signaling affects migration and proliferation differently in individual cancer cells due to nifedipine treatment. *Biochem Pharmacol.* 2020;171:113695. doi:10.1016/j.bcp.2019.113695
 72. Vaibavi SR, Sivasubramaniapandian M, Vaippully R, Edwina P, Roy B, Bajpai SK. Calcium-channel-blockers exhibit divergent regulation of cancer extravasation through the mechanical properties of cancer cells and underlying vascular endothelial cells. *Cell Biochem Biophys.* 2022;80(1):171-190. doi:10.1007/s12013-021-01035-3
 73. Das A, Pushparaj C, Bahí N, et al. Functional expression of voltage-gated calcium channels in human melanoma. *Pigment Cell Melanoma Res.* 2012;25(2):200-212. doi:10.1111/j.1755-148X.2012.00978.x
 74. Ji Y, Han Z, Shao L, Zhao Y. Ultrasound-targeted microbubble destruction of calcium channel subunit α 1D siRNA inhibits breast cancer via G protein-coupled receptor 30. *Oncol Rep.* 2016;36(4):1886-1892. doi:10.3892/or.2016.5031
 75. Wood H, Dickman A, Star A, Boland JW. Updates in palliative care - overview and recent advancements in the pharmacological management of cancer pain. *Clin Med.* 2018;18(1):17-22. doi:10.7861/clinmedicine.18-1-17
 76. Mathieson S, Lin C-WC, Underwood M, Eldabe S. Pregabalin and gabapentin for pain. *BMJ.* April 2020:m1315. doi:10.1136/bmj.m1315
 77. Brito BE, García MA, De Gouveia YM, et al. Concomitant Antihyperalgesic and Antitumor Effects of Gabapentin in a Murine Cancer Pain Model. *Int J Mol Sci.* 2021;22(18). doi:10.3390/ijms22189671

78. Dooley DJ, Taylor CP, Donevan S, Feltner D. Ca²⁺ channel $\alpha 2\delta$ ligands: novel modulators of neurotransmission. *Trends Pharmacol Sci.* 2007;28(2):75-82. doi:10.1016/j.tips.2006.12.006
79. Inzucchi SE, Bergenstal RM, Buse JB, et al. Management of Hyperglycemia in Type 2 Diabetes, 2015: A Patient-Centered Approach: Update to a Position Statement of the American Diabetes Association and the European Association for the Study of Diabetes. *Diabetes Care.* 2015;38(1):140-149. doi:10.2337/dc14-2441
80. ARGAUD D, ROTH H, WIERNSPERGER N, LEVERVE XM. Metformin decreases gluconeogenesis by enhancing the pyruvate kinase flux in isolated rat hepatocytes. *Eur J Biochem.* 1993;213(3):1341-1348. doi:10.1111/j.1432-1033.1993.tb17886.x
81. Gunton JE, Delhanty PJD, Takahashi S-I, Baxter RC. Metformin Rapidly Increases Insulin Receptor Activation in Human Liver and Signals Preferentially through Insulin-Receptor Substrate-2. *J Clin Endocrinol Metab.* 2003;88(3):1323-1332. doi:10.1210/jc.2002-021394
82. Jeon S-M. Regulation and function of AMPK in physiology and diseases. *Exp Mol Med.* 2016;48(7):e245. doi:10.1038/emm.2016.81
83. Ma T, Tian X, Zhang B, et al. Low-dose metformin targets the lysosomal AMPK pathway through PEN2. *Nature.* 2022;603(7899):159-165. doi:10.1038/s41586-022-04431-8
84. Wang Y, Xu W, Yan Z, et al. Metformin induces autophagy and G0/G1 phase cell cycle arrest in myeloma by targeting the AMPK/mTORC1 and mTORC2 pathways. *J Exp Clin Cancer Res.* 2018;37(1):63. doi:10.1186/s13046-018-0731-5
85. Shi W-Y, Xiao D, Wang L, et al. Therapeutic metformin/AMPK activation blocked lymphoma cell growth via inhibition of mTOR pathway and induction of autophagy. *Cell Death Dis.* 2012;3(3):e275-e275. doi:10.1038/cddis.2012.13
86. Alimova IN, Liu B, Fan Z, et al. Metformin inhibits breast cancer cell growth, colony formation and induces cell cycle arrest in vitro. *Cell Cycle.* 2009;8(6):909-915. doi:10.4161/cc.8.6.7933
87. Ooi MG, McMillin DW, Negri JM, et al. The Antidiabetic Biguanide Metformin Induces Growth Arrest in Multiple Myeloma Cells in Vitro, overcoming the Effect of Stromal Cells. *Blood.* 2008;112(11):2652-2652. doi:10.1182/blood.V112.11.2652.2652
