# Methods to assess PM<sub>2.5</sub> exposure from indoor sources in epidemiological studies: a review

# By Nekane Sandoval Diez

Student Number: 7053673

n.sandovaldiez@uu.nl

Examiners:

**Dr. G. (Gerard) Hoek** Fac. of Veterinary Medicine Dept. Population Health Sciences IRAS Dr. U. (Ulrike) Gehring

Fac. of Veterinary Medicine

Dept. Animals, science and society

IRAS

#### 0. Layman's Summary

Of all air pollutants, particles with a diameter of less than or equal to  $2.5 \ \mu m (PM_{2.5})$  have the greatest scientific evidence of adverse health effects, including respiratory, cardiovascular, nervous system effects, cancer, and mortality. Epidemiological studies are an invaluable and very important tool to establish the toxicity and effects of PM\_{2.5} on human health. Most epidemiological studies have measured or modelled ambient concentrations of PM\_{2.5} to estimate human exposure. However, humans spend most of their time indoors, where almost all exposure to air pollutants takes place.

Indoor PM<sub>2.5</sub> concentration is a mixture of both ambient and non-ambient particles. The ambient component refers to all the particles generated outdoors that infiltrate into the indoor environment through, for example, ventilation. Nonambient particles refer to all those particles that are produced directly indoors or by human personal activities. Many human activities can be sources of indoor PM<sub>2.5</sub>, although the most important are smoking, cooking, and heating. Outdoor and indoor PM<sub>2.5</sub> sources are different, so the particle's composition, toxicity, and human health effects may also be distinct. Therefore, it is important to treat exposure to outdoor- and indoor-generated PM<sub>2.5</sub> as two separate exposures to investigate their effects in epidemiological studies.

To separate indoor- and outdoor-generated PM<sub>2.5</sub> exposure, researchers have used tracer compounds whose production occurs almost exclusively outdoors and infiltrate into indoor environments, a method called source partitioning. However, the separation between indoor- and outdoor-generated PM<sub>2.5</sub> implies many logistical challenges for epidemiological research since it often entails a great economic cost for the researchers to carry out measurements in each of the residences or indoor environments. It can also be inconvenient for the participants. This is why most of the studies that measure indoor PM<sub>2.5</sub> concentrations do so in small study populations or for short periods of time. Having small study populations limits the ability of epidemiological studies to draw valid results.

In this literature review, we aimed to evaluate the approaches and techniques used to assess indoor-generated PM<sub>2.5</sub> exposure in epidemiological studies of respiratory health effects, pointing out their strengths and limitations. We identified a total of 29 epidemiological studies that carried out measurements of indoor PM<sub>2.5</sub> concentrations, of which only 5 applied methods to separate indoor- and outdoor-generated particles. All studies that applied source partitioning methods used sulphate or iron as tracer compounds, had small study populations, and studied short-term exposures. The results of studies applying source partitioning methods highlight the importance of investigating the specific relationship between indoor-generated PM<sub>2.5</sub> and respiratory health outcomes, as well as distinguishing it from outdoor sources.

# 1. Introduction

Numerous epidemiological studies have documented the relationship between air pollution and health. Among all the fractions of particulate matter (PM) in air pollution, fine PM (PM<sub>2.5</sub> or particles with aerodynamic diameters less than or equal to 2.5  $\mu$ m) shows the most substantial and consistent scientific evidence for relationships between short-term or long-term exposure and health [1]. PM<sub>2.5</sub> is associated with multiple adverse health outcomes, including respiratory, cardiovascular, and nervous system effects, cancer, and increased total mortality [1,2]. However, most epidemiological evidence is derived from studies that capture only the effect of ambient PM<sub>2.5</sub>, usually assessing exposure through outdoor monitoring stations or outdoor modelling. These studies do not address the health impact of nonambient fine PM.

People spend most of their lives in indoor microenvironments with an estimated 87% of time spent inside buildings [3], so these environments mediate human exposure to both outdoor- and indoor-generated particles. Since air exchange rates in indoor microenvironments have been reduced in modern buildings and the ambient concentration of PM<sub>2.5</sub> started to decrease in recent years [4], the contribution of indoor PM<sub>2.5</sub> sources to personal exposure will become increasingly important. PM<sub>2.5</sub> was found to be one of the most harmful non-biological indoor air pollutants, associated with the largest number of DALY losses, and responsible for most chronic health effects in homes without smoking [5]. Additionally, it has been reported that PM<sub>2.5</sub> from indoor sources may be the dominant fraction of integrated daily residential exposure, with nearly 30% of the disease burden from PM exposure attributable to indoor and outdoor sources have similar health effects, which is controversial. A pilot study suggested that indoor-generated fine PM may be more toxic and bioactive compared to fine PM from outdoor sources [7].

Personal exposure to  $PM_{2.5}$  includes ambient and nonambient components, both of which can differ in composition/size and produce different types of health effects or, at least, involve different sources that would be desirable to identify in order to implement cost-effective measures to reduce or prevent exposure [8]. It has been shown that there is usually a low-to-moderate correlation between indoor-generated and outdoor PM concentrations [9–12]; thus, studies that show a relationship between variations in health endpoints and outdoor PM cannot provide any information regarding the possible health effects of indoor-generated PM. Epidemiological studies with an appropriate characterization of particles from indoor sources are required to determine the health effects of indoor-generated PM\_2.5, as well as its composition-specific toxicity.

Two approaches can be used to assess the risks associated with indoor-generated PM<sub>2.5</sub>: risk assessment and epidemiological studies. The former approach is based on exposure data from human exposure studies, where exposure is determined by modelling or

conducting direct measurements of indoor PM<sub>2.5</sub> concentrations. Exposure data is then used in conjunction with exposure-response relationships derived from experimental or analytical toxicological and epidemiological studies to perform risk characterization. The most serious disadvantage of this approach is that it generally uses exposure-response relationships derived from outdoor settings [5,13], assuming that exposure to indoor-generated and outdoor-generated PM<sub>2.5</sub> are equal in terms of toxicity (PM<sub>2.5</sub> is said to be equitoxic). In addition, source variability, as well as temporal and personal variability of particles from indoor sources, are not considered.

In contrast, epidemiological studies of the effects of indoor-generated PM<sub>2.5</sub> need to assess exposure produced indoors and relate it to an outcome of interest in the studied population. In this way, compositional toxicity dependencies, indoor sources, and temporal or personal variability can be properly addressed. However, epidemiological studies are often limited by cost and feasibility constraints imposed by indoor exposure assessment and the inability to directly measure indoor-generated particles because outdoor sources also contribute significantly to indoor PM concentrations. Therefore, most studies attempting to investigate the health effects of indoor particulate matter are generally limited to small cohorts or perform exposure characterization based on more approximate exposure assessment methods, such as exposure indicators [14]. In this review, we do not address the well-described exposure indicator literature but focus on the more challenging PM monitoring/modelling approaches. In Supplement 1 we briefly discuss the large body of evidence of exposure indicators.

It is important to treat indoor-generated PM<sub>2.5</sub> as a distinct and separate exposure from outdoor-generated particles or other indoor-produced pollutants to ensure proper assessment of its related human health risks. Quantitatively assessing exposure to fine PM from indoor sources is a challenging task. Because of this, the present review aims to evaluate the current approaches and techniques used in epidemiological research to assess exposure to PM<sub>2.5</sub> from indoor origin, summarizing their strengths and limitations. We focused only on epidemiological studies reporting quantitative exposure measurements of the fine fraction of PM in indoor environments. Specifically, we restrict our literature search to direct (personal exposure monitoring) or indirect (exposure modelling techniques and indoor concentration measurements) exposure assessment methods, which represent better estimates for personal exposure than exposure indicators [15,16]. Since respiratory outcomes are the most common health effects of fine particulate matter exposure reported in the literature, this review was limited to epidemiological studies assessing respiratory health. Including all possible health outcomes was not feasible due to time constraints.

In this paper, we first describe our literature search (Section 2), followed by a background section including a summary of the fundamentals of exposure assessment methods to indoor air pollutants (Section 3). Section 4 gives a short overview of PM<sub>2.5</sub> indoor sources, levels observed in indoor microenvironments, and existing general

challenges to its exposure assessment. In Section 5, we present a review of the exposure assessment approaches applied in the identified epidemiological studies of respiratory health.

# 2. Methods

Only articles written in English and published in peer-reviewed journals about the association between indoor PM<sub>2.5</sub> exposure and respiratory health outcomes were included in this literature review. The literature search was performed using the electronic databases PubMed, Google Scholar, and Web of Science. Potentially relevant studies were identified and screened for retrieval using the following keywords: "indoor PM2.5", OR "PM2.5 indoor environments", OR "indoor fine particulate matter", AND "health effects", OR "asthma", "COPD", OR "lung cancer" OR "respiratory health", OR "epidemiological study", OR "cohort". In addition, we complemented the search using the most relevant references of the identified articles.

# 3. Exposure assessment to indoor particulate matter

In epidemiological studies, exposure assessment is a major component with the same importance as outcome assessment. Exposure in relation to particulate matter is generally expressed in terms of air concentration ( $\mu$ g/m<sup>3</sup>) and can be defined as the PM concentration near the breathing zone during a certain period (concentration x time) [11]. To represent an individual's total exposure to PM, we must consider all the microenvironments that contributed to breathable air for the individual and the time spent in those microenvironments. In other words, exposure is a function of both space and time and most studies report average exposures in a certain setting (e.g., 12- or 24-hour-averages).

Exposure to total PM<sub>2.5</sub> (T<sub>PM2.5</sub>) is constructed from ambient PM<sub>2.5</sub> (A<sub>PM2.5</sub>) and nonambient PM<sub>2.5</sub> (NA<sub>PM2.5</sub>) [8]. Nonambient PM<sub>2.5</sub> is in turn made up of indoorgenerated PM<sub>2.5</sub> and PM<sub>2.5</sub> generated from personal activities in any type of setting (PA<sub>PM2.5</sub>, also referred to as "personal cloud" [8]):

# $T_{PM2.5} = A_{PM2.5} + NA_{PM2.5}$ $NA_{PM2.5} = IG_{PM2.5} + PA_{PM2.5}$

Ambient  $PM_{2.5}$  (A<sub>PM2.5</sub>) are fine particles that are emitted (primary PM) or formed (secondary PM) in the ambient atmosphere. Indoor  $PM_{2.5}$  (I<sub>PM2.5</sub>) includes all fine particulate matter found indoors and can be divided into  $PM_{2.5}$  that is emitted or formed indoors (IG<sub>PM2.5</sub>) and PM from an ambient origin that infiltrates indoors depending on ventilation and infiltration conditions (AI<sub>PM2.5</sub>) [8]. Thus, the indoor  $PM_{2.5}$  concentration is given by:

$$I_{\rm PM2.5} = IG_{PM2.5} + AI_{PM2.5}$$

It is important to note that neither IGPM2.5 nor AIPM2.5 can be directly measured, so they must be derived from mass balance equations using other measurable quantities. The indoor PM<sub>2.5</sub> (I<sub>PM2.5</sub>) concentration can also be expressed through mass balance equations in equilibrium conditions as follows [17]:

$$I_{PM2.5} = \frac{Pa}{a+k} A_{PM2.5} + \frac{Q}{V(a+k)}$$

where  $I_{PM2.5}$  is the indoor PM<sub>2.5</sub> concentration (µg/m<sup>3</sup>),  $A_{PM2.5}$  is the ambient PM<sub>2.5</sub> concentration (µg/m<sup>3</sup>), P is the penetration efficiency (unitless), a is the air exchange rate (h<sup>-1</sup>), k is the decay rate (h<sup>-1</sup>), Q is the indoor sources strength (µg/h), and V is the volume of the indoor environment (m<sup>3</sup>).

The penetration efficiency of  $PM_{2.5}$  (*P*) can be defined as the fraction of ambient  $PM_{2.5}$  that is not removed from ambient air when it enters indoor environments.

From the above equation, PM<sub>2.5</sub> indoor concentrations can be solved for its two components [(8,17)]:

$$AI_{PM2.5} = \frac{Pa}{a+k} A_{PM2.5} = F_{INF} A_{PM2.5}$$
$$IG_{PM2.5} = \frac{Q}{V(a+k)}$$

where  $F_{INF}$  is the infiltration factor and describes the fraction of outdoor air that infiltrates indoors and remains suspended [17]. Thus,  $F_{INF}$  is a function of penetration rates (*P*) and decay rates (*k*). *P* can be defined as the fraction of ambient PM<sub>2.5</sub> that is not removed from ambient air when it enters indoor environments, and *k* describes the particle loss processes by diffusion or sedimentation.

To determine the effects of indoor-generated PM<sub>2.5</sub>, epidemiological studies must measure indoor and outdoor concentrations of PM<sub>2.5</sub> or directly measure personal exposure in indoor environments. The challenging task is the separation of indoor PM<sub>2.5</sub> concentrations or personal exposure measurements into its indoor-generated and outdoor-generated components. Measurements of the indoor source strength (Q) have rarely been done due to the difficulty of accurately measuring the emission rate of each indoor source. Similarly, the estimation of other indoor concentration parameters such as penetration efficiency and decay rates is a complicated task due to the influence exerted by other factors, like indoor and outdoor temperature, humidity, building characteristics, and wind speed [18]. Therefore, studies usually must rely on source partitioning methods that estimate  $F_{INF}$  to calculate the fraction of PM<sub>2.5</sub> generated outdoors and derive the indoor-generated component from the total indoor PM<sub>2.5</sub> concentration (estimation of  $F_{INF}$  using a tracer method is discussed in more detail in Section 5 of this review):

## $IG_{PM2.5} = I_{PM2.5} - AI_{PM2.5}$

There are direct and indirect methods for exposure assessment and measurement of indoor PM<sub>2.5</sub> concentrations [2,11]. Direct approaches refer to those methods where exposure is measured through a monitoring device, such as Personal Exposure Monitors (PEM), or through biomarkers of exposure. PEMs are worn by study subjects and record detailed exposure data as the individual remains in a given microenvironment, generally through filter-based mass measurements or light scattering instruments [11]. When combined with time-activity data, they can provide detailed exposure information from indoor sources. However, PEMs usually represent a great burden for the participants and entail a high cost for the researchers; thus, serving only to evaluate short-term effects and within limited sample sizes. Currently, there is no biomarker that can be used in epidemiological studies to reflect PM<sub>2.5</sub> exposure, although it is a topic under investigation [19,20].

Indirect approaches make use of modelling techniques and other measurable quantities to derive or estimate personal exposure. Microenvironmental monitoring measures the concentration of PM<sub>2.5</sub> indoors and, along with time-activity data, can be used to determine integrated exposures for the time spent in each microenvironment and identify sources. To reliably reflect personal exposure, microenvironmental measurements must have a homogeneous distribution in time and space during the measurement period [15]. Both microenvironmental monitoring and time-activity data are input information for modelling exposure techniques.

Exposure models with varying levels of complexity are fitted using two general approaches: time-series and time-average [15]. In the former, microenvironmental exposures are estimated sequentially over time. For the time-average approach, the estimates of exposure are calculated for the time spent on average in each microenvironment. Modelling techniques can be further classified into deterministic and stochastic models [16]. While deterministic models (also known as physical models) mathematically describe the relationship between sources and PM concentration based on prior knowledge of pollutant properties, stochastic models (also known as statistical models) are based on the measured concentration of a representative sample of microenvironments which are regressed by certain variables [16].

#### 4. PM<sub>2.5</sub> indoor levels and sources: challenges for exposure assessment

Indoor PM emissions can be sporadic, episodic, or continuous, and are site and time specific. Smoking, cooking, and heating constitute the most important PM<sub>2.5</sub> indoor sources. Simoni et al. showed that homes with a smoker had about  $33 \,\mu\text{g/m}^{-3}$  higher 48-hour average PM<sub>2.5</sub> concentrations, with a reported 48-h average increment of 0.2  $\mu\text{g/m}^{3}$  for each smoked cigarette [21]. Coal, wood, or other biomass fuel combustion for cooking or heating are also recognized as important sources of fine PM [2,22], with the largest peaks of indoor PM<sub>2.5</sub> concentrations found to be during cooking activities

[23,24]. Other known indoor sources of PM<sub>2.5</sub> are human movement [25], burning candles or incense [26], cleaning activities like vacuuming and sweeping [(8,27], usage of printers, fax machines, or photocopiers [(28)], and the application of anti-insect products, cleaning agents or cosmetics [29,30].

Indoor PM shows large spatial (within and between indoor environments) and temporal variability and may have higher concentration levels than outdoor PM, even in non-smoking households [31]. Morawska and Salthammer reported in a review of 14 representative studies that for naturally ventilated buildings in the absence of known indoor sources, the median value of the indoor to outdoor concentration ratio (I/O ratio) of PM<sub>2.5</sub> was 0.91, ranging from 0.54 to 1.08, showing the significant contribution of outdoor air as a source of indoor particles. In contrast, when known indoor sources were present I/O ratios ranged from 1 to 2.4, with a median value of 1.21 [25]. Furthermore, a study conducted in four European countries differentiating between the outdoor- and indoor-generated components of the PM<sub>2.5</sub> indoor concentration found that the contribution of indoor sources in non-smoking households represented 20-30% of the total indoor levels, with reported indoor- and outdoor-generated concentrations of 3–5  $\mu$ g/m3 and 6-20  $\mu$ g/m3, respectively [32].

The assessment of exposure to PM<sub>2.5</sub> generated indoors is complex in part because it has not been established which physical or chemical properties are responsible for its toxicity. PM<sub>2.5</sub> also represents a mixture of pollutants, unlike other well-known criteria gaseous pollutants like carbon monoxide (CO). CO is a widely distributed odourless and colourless gas mainly formed from anthropogenic emissions of incomplete combustion of carbonaceous material like coal, wood, natural gas, petrol, and kerosene [33]. Similar to fine PM, indoor concentrations of CO are the result of infiltration of ambient CO and the presence of nonambient combustion sources [34]. However, as its toxicity is not source-specific, the distinction between indoor and outdoor CO sources is not important for the study of health effects and both types of sources contribute to a relevant total exposure. Thus, exposure to CO can be assessed in epidemiological studies with a highly specific internal dose metric like carboxyhaemoglobin (COHb) or through personal monitoring analytic methods that estimate total personal exposure [34].