88. Buzzai M, Jones RG, Amaravadi RK, et al. Systemic Treatment with the Antidiabetic Drug Metformin Selectively Impairs p53-Deficient Tumor Cell Growth. *Cancer Res.* 2007;67(14):6745-6752. doi:10.1158/0008-5472.CAN-06-4447
89. Wheaton WW, Weinberg SE, Hamanaka RB, et al. Metformin inhibits mitochondrial complex I of cancer cells to reduce tumorigenesis. *Elife.* 2014;3. doi:10.7554/eLife.02242
90. Li P, Zhao M, Parris AB, Feng X, Yang X. p53 is required for metformin-induced growth inhibition, senescence and apoptosis in breast cancer cells. *Biochem Biophys Res Commun.* 2015;464(4):1267-1274. doi:10.1016/j.bbrc.2015.07.117
91. Ferretti AC, Hidalgo F, Tonucci FM, et al. Metformin and glucose starvation decrease the migratory ability of hepatocellular carcinoma cells: targeting AMPK activation to control migration. *Sci Rep.* 2019;9(1):2815. doi:10.1038/s41598-019-39556-w
92. Wandee J, Prawan A, Senggunprai L, Kongpetch S, Tusskorn O, Kukongviriyapan V. Metformin enhances cisplatin induced inhibition of cholangiocarcinoma cells via AMPK-mTOR pathway. *Life Sci.* 2018;207:172-183. doi:10.1016/j.lfs.2018.05.046
93. Cerezo M, Tichet M, Abbe P, et al. Metformin Blocks Melanoma Invasion and Metastasis Development in AMPK/p53-Dependent Manner. *Mol Cancer Ther.* 2013;12(8):1605-1615.

doi:10.1158/1535-7163.MCT-12-1226-T

94. Pawelczyk L, Spaczynski RZ, Banaszewska B, Duleba AJ. Metformin therapy increases insulin-like growth factor binding protein-1 in hyperinsulinemic women with polycystic ovary syndrome. *Eur J Obstet Gynecol Reprod Biol.* 2004;113(2):209-213. doi:10.1016/j.ejogrb.2003.09.031
95. Pollak M. Insulin and insulin-like growth factor signalling in neoplasia. *Nat Rev Cancer.* 2008;8(12):915-928. doi:10.1038/nrc2536
96. Memmott RM, Mercado JR, Maier CR, Kawabata S, Fox SD, Dennis PA. Metformin Prevents Tobacco Carcinogen-Induced Lung Tumorigenesis. *Cancer Prev Res.* 2010;3(9):1066-1076. doi:10.1158/1940-6207.CAPR-10-0055
97. Yang Y-X, Metz DC. Safety of proton pump inhibitor exposure. *Gastroenterology.* 2010;139(4):1115-1127. doi:10.1053/j.gastro.2010.08.023
98. Moriyama Y, Patel V, Ueda I, Futai M. Evidence for a common binding site for omeprazole and N-ethylmaleimide in subunit A of chromaffin granule vacuolar-type H(+)-ATPase. *Biochem Biophys Res Commun.* 1993;196(2):699-706. doi:10.1006/bbrc.1993.2306
99. Fako VE, Wu X, Pflug B, Liu J-Y, Zhang J-T. Repositioning proton pump inhibitors as anticancer drugs by targeting the thioesterase domain of human fatty acid synthase. *J Med Chem.* 2015;58(2):778-784. doi:10.1021/jm501543u
100. De Milito A, Canese R, Marino ML, et al. pH-dependent antitumor activity of proton pump inhibitors against human melanoma is mediated by inhibition of tumor acidity. *Int J Cancer.* 2010;127(1):207-219. doi:10.1002/ijc.25009
101. Zhang B, Yang Y, Shi X, et al. Proton pump inhibitor pantoprazole abrogates adriamycin-resistant gastric cancer cell invasiveness via suppression of Akt/GSK- β / β -catenin signaling and epithelial-mesenchymal transition. *Cancer Lett.* 2015;356(2):704-712. doi:10.1016/j.canlet.2014.10.016
102. Feng S, Zheng Z, Feng L, et al. Proton pump inhibitor pantoprazole inhibits the proliferation, self-renewal and chemoresistance of gastric cancer stem cells via the EMT/ β -catenin pathways. *Oncol Rep.* 2016;36(6):3207-3214. doi:10.3892/or.2016.5154
103. Wang CJ, Li D, Danielson JA, et al. Proton pump inhibitors suppress DNA damage repair and sensitize treatment resistance in breast cancer by targeting fatty acid synthase. *Cancer Lett.* 2021;509:1-12. doi:10.1016/j.canlet.2021.03.026