The concentration, sources, and characteristics of indoor particles are also highly indoorspecific and can vary within and between microenvironments. Carrying out activities that generate indoor PM<sub>2.5</sub> usually results in a heterogeneous distribution of particles throughout space [25], so it would be important to consider whether PM<sub>2.5</sub> concentration measurements in a specific compartment of a microenvironment (such as the living room in a residence) constitutes a relevant estimate of exposure. As already noted in section 3, microenvironmental monitoring is only a reliable estimate of exposure if PM<sub>2.5</sub> is well-mixed indoors. If this does not happen, the exposure could be underestimated or overestimated, and the effect estimates are likely biased to the null (classical error model) [35]. Although the residence represents the indoor microenvironment where people spend most of their time [3], sources of residential PM<sub>2.5</sub> can differ from those found in other microenvironments also relevant for health (e.g., schools, vehicles, hospitals, etc.). Therefore, exposure-response relationships are subject to the assumption that all indoor environments are well represented by the microenvironment in which the concentration measurements are made (usually the residence). In this case, the use of time-activity data becomes of utmost importance to estimate exposure levels relevant to the time spent in each environment. Additionally, the variability of emission factors between sources of the same type is substantial. For example, PM<sub>2.5</sub> emissions from indoor cooking can vary between developed and developing countries, where the type of stove used is different (fuel or wood stoves vs electric stoves). The fine particles produced can also vary according to the cooking style, type of food cooked, type of oil used, and cooking temperature [36].

# 5. <u>Indoor PM<sub>2.5</sub> exposure assessment methods in epidemiological studies of</u> <u>respiratory health</u>

We identified 29 studies that performed quantitative measurements of PM<sub>2.5</sub> in indoor environments to determine the effects of indoor fine PM on respiratory health endpoints. Tables 1 and 2 show the key study design characteristics of the identified epidemiological studies evaluating associations with respiratory health of short-term (n=20) and long-term exposure (n=9). Short-term exposure studies were primarily designed as panel studies and had, on average, a smaller sample size than long-term studies (median sample size of 46 and 150, respectively). Most studies have been published from 2010 onwards and have been conducted mostly in North America (n=17, 58.6%) and Europe (n=8, 27.6%), with a minority carried out in Asia (n=4, 13.8%) and without representation from studies performed in South America or Africa. Finally, the majority of the selected studies evaluated respiratory outcomes in vulnerable populations, such as children or older adults with asthma or COPD.

Tables 3 and 4 list the exposure assessment methods for short- and long-term effects epidemiological studies, respectively. Of the identified studies, nearly all conducted measurements of indoor  $PM_{2.5}$  concentrations at the participants' residence (n=28); with only one study performing measurements in a different indoor microenvironment (i.e. hospital, Ma et al. [37]). Of the places where the measurements were made inside the residences, the living room was the most common (n=20, 69.0%), followed by the bedroom (n=10, 34.5%), and only four studies made measurements in another room (e.g., kitchen or farm workplace). Six studies also reported conducting personal exposure monitoring (PEM) and 13 collected time-activity data.

## 5.1 Epidemiological studies using source partitioning methods

Only five epidemiological studies performed source partitioning of indoor concentrations between indoor-generated and outdoor-generated PM<sub>2.5</sub> [38–42]

(Table 3). All of them used a tracer element without indoor sources as a method to estimate the infiltration factor ( $F_{INF}$ ) and the application of deterministic models to derive the fraction of PM<sub>2.5</sub> from indoor sources using mass balance equations. One of these studies also applied a stochastic model to complement the estimates (Koening et al. [40]).

The use of a tracer element to estimate the fraction of outdoor-generated  $PM_{2.5}$  is based on the assumptions that such an element primarily comes from outdoor sources, indoor and personal activity sources are practically negligible, and the physical behaviour of the element is similar to that of other outdoor  $PM_{2.5}$  constituents [43]. A tracer compound allows the estimation of  $F_{INF}$  without having to measure the penetration efficiency (P), air exchange rate (a), and decay rate (k) of  $PM_{2.5}$  for each indoor setting, which would be unfeasible in epidemiological studies. However, the costs of carrying out filtered-based measurements to identify tracer compounds are still high enough to prevent their application in epidemiological studies of long-term exposure since we did not find any long-term studies using this method.

It should be noted that the use of a tracer element (sulphur or iron in our identified studies) could also add uncertainty to indoor PM<sub>2.5</sub> exposure estimates. The largest fraction of sulphur-bound particles is in the sub-micron particle size range compared to PM which is more concentrated in larger particle sizes. Because of this, penetration and decay rates of PM, and thus infiltration rates, may be different from that of PM-bound sulphur, leading to potential misclassification [32].

The Vancouver Panel Study (Ebelt et al. [38], Wilson and Brauer [39]) was the first to investigate the association between ambient and nonambient PM<sub>2.5</sub> with lung and cardiovascular function in 16 adults with COPD. Total personal exposures for PM<sub>2.5</sub> and sulphate were measured for each participant and, together with time-activity data describing the amount of time spent outdoors, the ambient exposure fraction was calculated from the measured ambient concentration at five central stations within the study area. Sulphate was used as a tracer compound of the outdoor-generated PM<sub>2.5</sub> since it has practically no indoor or personal activity sources [43,44] and, thus, the total personal exposure to sulphate can be assumed to come entirely from outdoor sources. The nonambient component was calculated as the subtraction of the total exposure minus the ambient component. Unlike the other studies, this study used PEM and time activity data instead of indoor air monitoring concentrations to estimate the nonambient exposure. Furthermore, the study excluded a major indoor source of PM<sub>2.5</sub> by not including participants who smoked or lived in smoking households, so the results mainly reflect the effect of other major indoor sources than smoking or personal activities performed in outdoor or indoor settings.

The other three studies reporting source partitioning also applied deterministic models to differentiate the indoor-generated  $(IG_{PM2.5})$  and outdoor-generated  $(AI_{PM2.5})$ 

components of indoor PM<sub>2.5</sub> concentration ( $I_{PM2.5}$ ) (Table 3). Koening et al. [40] and Habre et al. [41] applied mass balance equations to estimate the infiltration factor of the residences of each of the children with asthma who were included in their studies. Sulphate was used as a tracer method of outdoor sources in the mass balance model. In addition, Koening et al. fitted a predictive model to estimate  $F_{INF}$  since the measurements of sulphate using a radiance nephelometer were only valid for a subsample of the residences. The predictive model applied was developed in a previous study [45] and considered residence indicators such as the type of dwelling, the use of air cleaners, and surrogates of home ventilation conditions. The authors reported a good agreement between estimates of  $F_{INF}$  using the sulphate-tracer method and the predictive model. Compared to the others, the study by Chi et al. [42] was the only epidemiological study differentiating between exposure to particles of indoor origin and those generated outdoors that was carried out in a non-first-world country and used a different tracer element: iron. It has been demonstrated that iron, and not sulphate, is a better indicator of particulate matter from outdoor sources in China in relation to the existence of different anthropogenic PM<sub>2.5</sub> sources in this country [46].

Air monitoring at a personal level or measurement of indoor PM<sub>2.5</sub> concentrations limits the applicability of source partitioning methods at larger scales and for long-term exposures, as it is labour-intensive and expensive. This is evidenced by the small sample size of the identified studies that applied these methods, which were small-scale panel studies with sample sizes ranging from 16 to 75. Therefore, the major disadvantage of source partitioning methods, and of all indoor PM<sub>2.5</sub> studies in general, is low statistical power due to small sample size. An underpowered study implies problems in the veracity of the results because the probability of finding a genuine true result is low (high rate of false negatives) and the probability that a significant finding is true is decreased (low Positive Predictive Value or PPV) [47]. In addition, external validity may also be compromised as findings tend to be less replicable [47,48].

Another challenge for source partitioning methods is the high temporal and spatial variability of indoor PM<sub>2.5</sub> concentrations, and how to capture this information to reduce error in exposure determination without losing feasibility. The level of complexity of mass balance models could become large enough to prevent their application in an epidemiological setting if the input information required is too detailed. For example, indoor concentrations of PM<sub>2.5</sub> and the contributions from indoor or outdoor sources can vary greatly throughout the year depending on the season. In winter and autumn, where ventilation rates are typically lower, indoor sources may be more important for total indoor PM<sub>2.5</sub> concentration than in spring and summer [49]. The temporal variability could also be affected by human activity: at night when people are asleep and static for long periods, indoor particle concentrations are usually lower than during the day [24]. These aspects can be addressed in part in the study design phase, conducting

the research only in a certain season, and collecting time-activity data to develop relevant exposure estimates.

In the identified studies with reported source partitioning, there is consistent evidence of differential effects and degrees of toxicity between fine PM from indoor and outdoor sources (Table 5). The studies conducted in Canada [38,39] found that the association estimates with different health outcomes were of greater magnitude for particles of ambient origin than for nonambient exposures. In the same line, Koening et al. [40] saw that PM<sub>2.5</sub> of outdoor origin was more potent per unit mass than particles of indoor origin for the development of airway inflammation. However, they also observed that PM<sub>2.5</sub> of indoor origin was associated with decrements in lung functions, indicating that the potential effects of indoor-vs outdoor-generated particles differed for different health endpoints. Another study conducted on children with asthma also found a difference in asthma-related symptoms: while PM from indoor sources was significantly associated with odds of more severe wheezing, PM from outdoor sources was associated with odds of more severe cough [41]. Finally, Chi et al. reported that outdoor-generated PM<sub>2.5</sub> was associated with increased blood pressure levels and with decreased pulmonary function in healthy elderly adults, whereas PM<sub>2.5</sub> from indoor sources was linked with decreased pulmonary function in COPD patients [42]. Therefore, the results of these studies suggest that health effects may vary between indoor and outdoor particles, depending on health outcome, population, and type of indoor source.

#### 5.2 Epidemiological studies not using source partitioning methods

In the studies that did not report source partitioning methods (n=24), indoor PM<sub>2.5</sub> concentration was used as a proxy for exposure to indoor-generated PM<sub>2.5</sub>. Some studies attempted to differentiate the effects of outdoor and indoor PM using different methods. 14 addressed the issue at the study design phase (e.g., limiting the inclusion of smokers or choosing populations with high levels of in-home combustion) [37,50–62], and eight did so in the statistical analysis [63–70], either with multiple-exposure models (i.e., adjusting the relationship between indoor PM<sub>2.5</sub> and health endpoints for outdoor concentrations) or adjusting the relationship between indoor PM<sub>2.5</sub> and health endpoints for questionnaire-based indicators (i.e., smoking, cooking, and residential behaviours). Finally, two did not mention performing any restrictions or adjustments in relation to the indoor concentrations [71,72]. The decision to take any of the above approaches depended largely on the study question and the aims of the research.

Studies that took the total concentration of indoor PM<sub>2.5</sub> for the analysis of respiratory effects assumed that there was no difference between indoor- and outdoor-generated particles in terms of toxicity and type of health outcome analysed. Kim et al. conducted a randomized intervention study to evaluate the efficacy of air purifiers with High-Efficiency Particulate Air (HEPA) filters in reducing indoor levels of PM<sub>2.5</sub> and the consequent effect on respiratory function in asthmatic children [71]. Because this study

was focused on reducing total levels of indoor fine PM, it was not necessary to distinguish between indoor and outdoor sources. The birth cohort study carried out by Raaschou-Nielsen et al. aimed to study the long-term relationship of indoor air pollutants with bronchial hyperresponsiveness measured through wheezing symptoms in infants [72]. In this study, the interest in indoor PM<sub>2.5</sub> was to obtain an estimate of total exposure, rather than treating indoor- and outdoor-generated PM<sub>2.5</sub> as distinct exposures.

Limiting the study population by indoor sources was the most used method to differentiate between indoor and outdoor sources in studies using only indoor concentrations as exposure metric; however, this approach does not resolve the controversial assumption of assuming equitoxicity between outdoor-generated particles and indoor-generated PM<sub>2.5</sub>. For example, several studies limited the study population to non-smokers living in smoker-free homes [51,54,56–59,61], thus giving priority to the analysis of the effects of outdoor-generated PM<sub>2.5</sub> found indoors and assumed little or no effect from other PM<sub>2.5</sub> indoor sources. In contrast, Butz et al. [50] and Peng et al. [60] limited their study population to children residing in smoking households to assess the health impact of air cleaners reducing PM<sub>2.5</sub> from second-hand tobacco smoke (SHS).

Adjusting effect estimates for outdoor  $PM_{2.5}$  concentrations could be an effective method to capture exposure-response relationships of indoor-generated  $PM_{2.5}$  provided that outdoor concentrations constitute a good proxy of exposure to outdoor-generated PM that infiltrated indoors, that is that the correlation between  $AI_{PM2.5}$  and outdoor concentrations is high, and that outdoor-generated PM<sub>2.5</sub> is an important contributor to total indoor PM<sub>2.5</sub> concentrations. If indoor concentrations are statistically independent of ambient concentrations, then the relation between indoor PM<sub>2.5</sub> and the health outcome of interest cannot be confounded by ambient concentrations [8]. Using measurements made right outside the indoor location under study or modelling techniques will be better than using central site concentrations due to high geographic variability. In the same way, adjusting for indicators of indoor exposure (for example, hair nicotine) will be effective if the indicator reflects a true exposure to the indoor source.

Studies that did not perform source partitioning were also limited by the cost, feasibility, and participant burden limitations of indoor air and personal monitoring methods, although to a lesser extent than source partitioning studies. Indication of this is that the study samples were generally higher and that all the long-term exposure studies did not perform source partitioning. These studies added uncertainty to the exposure assessment of indoor-generated PM<sub>2.5</sub> since they did not make a straightforward distinction between outdoor and indoor sources.

#### 6. Conclusions

Not many epidemiological studies of respiratory health effects have performed indoor PM<sub>2.5</sub> monitoring, compared to the large number of research conducted with exposure indicators. Even fewer studies have applied source partitioning methods to separate indoor- and outdoor-generated particles. Current source partitioning methods are a complicated task that involves several logistical challenges to large-scale epidemiological studies, particularly those of long-term exposure. The five studies that applied sourcepartitioning were all short-term exposure studies in small study populations. The results from studies that did apply source partitioning methods underscore the importance of investigating the specific exposure-response relationship for PM<sub>2.5</sub> originated indoors and distinguishing it from that of outdoor sources. While undeniably useful in addressing research questions based on identified indoor sources, studies using exposure surrogates or indoor concentrations alone are not enough to fill the gap of knowledge that still exists about the potential human health impact of indoor-generated  $PM_{2.5}$ . Therefore, more research is needed to establish the specific degree and type of PM toxicity from indoor sources in epidemiological studies, especially applying larger sample sizes and involving different populations. Source partitioning methods using trace elements from outdoor sources represent a reliable way to estimate a subject's exposure to indoor- and outdoor-generated PM<sub>2.5</sub>, with a reduction of exposure measurement errors.

Authors	Year	Type of study	Country	Population	Sample size	Outcomes
Balmes et al.	2014	Cross-sectional	USA	Adults with asthma and rhinitis	302	Respiratory symptoms and lung function (FEV1)
Chi et al.	2019	Panelstudy	China	COPD patients and healthy spouses	75	Resting BP, PEF and FEV1
Delfino et al.	2004	Panelstudy	USA	Children with asthma	19	Lung function (FEV1)
Ebelt et al.	2005	Panelstudy	Canada	Patients with COPD	16	Lung function (FEV1, change in FEV1), systolic and diastolic BP, SVE, HR, HR variability.
Habre et all.	2014	Panelstudy	USA	Children with asthma	36	Daily asthma caught and wheeze scores
Hartog et al.	2009	Panelstudy	Finland, Greece, Netherlands and UK	Adult patients with asthma or COPD	135	Lung function (FVC, FEV1, PEF)
Isiugo et al.	2019	Panelstudy	USA	Children with asthma	44	Lung function (FEV1, FVC, FEV1/FVC, FEF)
Jansen et al.	2005	Panelstudy	USA	Adult patients with asthma or COPD	16	$FE_{NO},lungfunction(FEV1),BP,SaO_2,andpulserate.$
Karottki et al.	2014	Cross-sectional study	Denmark	Adults	78	MVF, lung function, biomarkers related to inflammation monocyte activation and the prediabetic marker HbA1c
Karottki et al.	2015	Panelstudy	Denmark	Elderly adults	48	MVF, lung function, biomarkers related to inflammation monocyte activation and the prediabetic marker HbA1c
Kim et al.	2020	Randomized intervention study	Korea	Children with asthma	26	Daily PEFR
Koenig et al.	2005	Panelstudy	USA	19 asthmatic children	19	eNO, and lung function (FEV1, FVC, MEF)
Ma et al.	2008	Panelstudy	Japan	Hospitalized children with asthma	19	PEF and wheezing
Maesano et al.	2019	Cross-sectional study	France	Adults (farmers)	109	Asthma, COPD, early airway obstruction
Osman et al.	2007	Cross-sectional study	UK	Patients with COPD	148	Respiratory-specific health status (St. George's Respiratory Health Questionnaire)
Simoni et al.	2002	Cross-sectional study	Italy	Adults	383	Asthmatic symptoms and PEF variability
Simoni et al.	2004	Cross-sectional study	Italy	Adults	1091	Acute respiratory illness, asthmatic symptoms, irritant symptoms, PEF variability
Trenga et al.	2006	Panelstudy	USA	Children with asthma and elderly adults	74	Lung function (FEV1 and PEF in adults, MMEF, PEF, FEV1, and symptoms in children),
Weichenthal et al.	2012	Randomized intervention study	Canada	Adults	37	Lung function, BP, and endothelial function
Wilson and Brauer	2006	Panelstudy	Canada	Patients with COPD	16	Lung function (FEV1, change in FEV1), systolic and diastolic BP, SVE, HR, HR variability

Table 1. General characteristics of the reviewed epidemiological studies assessing short-term exposure to indoor  $PM_{2.5}$ 

 $COPD=chronic obstructive pulmonary disease; FEV1=forced expiratory volume in the first second; FVC=forced vital capacity; PEF=peak expiratory flow; FEF=forced expiratory flow; MEF=maximal expiratory flow; MMEF=maximal midexpiratory flow; FE_{NO}=fractional exhaled nitric oxide; SaO_2=oxygen saturation; BP=Blood pressure; MVF=microvascular function; eNO=exhaled nitric oxide; HR=Heart rate.$ 