104. Babu D, Mudiraj A, Yadav N, Y.B.V.K. C, Panigrahi M, Prakash Babu P. Rabeprazole has efficacy per se and reduces resistance to temozolomide in glioma via EMT inhibition. *Cell Oncol.* 2021;44(4):889-905. doi:10.1007/s13402-021-00609-w
105. Paškevičiūtė M, Petrikaitė V. Proton Pump Inhibitors Modulate Transport Of Doxorubicin And Its Liposomal Form Into 2D And 3D Breast Cancer Cell Cultures. *Cancer Manag Res.* 2019;11:9761-9769. doi:10.2147/CMAR.S224097
106. Luciani F, Spada M, De Milito A, et al. Effect of Proton Pump Inhibitor Pretreatment on Resistance of Solid Tumors to Cytotoxic Drugs. *JNCI J Natl Cancer Inst.* 2004;96(22):1702-1713. doi:10.1093/jnci/djh305
107. Martínez-Zaguilán R, Sennoune SR. Vacuolar H⁺-ATPase Signaling in Cancer. In: *Regulation of Ca²⁺-ATPases, V-ATPases and F-ATPases.* Cham: Springer International Publishing; 2016:371-392. doi:10.1007/978-3-319-24780-9_18

108. Lee AH, Tannock IF. Heterogeneity of intracellular pH and of mechanisms that regulate intracellular pH in populations of cultured cells. *Cancer Res.* 1998;58(9):1901-1908. <http://www.ncbi.nlm.nih.gov/pubmed/9581831>.
109. Raghunand N, Martínez-Zaguilán R, Wright SH, Gillies RJ. pH and drug resistance. II. turnover of acidic vesicles and resistance to weakly basic chemotherapeutic drugs. *Biochem Pharmacol.* 1999;57(9):1047-1058. doi:10.1016/S0006-2952(99)00021-0
110. Sun L, Yao Y, Pan G, et al. Small interfering RNA-mediated knockdown of fatty acid synthase attenuates the proliferation and metastasis of human gastric cancer cells via the mTOR/Gli1 signaling pathway. *Oncol Lett.* May 2018. doi:10.3892/ol.2018.8648
111. Wu X, Dong Z, Wang CJ, et al. FASN regulates cellular response to genotoxic treatments by increasing PARP-1 expression and DNA repair activity via NF-κB and SP1. *Proc Natl Acad Sci.* 2016;113(45). doi:10.1073/pnas.1609934113
112. Kara A, Özgür A, Nalbantoğlu S, Karadağ A. DNA repair pathways and their roles in drug resistance for lung adenocarcinoma. *Mol Biol Rep.* 2021;48(4):3813-3825. doi:10.1007/s11033-021-06314-z
113. Flavahan WA, Gaskell E, Bernstein BE. Epigenetic plasticity and the hallmarks of cancer. *Science.* 2017;357(6348). doi:10.1126/science.aal2380
114. Sarmiento-Ribeiro AB, Scorilas A, Gonçalves AC, Efferth T, Trougakos IP. The emergence of drug resistance to targeted cancer therapies: Clinical evidence. *Drug Resist Updat.* 2019;47:100646. doi:10.1016/j.drup.2019.100646
115. Sun W, Sanderson PE, Zheng W. Drug combination therapy increases successful drug repositioning. *Drug Discov Today.* 2016;21(7):1189-1195. doi:10.1016/j.drudis.2016.05.015
116. Xia T, He Q, Shi K, et al. Losartan loaded liposomes improve the antitumor efficacy of liposomal paclitaxel modified with pH sensitive peptides by inhibition of collagen in breast cancer. *Pharm Dev Technol.* 2018;23(1):13-21. doi:10.1080/10837450.2016.1265553
117. Shields CL, Furuta M, Thangappan A, et al. Metastasis of uveal melanoma millimeter-by-millimeter in 8033 consecutive eyes. *Arch Ophthalmol (Chicago, Ill 1960).* 2009;127(8):989-998. doi:10.1001/archophthalmol.2009.208
118. Sharma A, Jacob A, Tandon M, Kumar D. Orphan drug: Development trends and strategies. *J Pharm Bioallied Sci.* 2010;2(4):290-299. doi:10.4103/0975-7406.72128
119. Salacz ME, Watson KR, Schomas DA. Glioblastoma: Part I. Current state of affairs. *Mo Med.* 108(3):187-194. <http://www.ncbi.nlm.nih.gov/pubmed/21736079>.
120. van den Berg S, de Visser S, Leufkens HGM, Hollak CEM. Drug Repurposing for Rare Diseases: A Role for Academia. *Front Pharmacol.* 2021;12. doi:10.3389/fphar.2021.746987