Authors	Year	Type of study	Country	Population	Sample size	Follow-up	Outcomes
Butz et al.	2011	Randomized intervention study	USA	Children with asthma	126	6 months	Asthma symptom-free days (difference between baseline and follow-up)
Gurley et al.	2013	Birth cohort study	Bangladesh	Children	257	2 years	Incidence of acute lower respiratory infection (ALRI)
Hansel et all.	2013	Longitudinal cohort study	USA	Patients with COPD	84	6 months	Respiratory symptoms, rescue medication use, and COPD exacerbations
McCormack et al.	2009	Longitudinal cohort study	USA	Children with asthma	150	6 months	Respiratory symptoms, rescue medication use, acute health care use
McCormack et al.	2011	Longitudinal cohort study	USA	Children with asthma	150	6 months	Respiratory symptoms, rescue medication use, acute health care use
Neas et al.	1994	Longitudinal cohort study	USA	Children	1237	2 years	Cumulative incidence of respiratory symptoms, lung function (FVC, FEV1, FEV1/FVC, FEF25-75%, FEF25- 75%/FVC)
Peng et al.	2018	Randomized intervention study	USA	Children with asthma	75	6 months	Asthma symptom-free days (difference between baseline and follow-up)
Raaschou-Nielsen et al.	2010	Birth cohort study	Denmark	Children	378	18 months	Number of days with wheezing symptoms
Walker et al.	2022	Randomized intervention study	USA	Children	461	1 years (2 winter seasons)	LRTI

Table 2. General characteristics of the reviewed epidemiological studies assessing long-term exposure to indoor PM<sub>2.5</sub>

FEV1=forced expiratory volume in the first second; FVC=forced vital capacity; FEF25-75%=forced mid-expiratory flow at 25-75% of FVC; BP=Blood pressure; SVE=supraventricular ectopy; HR=Heart rate; LRTI=lower respiratory tract infections

Authors	Year	Exposure measurements	Instrument	Measurement period	Measurement season	Source partitioning	Time-activity data	Restriction of indoor sources
Balmes et al.	2014	Indoor-residence (kitchen, living room), outdoor-residence and central station	Nephelometer (DustTrak)	1 monitoring session of 3 min	Spring and summer	No	No	No, but hair nicotine was used as a covariate in multi-exposure models.
Chi et al.	2019	Indoor-residence (living room) and outdoor-residence	SKC sampling systems with Teflon filter	1 monitoring session of 5 days	Winter and spring	Yes, iron-tracer method	No	Yes, not smoking was an inclusion criterion for healthy spouses. Smoking status was used as a covariate.
Delfino et al.	2004	PEM, indoor-residence (living room), outdoor-residence and central station	Passive nephelometer (Personal dataRAM) and Harvard Impactors with Teflon filters	1 monitoring session of 2 weeks	Spring and fall	No	Yes (amount of time spent outdoors/indoors)	Yes, inclusion criteria included no active smoking or passive exposure to tobacco smoke at home.
Ebelt et al.	2005	PEM and concentrations at 5 central site stations	Harvard Impactors	7 monitoring sessions of 24 hours	Spring and summer	Yes, sulphate- tracer method	Yes (amount of time spent outdoors/indoors)	Yes, inclusion only of non-smokers living in non-smoking households.
Habre et all.	2014	Concentrations at indoor- residence (living room) and central site station	Nephelometer (DustTrak) with Teflon filters (indoor), Modified Harvard Impactors (outdoor)	2 monitoring sessions of 2 weeks	All year	Yes, sulphate- tracer method	No	Yes, exclusion criteria for the presence of a smoker in the residence.
Hartog et al.	2009	Concentrations at indoor- residence (living room), outdoor- residence and central cite station	Harvard Impactors	1 monitoring session of 1 week	All year	No	No	Yes, selection of non-smokers living in non-smoking households.
lsiugo et al.	2019	Concentrations at indoor- residence (bedrooms) and outdoor-residence	Teflon filters with a single-stage Personal Modular Impactor (SKC)	2 monitoring sessions of 48 hours	All year	No	No	No, but UVPM was measured (PM2.5 emitted from smouldering organics such as smoking cigarettes and burning fireplace wood)
Jansen et al.	2005	PEM and concentrations at indoor-residence (living room), outdoor-residence and central cite station.	MPEM <sub>10</sub> (personal exposure) and Harvard Impactors	1 monitoring session of 12 days	Winter	No	Yes (amount of time spent outdoors/indoors, household behaviours)	Yes, inclusion only of non-smokers.
Karottki et al.	2014	Concentrations at indoor- residence (living room) and at a central cite station.	Fluoropore Membrane PTFE filters	1 monitoring session of 2 days	Winter	No	Yes (household behaviours)	Yes, inclusion only of non-smokers.

Table 3. Exposure assessment m	ethods described	in epidemiological	l studies of short-terr	n exposure to indoc	$r PM_{2.5}$
•					2.0

PEM=personal exposure monitor; MPEM<sub>10</sub> = Marple Personal Environmental Monitors for PM10; PTFE=Polytetrafluoroethylene; TEOM=Tapered-element oscillating microbalance, UVPM=ultraviolet absorbing particulate matter

Authors	Year	Exposure measurements	Instrument	Measurement period	Measurement season	Source partitioning	Time-activity data	Restriction of indoor sources
Kim et al.	2020	Concentrations at indoor- residence (living room)	PurpleAir light scattering sensor	1 monitoring session of 7 weeks	Fall	No	No	No
Koenig et al.	2005	PEM and concentrations at indoor-residence (living room) and outdoor-residence.	Harvard Impactors, Radiance nephelometers, and Harvard personal environmental monitors	1 monitoring session of 10 days	Winter and spring	Yes, sulphate- tracer method and recursive mass balance model	Yes (amount of time spent outdoors/indoors)	No
Ma et al.	2008	Concentrations at indoor-hospita (2 rooms and a hall), outdoor- hospital (entrance), and central cite station near hospital.	TEOM (central cite), dust monitor with a laser diode light scattering device (indoor and outdoor).	7 monitoring sessions of 24 hours	Winter	No	No	No, but the study only included children under long-term hospitalization.
Maesano et al.	2019	Concentrations at farm-residence (living room/bedroom) and farm- workplace (granary/stable)	AEROCET 531S device	1 monitoring session of 2 min	Spring	No	No	No, but smoking status, presence of pets, moulds, and wood heating were included as covariates.
Osman et al.	2007	Concentrations at indoor- residence (living room) and at a central cite station.	DustTrak light scattering monitor	1 monitoring session of 12 hours	Winter and spring	No	No	No, but smoking status was included as a covariate.
Simoni et al.	2002	Concentrations at indoor- residence (living room)	Dorr Oliver-type preselector	2 monitoring sessions of 1 week	Winter and summer	No	Yes (amount of time spent outdoors/indoors)	No, but stratified analysis by smoking status.
Simoni et al.	2004	Concentrations at indoor- residence (living room)	Dorr Oliver-type preselector	3 monitoring sessions of 1- week	Winter and summer	No	Yes (amount of time spent outdoors/indoors)	No, but smoking status was included as a covariate
Trenga et al.	2006	PEM and concentrations at indoor-residence (living room), outdoor-residence and central cite station.	Harvard Impactors (indoor and outdoor-residence), Harvard Personal Environmental Monitor	26 monitoring sessions of 5-10 days	All year	No	No	Yes, inclusion only of non-smokers.
Weichenthal et al.	2012	Concentrations at indoor- residence (living room) and at a central cite station.	Harvard cascade impactors	1 monitoring session of 3 weeks	Winter	No	No	No, but daily average number of cigarettes smoked indoors was included as covariate.
Wilson and Brauer	2006	PEM and concentrations at 5 central site stations	Harvard Impactors	7 monitoring sessions of 24 hours	Spring and summer	Yes, sulphate- tracer method	Yes (amount of time spent outdoors/indoors)	Yes, inclusion only of non-smokers living in non-smoking households.

Table 3. Exposure assessment methods described in epidemiological studies of short-term exposure to indoor PM<sub>2.5</sub> (CONT.)

PEM=personal exposure monitor; MPEM<sub>10</sub> = Marple Personal Environmental Monitors for PM10; PTFE=Polytetrafluoroethylene; TEOM=Tapered-element oscillating microbalance, UVPM=ultraviolet absorbing particulate matter

Authors	Year	Exposure measurements	Instrument	Measurement period	Measurement season	Source partitionin g	Time-activity data	Restriction of indoor sources
Butz et al.	2011	Indoor-residence (living room and bedroom)	MSP impactors (MSP Corp.)	2 monitoring sessions of 7 days	All year	No	Yes (amount of time spent outdoors/indoors)	Yes, inclusion of only children residing with a smoker.
Gurley et al.	2013	Indoor-residence (bedroom)	Berkeley Air monitoring device	1 monitoring session per month of 24 hours	All year	No	No	No, but study was conducted in a low-income urban community.
Hansel et all.	2013	Indoor-residence (bedroom, living room)	MSP impactors (MSP Corp.)	3 monitoring sessions of 1- week	All year	No	Yes (household behaviours)	Yes, inclusion of former smokers only. Assessment of SHS exposure by air/hair nicotine (confounding variable).
McCormack et al.	2009	Indoor-residence (bedroom) and central site station	MSP impactors (indoor) and PartisolPlus sequential air sampler (outdoor)	3 monitoring sessions of 3 consecutive days	All year	No	Yes (household behaviours)	No, but ambient PM was included as a covariate.
McCormack et al.	2011	Indoor-residence (bedroom) and central site station	MSP impactors (indoor) and PartisolPlus sequential air sampler (outdoor)	3 monitoring sessions of 3 consecutive days	All year	No	Yes (amount of time spent outdoors/indoors, household behaviours)	No, but ambient PM was included as a covariate.
Neas et al.	1994	Indoor-residence (bedroom)	Harvard Impactors	2 monitoring sessions of 1- week	Winter and summer	No	No	Yes, inclusion of children residing in never-smoking households
Peng et al.	2018	Indoor-residence (living room and bedroom)	MSP impactors (MSP Corp.)	2 monitoring sessions of 7 days	All year	No	No	Yes, inclusion of only children residing with a smoker.
Raaschou-Nielsen et al.	2010	Indoor-residence (living room)	KTL PM2.5 cyclone and a BGI400 pump	3 monitoring sessions of 1- week	All year	No	No	No
Walker et al.	2022	Indoor-residence (kitchen)	DustTrak light-scattering monitor	1 monitoring session of 6 days	Winter	No	Yes (household behaviours)	Yes, inclusion only of children residing in households that use a wood stove as primary heating source.

Table 4. Exposure assessment methods described in epidemiological studies of long-term exposure to indoor  $PM_{2.5}$ 

SHS=second hand smoking

Study	Exposure	Health outcome	Type of effect estimate	Effect estimate	95% CI
	Indoor-generated PM2.5	FEV1	Percentage deviation for IQR increase	-0.5%	(-0.9%, -0.1%)
Chi et al.	Outdoor-generated PM2.5	PEF	Percentage deviation for IQR increase	-3.7%	(-7.1%, -0.3%)
	PM2.5 indoor concentration	FEV1	Percentage deviation for IQR increase	-1.7%	(-2.9%, -0.5%)
	Ambient DM2 5	FEV <sub>1</sub> (ml)	Effect estimates for IQR increase	-14.42	(-40.22, 11.38)
Vancouver Panel Study (Ebelt et al., Wilson and Brauer)	Ambient PMZ.5	ΔFEV₁(ml)	Effect estimates for IQR increase	-28.93	(-54.55, -3.30)
		FEV <sub>1</sub> (ml)	FEV <sub>1</sub> (ml) Effect estimates for IQR increase		(0.21, 21.73)
	Nonamplent PM2.5	$\Delta FEV_1$ (ml)	Effect estimates for IQR increase	4.01	(-6.32, 14.33)
	Indeer generated DM2.5	Asthma caught symptoms	OR for a SD increase	1.2	(0.88, 1.64)
	Indoor-generated PMZ.5	Asthma wheeze symptoms	OR for a SD increase	1.55	(1.05, 2.28)
Liebre et el	Outdoor concreted DM2.5	Asthma caught symptoms	OR for a SD increase	1.27	(0.90, 1.77)
Habre et al.	Outdoor-generated PM2.5	Asthma wheeze symptoms	OR for a SD increase	1.13	(0.75, 1.72)
		Asthma caught symptoms	OR for a SD increase	1.22	(0.91, 1.63)
	PMZ.5 Indoor concentration	Asthma wheeze symptoms	OR for a SD increase	1.57	(1.09, 2.26)
Keening et al	Indoor-generated PM2.5	eNO	Change per 10 µg/m <sup>3</sup> estimated PM2.5	No use of ICS: 3.29 Use of ICS: -4.94	No use of ICS : (-1.14, 7.73) Use of ICS : (-10.94, 1.06)
Kuching et al.	Outdoor-generated PM2.5	eNO	Change per 10 µg/m <sup>3</sup> estimated PM2.5	No use of ICS: 4.98; Use of ICS: -0.19	No use of ICS : (0.28, 9.69) Use of ICS : (-3.77, 7.12)

Table 5. Effect estimates of	oserved in epidemiolo	gical studies of respira	atory health effects usin	g source partitioning
				<u> </u>

FEV1= forced expiratory volume in the first second; PEF=peak expiratory flow;  $\Delta$ FEV<sub>1</sub> = change in FEV1; eNo= Exhaled nitric oxide; IQR=interquartile range; SD=standard deviation; ICS: inhaled corticosteroid

#### References

- 1. U.S. EPA. Integrated Science Assessment (ISA) for Particulate Matter. Washington, DC: U.S. Environmental Protection Agency; 2019. 1967 p. (EPA/600/R-19/188).
- 2. World Health Organization. WHO global air quality guidelines: particulate matter (PM2.5 and PM10), ozone, nitrogen dioxide, sulfur dioxide and carbon monoxide [Internet]. Geneva: World Health Organization; 2021 [cited 2022 Jun 21]. Available from: https://apps.who.int/iris/handle/10665/345329
- 3. Klepeis NE, Nelson WC, Ott WR, Robinson JP, Tsang AM, Switzer P, et al. The National Human Activity Pattern Survey (NHAPS): a resource for assessing exposure to environmental pollutants. J Expo Sci Environ Epidemiol. 2001 Jul 1;11(3):231–52.
- 4. Li N, Friedrich R. Methodology for Estimating the Lifelong Exposure to PM2.5 and NO2—The Application to European Population Subgroups. Atmosphere. 2019 Sep;10(9):507.
- 5. Logue JM, Price PN, Sherman MH, Singer BC. A method to estimate the chronic health impact of air pollutants in U.S. residences. Environ Health Perspect. 2012 Feb;120(2):216–22.
- 6. Morawska L, Afshari A, Bae GN, Buonanno G, Chao CYH, Hänninen O, et al. Indoor aerosols: from personal exposure to risk assessment. Indoor Air. 2013;23(6):462–87.
- Long CM, Suh HH, Kobzik L, Catalano PJ, Ning Y, Koutrakis P. A Pilot Investigation of the Relative Toxicity of Indoor and Outdoor Fine Particles: In Vitro Effects of Endotoxin and Other Particulate Properties. Environ Health Perspect. 2001;109(10):8.
- 8. Wilson WE, Mage DT, Grant LD. Estimating separately personal exposure to ambient and nonambient particulate matter for epidemiology and risk assessment: why and how. J Air Waste Manag Assoc 1995. 2000 Jul;50(7):1167–83.
- 9. Spengler JD, Treitman RD, Tosteson TD, Mage DT, Soczek MLou. Personal exposures to respirable particulates and implications for air pollution epidemiology. Environ Sci Technol. 1985 Aug;19(8):700–7.
- 10. Clayton CA, Perritt RL, Pellizzari ED, Thomas KW, Whitmore RW, Wallace LA, et al. Particle Total Exposure Assessment Methodology (PTEAM) study: distributions of aerosol and elemental concentrations in personal, indoor, and outdoor air samples in a southern California community. J Expo Anal Environ Epidemiol. 1993 Jun;3(2):227–50.

- U.S. EPA. Air Quality Criteria for Particulate Matter (Final Report, 2004) [Internet]. Washington, D.C: U.S. Environmental Protection Agency; [cited 2022 Jun 21]. Report No.: EPA 600/P-95/001. Available from: https://cfpub.epa.gov/ncea/risk/recordisplay.cfm?deid=87903
- 12. Mohammed MOA, Song WW, Ma WL, Li WL, Ambuchi JJ, Thabit M, et al. Trends in indoor–outdoor PM2.5 research: A systematic review of studies conducted during the last decade (2003–2013). Atmospheric Pollut Res. 2015 Sep;6(5):893–903.
- 13. Li Z, Wen Q, Zhang R. Sources, health effects and control strategies of indoor fine particulate matter (PM2.5): A review. Sci Total Environ. 2017 May 15;586:610–22.
- 14. Loo CKJ, Foty RG, Wheeler AJ, Miller JD, Evans G, Stieb DM, et al. Do Questions Reflecting Indoor Air Pollutant Exposure from a Questionnaire Predict Direct Measure of Exposure in Owner-Occupied Houses? Int J Environ Res Public Health. 2010 Aug;7(8):3270–97.
- 15. Jantunen M, Jaakkola JJK, Krzyżanowski M, editors. Assessment of exposure to indoor air pollutants. Copenhagen: World Health Organization Regional office for Europe; 1997. 139 p. (WHO regional publications).
- 16. Nieuwenhuijsen MJ, editor. Exposure assessment in environmental epidemiology. Second edition. Oxford ; New York: Oxford University Press; 2015. 405 p.
- 17. Hoek G, Kos G, Harrison R, de Hartog J, Meliefste K, ten Brink H, et al. Indooroutdoor relationships of particle number and mass in four European cities. Atmos Environ. 2008 Jan;42(1):156–69.
- 18. Abt E, Suh HH, Catalano P, Koutrakis P. Relative Contribution of Outdoor and Indoor Particle Sources to Indoor Concentrations. Environ Sci Technol. 2000 Sep 1;34(17):3579–87.
- 19. Chu H, Huang FQ, Yuan Q, Fan Y, Xin J, Du M, et al. Metabolomics identifying biomarkers of PM2.5 exposure for vulnerable population: based on a prospective cohort study. Environ Sci Pollut Res Int. 2021 Mar;28(12):14586–96.
- Sørensen M, Autrup H, Hertel O, Wallin H, Knudsen LE, Loft S. Personal Exposure to PM2.5 and Biomarkers of DNA Damage1. Cancer Epidemiol Biomarkers Prev. 2003 Mar 1;12(3):191–6.
- Simoni M, Biavati P, Carrozzi L, Viegi G, Paoletti P, Matteucci G, et al. The Po River Delta (North Italy) Indoor Epidemiological Study: Home Characteristics, Indoor Pollutants, and Subjects' Daily Activity Pattern. Indoor Air. 1998;8(2):70–9.

- 22. Shen G, Xue M, Chen Y, Yang C, Li W, Shen H, et al. Comparison of carbonaceous particulate matter emission factors among different solid fuels burned in residential stoves. Atmos Environ. 2014 Jun;89:337–45.
- 23. Long CM, Suh HH, Koutrakis P. Characterization of indoor particle sources using continuous mass and size monitors. J Air Waste Manag Assoc 1995. 2000 Jul;50(7):1236–50.
- 24. Bhangar S, Mullen NA, Hering SV, Kreisberg NM, Nazaroff WW. Ultrafine particle concentrations and exposures in seven residences in northern California. Indoor Air. 2011;21(2):132–44.
- Morawska L, Salthammer T. Indoor Environment: Airborne Particles and Settled Dus. In: Morawska L, Salthammer T, editors. Indoor Environment [Internet]. Weinheim, Germany: Wiley-VCH Verlag GmbH & Co. KGaA; 2003 [cited 2022 Jul 14]. p. 1–46. Available from: https://onlinelibrary.wiley.com/doi/10.1002/9783527610013.ch1
- 26. See SW, Balasubramanian R. Characterization of fine particle emissions from incense burning. Build Environ. 2011 May;46(5):1074–80.
- 27. Martins NR, Carrilho da Graça G. Impact of PM2.5 in indoor urban environments: A review. Sustain Cities Soc. 2018 Oct 1;42:259–75.
- 28. Quang TN, He C, Morawska L, Knibbs LD. Influence of ventilation and filtration on indoor particle concentrations in urban office buildings. Atmos Environ. 2013 Nov 1;79:41–52.
- 29. Wang X, Bi X, Chen D, Sheng G, Fu J. Hospital indoor respirable particles and carbonaceous composition. Build Environ. 2006 Aug 1;41(8):992–1000.
- 30. Stabile L, Fuoco FC, Buonanno G. Characteristics of particles and black carbon emitted by combustion of incenses, candles and anti-mosquito products. Build Environ. 2012 Oct 1;56:184–91.
- 31. Adgate JL, Ramachandran G, Pratt GC, Waller LA, Sexton K. Spatial and temporal variability in outdoor, indoor, and personal PM2.5 exposure. Atmos Environ. 2002 Jul 1;36(20):3255–65.
- 32. Hänninen OO, Lebret E, Ilacqua V, Katsouyanni K, Künzli N, Srám RJ, et al. Infiltration of ambient PM2.5 and levels of indoor generated non-ETS PM2.5 in residences of four European cities. Atmos Environ. 2004 Dec 1;38(37):6411–23.
- 33. Penney D, Benignus V, Kephalopoulos S, Kotzias D, Kleinman M, Verrier A. Carbon monoxide [Internet]. WHO Guidelines for Indoor Air Quality: Selected Pollutants.

World Health Organization; 2010 [cited 2022 Jul 12]. Available from: https://www.ncbi.nlm.nih.gov/books/NBK138710/

- 34. U.S. EPA. Quantitative Risk and Exposure Assessment for Carbon Monoxide. 2010 Jul p. 376. Report No.: EPA-452/R-10-009.
- 35. Zeger SL, Thomas D, Dominici F, Samet JM, Schwartz J, Dockery D, et al. Exposure measurement error in time-series studies of air pollution: concepts and consequences. Environ Health Perspect. 2000 May;108(5):419–26.
- 36. Buonanno G, Morawska L, Stabile L. Particle emission factors during cooking activities. Atmos Environ. 2009 Jun;43(20):3235–42.
- Ma L, Shima M, Yoda Y, Yamamoto H, Nakai S, Tamura K, et al. Effects of Airborne Particulate Matter on Respiratory Morbidity in Asthmatic Children. J Epidemiol. 2008;18(3):97–110.
- Ebelt ST, Wilson WE, Brauer M. Exposure to Ambient and Nonambient Components of Particulate Matter: A Comparison of Health Effects. Epidemiology. 2005 May;16(3):396–405.
- 39. Wilson WE, Brauer M. Estimation of ambient and non-ambient components of particulate matter exposure from a personal monitoring panel study. J Expo Sci Environ Epidemiol. 2006 May;16(3):264–74.
- 40. Koenig JQ, Mar TF, Allen RW, Jansen K, Lumley T, Sullivan JH, et al. Pulmonary Effects of Indoor- and Outdoor-Generated Particles in Children with Asthma. Environ Health Perspect. 2005 Apr;113(4):499–503.
- 41. Habre R, Moshier E, Castro W, Nath A, Grunin A, Rohr A, et al. The effects of PM2.5 and its components from indoor and outdoor sources on cough and wheeze symptoms in asthmatic children. J Expo Sci Environ Epidemiol. 2014 Jul;24(4):380–7.
- 42. Chi R, Chen C, Li H, Pan L, Zhao B, Deng F, et al. Different health effects of indoorand outdoor-originated PM2.5 on cardiopulmonary function in COPD patients and healthy elderly adults. Indoor Air. 2019;29(2):192–201.
- 43. Sarnat JA, Long CM, Koutrakis P, Coull BA, Schwartz J, Suh HH. Using Sulfur as a Tracer of Outdoor Fine Particulate Matter. Environ Sci Technol. 2002 Dec 1;36(24):5305–14.
- 44. Oglesby L, Künzli N, Röösli M, Braun-Fahrländer C, Mathys P, Stern W, et al. Validity of Ambient Levels of Fine Particles as Surrogate for Personal Exposure to Outdoor Air Pollution—Results of the European EXPOLIS-EAS Study (Swiss Center Basel). J Air Waste Manag Assoc. 2000 Jul 1;50(7):1251–61.

- 45. Allen R, Wallace L, Larson T, Sheppard L, Liu LJS. Estimated hourly personal exposures to ambient and nonambient particulate matter among sensitive populations in Seattle, Washington. J Air Waste Manag Assoc 1995. 2004 Sep;54(9):1197–211.
- 46. Ji W, Li H, Zhao B, Deng F. Tracer element for indoor PM2.5 in China migrated from outdoor. Atmos Environ. 2018 Mar;176:171–8.
- 47. Button KS, Ioannidis JPA, Mokrysz C, Nosek BA, Flint J, Robinson ESJ, et al. Power failure: why small sample size undermines the reliability of neuroscience. Nat Rev Neurosci. 2013 May;14(5):365–76.
- 48. Forstmeier W, Wagenmakers EJ, Parker TH. Detecting and avoiding likely falsepositive findings – a practical guide. Biol Rev. 2017;92(4):1941–68.
- 49. Wallace L. Indoor Particles: A Review. J Air Waste Manag Assoc. 1996 Feb;46(2):98–126.
- 50. Butz AM, Matsui EC, Breysse P, Curtin-Brosnan J, Eggleston P, Diette G, et al. A randomized trial of air cleaners and a health coach to improve indoor air quality for inner-city children with asthma and secondhand smoke exposure. Arch Pediatr Adolesc Med. 2011 Aug;165(8):741–8.
- 51. Delfino RJ, Quintana PJE, Floro J, Gastañaga VM, Samimi BS, Kleinman MT, et al. Association of FEV1 in asthmatic children with personal and microenvironmental exposure to airborne particulate matter. Environ Health Perspect. 2004 Jun;112(8):932–41.
- Gurley ES, Homaira N, Salje H, Ram PK, Haque R, Petri W, et al. Indoor exposure to particulate matter and the incidence of acute lower respiratory infections among children: A birth cohort study in urban Bangladesh. Indoor Air. 2013;23(5):379– 86.
- 53. Hansel NN, McCormack MC, Belli AJ, Matsui EC, Peng RD, Aloe C, et al. In-Home Air Pollution Is Linked to Respiratory Morbidity in Former Smokers with Chronic Obstructive Pulmonary Disease. Am J Respir Crit Care Med. 2013 May 15;187(10):1085–90.
- 54. Hartog JJ de, Ayres JG, Karakatsani A, Analitis A, Brink H ten, Hameri K, et al. Lung function and indicators of exposure to indoor and outdoor particulate matter among asthma and COPD patients. Occup Environ Med. 2010 Jan 1;67(1):2–10.
- 55. Isiugo K, Jandarov R, Cox J, Ryan P, Newman N, Grinshpun SA, et al. Indoor particulate matter and lung function in children. Sci Total Environ. 2019 May;663:408–17.

- 56. Jansen KL, Larson TV, Koenig JQ, Mar TF, Fields C, Stewart J, et al. Associations between Health Effects and Particulate Matter and Black Carbon in Subjects with Respiratory Disease. Environ Health Perspect. 2005 Dec;113(12):1741–6.
- 57. Karottki DG, Bekö G, Clausen G, Madsen AM, Andersen ZJ, Massling A, et al. Cardiovascular and lung function in relation to outdoor and indoor exposure to fine and ultrafine particulate matter in middle-aged subjects. Environ Int. 2014 Dec;73:372–81.
- 58. Karottki DG, Spilak M, Frederiksen M, Jovanovic Andersen Z, Madsen AM, Ketzel M, et al. Indoor and outdoor exposure to ultrafine, fine and microbiologically derived particulate matter related to cardiovascular and respiratory effects in a panel of elderly urban citizens. Int J Environ Res Public Health. 2015 Feb 2;12(2):1667–86.
- 59. Neas LM, Dockery DW, Ware JH, Spengler JD, Ferris BG Jr, Speizer FE. Concentration of Indoor Particulate Matter as a Determinant of Respiratory Health in Children. Am J Epidemiol. 1994 Jun 1;139(11):1088–99.
- 60. Peng RD, Butz AM, Hackstadt AJ, Williams DL, Diette GB, Breysse PN, et al. Estimating the health benefit of reducing indoor air pollution in a randomized environmental intervention. J R Stat Soc Ser A Stat Soc. 2015 Feb;178(2):425–43.
- 61. Trenga CA, Sullivan JH, Schildcrout JS, Shepherd KP, Shapiro GG, Liu LJS, et al. Effect of particulate air pollution on lung function in adult and pediatric subjects in a Seattle panel study. Chest. 2006 Jun;129(6):1614–22.
- 62. Walker ES, Semmens EO, Belcourt A, Boyer BB, Erdei E, Graham J, et al. Efficacy of Air Filtration and Education Interventions on Indoor Fine Particulate Matter and Child Lower Respiratory Tract Infections among Rural U.S. Homes Heated with Wood Stoves: Results from the KidsAIR Randomized Trial. Environ Health Perspect. 130(4):047002.
- 63. Balmes JR, Cisternas M, Quinlan PJ, Trupin L, Lurmann FW, Katz PP, et al. Annual average ambient particulate matter exposure estimates, measured home particulate matter, and hair nicotine are associated with respiratory outcomes in adults with asthma. Environ Res. 2014 Feb;129:1–10.
- 64. Maesano CN, Caillaud D, Youssouf H, Banerjee S, Prud'Homme J, Audi C, et al. Indoor exposure to particulate matter and volatile organic compounds in dwellings and workplaces and respiratory health in French farmers. Multidiscip Respir Med. 2019 Dec;14(1):33.
- 65. McCormack MC, Breysse PN, Matsui EC, Hansel NN, Williams D, Curtin-Brosnan J, et al. In-Home Particle Concentrations and Childhood Asthma Morbidity. Environ Health Perspect. 2009 Feb;117(2):294–8.

- 66. McCormack MC, Breysse PN, Matsui EC, Hansel NN, Peng RD, Curtin-Brosnan J, et al. Indoor particulate matter increases asthma morbidity in children with nonatopic and atopic asthma. Ann Allergy Asthma Immunol. 2011 Apr;106(4):308–15.
- 67. Osman LM, Douglas JG, Garden C, Reglitz K, Lyon J, Gordon S, et al. Indoor Air Quality in Homes of Patients with Chronic Obstructive Pulmonary Disease. Am J Respir Crit Care Med. 2007 Sep;176(5):465–72.
- 68. Simoni M, Carrozzi L, Baldacci S, Scognamiglio A, Di Pede F, Sapigni T, et al. The Po River Delta (north Italy) indoor epidemiological study: effects of pollutant exposure on acute respiratory symptoms and respiratory function in adults. Arch Environ Health. 2002 Apr;57(2):130–6.
- 69. Simoni M, Scognamiglio A, Carrozzi L, Baldacci S, Angino A, Pistelli F, et al. Indoor exposures and acute respiratory effects in two general population samples from a rural and an urban area in Italy. J Expo Anal Environ Epidemiol. 2004;14 Suppl 1:S144-152.
- 70. Weichenthal S, Mallach G, Kulka R, Black A, Wheeler A, You H, et al. A randomized double-blind crossover study of indoor air filtration and acute changes in cardiorespiratory health in a First Nations community. Indoor Air. 2013 Jun;23(3):175–84.
- 71. Kim S, Lee J, Park S, Rudasingwa G, Lee S, Yu S, et al. Association between Peak Expiratory Flow Rate and Exposure Level to Indoor PM2.5 in Asthmatic Children, Using Data from the Escort Intervention Study. Int J Environ Res Public Health. 2020 Oct 21;17(20):E7667.
- 72. Raaschou-Nielsen O, Hermansen MN, Loland L, Buchvald F, Pipper CB, Sørensen M, et al. Long-term exposure to indoor air pollution and wheezing symptoms in infants. Indoor Air. 2010;20(2):159–67.

#### Supplement 1. Exposure indicator epidemiological studies

There is a large body of literature reporting health effects from indoor sources based on exposure questionnaire indicators for smoking, second-hand tobacco smoke (SHS), fuelburning while cooking, or other in-home combustion process; although biomarkers have also been applied to assign exposure (e.g., nicotine in hair or urine for second-hand tobacco smoke) [1–6]. The use of exposure surrogates has the advantage of allowing larger study samples and greater statistical power, as well as longer follow-up periods for long-term effects. This is because exposure surrogates have greater accessibility and affordability, are simpler to administer, more cost-effective, and warrant a lower participant burden compared to quantitative methods. However, if the aim is to determine the health effects of PM<sub>2.5</sub> from indoor sources, exposure indicators entail certain disadvantages and risks for an accurate evaluation of exposure.

First, exposure indicators are not specific to a certain type of pollutant. While it is well established that smoking, cooking, and candle/incense burning are some of the largest sources of indoor  $PM_{2.5}$  [7–9], these sources often produce a complex mixture of pollutants that can act separately or synergistically to affect human health. The use of exposure surrogates may lead to the attribution of health effects to PM<sub>2.5</sub> that are actually produced by other particles or compounds, with an under- or overestimation of the effect estimates (confounding bias). For example, carbon monoxide (CO) is another highly toxic pollutant present in the gas phase of tobacco smoke [10] and its chronic inhalation at doses corresponding to SHS has been associated with effects on the neurological, respiratory, and cardiovascular systems [11,12]. The evaluation of a mixture of pollutants can also be an advantage, depending on the study question to be analysed. Second, questionnaire-based indicators are more prone to exposure misclassification and recall bias. In general, self-reported exposure tends to underestimate real exposure, and correlations between questionnaire-based exposure indicators and quantitative exposure measurements can be highly variable and dependent on the exposure definition and time since exposure [13]. Even if the misclassification of exposure is random and not systematic, when the magnitude of the actual relationship between exposure and the health effect is small, inaccurate exposure assessment can lead to a statistical error type 2, in which the conclusion would be that such a relationship does not exist when in fact it does. Finally, there are indoor sources of PM<sub>2.5</sub> that simply cannot be determined using exposure indicators and that contribute to the potential risk to human health from indoor PM<sub>2.5</sub>. Examples of this are the fine particles that come from resuspension by human indoor activities and whose exposure can only be estimated using modelling methods [14,15].

- 1. Hystad P, Duong M, Brauer M, Larkin A, Arku R, Kurmi OP, et al. Health Effects of Household Solid Fuel Use: Findings from 11 Countries within the Prospective Urban and Rural Epidemiology Study. Environ Health Perspect. 127(5):057003.
- 2. Torres-Duque C, Maldonado D, Perez-Padilla R, Ezzati M, Viegi G, on behalf of the Forum of International Respiratory Societies (FIRS) Task Force on Health Effects of Biomass Exposure. Biomass Fuels and Respiratory Diseases: A Review of the Evidence. Proc Am Thorac Soc. 2008 Jul 15;5(5):577–90.
- Zhang X, Rao L, Liu Q, Yang Q. Meta-analysis of associations between cooking oil fumes exposure and lung cancer risk. Indoor Built Environ. 2022 Mar;31(3):820– 37.
- 4. Naeher LP, Brauer M, Lipsett M, Zelikoff JT, Simpson CD, Koenig JQ, et al. Woodsmoke Health Effects: A Review. Inhal Toxicol. 2007 Jan;19(1):67–106.
- 5. Dunbar A, Gotsis W, Frishman W. Second-Hand Tobacco Smoke and Cardiovascular Disease Risk: An Epidemiological Review. Cardiol Rev. 2013 Mar;21(2):94–100.
- 6. Pan A, Clark ML, Ang LW, Yu MC, Yuan JM, Koh WP. Incense use and cardiovascular mortality among Chinese in Singapore: the Singapore Chinese Health Study. Environ Health Perspect. 2014 Dec;122(12):1279–84.
- 7. Abt E, Suh HH, Catalano P, Koutrakis P. Relative Contribution of Outdoor and Indoor Particle Sources to Indoor Concentrations. Environ Sci Technol. 2000 Sep 1;34(17):3579–87.
- 8. Brauer M, Hirtle R, Lang B, Ott W. Assessment of indoor fine aerosol contributions from environmental tobacco smoke and cooking with a portable nephelometer. J Expo Sci Environ Epidemiol. 2000 Apr;10(2):136–44.
- 9. Long CM, Suh HH, Koutrakis P. Characterization of indoor particle sources using continuous mass and size monitors. J Air Waste Manag Assoc 1995. 2000 Jul;50(7):1236–50.
- Talhout R, Schulz T, Florek E, Van Benthem J, Wester P, Opperhuizen A. Hazardous Compounds in Tobacco Smoke. Int J Environ Res Public Health. 2011 Feb;8(2):613–28.
- 11. Sørhaug S, Steinshamn S, Nilsen OG, Waldum HL. Chronic inhalation of carbon monoxide: effects on the respiratory and cardiovascular system at doses corresponding to tobacco smoking. Toxicology. 2006 Dec 7;228(2–3):280–90.
- 12. Raub JA, Benignus VA. Carbon monoxide and the nervous system. Neurosci Biobehav Rev. 2002 Dec;26(8):925–40.

- 13. Avila-Tang E, Elf JL, Cummings KM, Fong GT, Hovell MF, Klein JD, et al. Assessing secondhand smoke exposure with reported measures. Tob Control. 2013 May;22(3):156–63.
- 14. Qian J, Peccia J, Ferro AR. Walking-induced particle resuspension in indoor environments. Atmos Environ. 2014 Jun;89:464–81.
- 15. Qian J, Ferro AR. Resuspension of Dust Particles in a Chamber and Associated Environmental Factors. Aerosol Sci Technol. 2008 May 29;42(7):566–78.