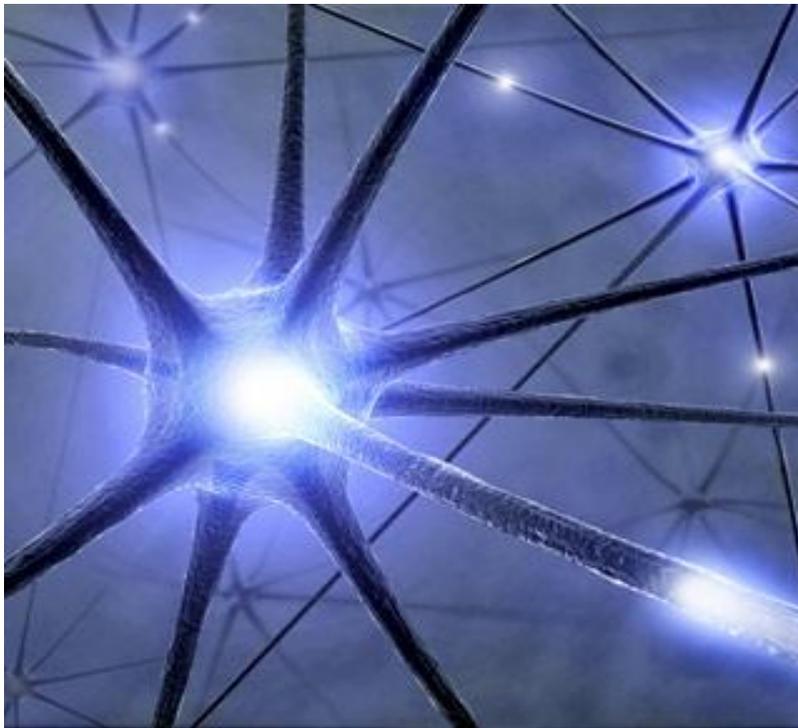


DISEASE MECHANISM AND MODELS FOR ALS

NEW CONCEPT: AGING AS MAJOR REGULATOR OF ALS PATHOLOGY

A REVIEW ON THE CURRENT KNOWLEDGE OF ALS IS GIVEN BY DISCUSSING DISEASE MECHANISMS AND MODELS THAT ARE USED TO COMBAT THE BATTLE AGAINST ITS PATHOLOGY. FURTHERMORE, A NEW CONCEPT WILL BE PROPOSED.



Susanne Elise Baars

December 2012

Image on front page:

Researchers suggest that stimulation of cell signaling delays the onset of ALS, by causing motor neurons 'humming'.

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Image reference: www.scientificamerican.com/article.cfm?id=potential-new-weapon-against-als

HUBRECHT INSTITUTE

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ABSTRACT

Amyotrophic lateral sclerosis (ALS, Lou Gehrig's disease) is a severe adult-onset neurodegenerative disease, characterized by progressive premature loss of upper and lower motor neurons. Eventually the disease leads to death due to respiratory failure¹. ALS is the third most common neurodegenerative disease² and cause of adult mortality³. It appears to be caused by a complex interaction among genetics and environmental factors². While approximately 10% of ALS cases are familial, the remaining cases are believed to be sporadic. However, the exact working mechanism underlying the selective motor neuron death in ALS is not resolved yet. Since a wide variety of cellular processes are involved in ALS, a general, systemic principle might underlie it.

Recent data provides compelling evidence for a major role of aging in the development of ALS^{4,5,6,7}. This hypothesis was enhanced by the discovery that reduced Insulin/IGF signalling (IIS) correlates with lifespan extension⁸ and a decreased risk to develop ALS⁹. Its pathway is not only regulating lifespan, metabolism and stress resistance⁴, TDP-43 has also showed to be part of it, a well known ALS gene⁹. TDP-43 is primarily a nuclear protein that participates in common heteromultimeric complexes, which is involved into diverse RNA processes and stress granule formation^{16,10}. Furthermore, TDP-43 is identified as the major component of insoluble cytoplasmic inclusions in both sALS as fALS, and associated with the oxidative stress response. It seems likely that this TDP-43 protein mediates longevity through activation of the IIS pathway under stressful conditions¹¹. Besides the activation of various cellular cascades, there is a vicious circle in which TDP-43 expression becomes upregulated. Interestingly, it has been found that during stress, the threshold for TDP-43 phosphorylation is lowered. This lead to increased proteotoxicity¹². This in turn seems to cause and/or to accelerate motor neuron degeneration.

However, besides aging, also heavy exercise is in part mediated by the IIS pathway and involved in ALS development¹³. During heavy exercise, enormous amount of ROS are generated which causes oxidative stress and disturb the epigenetic codes¹⁴.

Interestingly, age-related changes seemed to be determined by the age of the systemic environment, rather than by the cell-autonomous age¹⁵. Therefore, it has been suggested that signals from the systemic environment drive age-related, intrinsic changes, mediated by the IIS pathway. This activates a cascade of events that eventually can lead to ALS pathology. However, current animal models are not suitable to further investigate this postulation. Not only did the obtained results fail when translated to the human situation, also are fundamental genetic and anatomical differences between both species in age-related pathways found^{16,17}. Therefore, new and improved models are needed. In vitro cell based therapies seems to be the first feasible model to investigate the disease mechanism for both sALS and fALS by utilizing induced pluripotent stem cells (iPSCs)¹⁸. iPSCs can give us insights into disease mechanisms, drug discovery, cell therapy and a potential new diagnostic method for patients with ALS^{19,20,21}. Furthermore, will this technique enable the identification of a systemic regulation of aging on ALS pathology in patient derived iPSCs.

ABBREVIATIONS

ALS	Amyotrophic lateral sclerosis
MN	Motor Neurons
UMN	Upper Motor Neurons
LMN	Lower Motor Neurons
FTLD	frontotemporal dementia
sALS	Sporadic ALS
fALS	fALS
ER stress	endoplasmic reticulum stress
UPR	unfolded protein response
IGF1	insulin-like growth factor 1
TNF α	tumour necrosis factor
BBB	blood brain barrier
BSCB	blood spinal cord barrier
MP2	myelin protein 2
MMp9	matric metalloproteinase 9
iPSC	induced pluripotent stem cells
ESCs	Embryonic stem cells
Human iPSC	Human Induced Pluripotent Stem Cell

INTRODUCTION

Amyotrophic lateral sclerosis (ALS, Lou Gehrig's disease) is a severe adult-onset neurodegenerative disease, characterized by progressive premature loss of upper and lower motor neurons (MNs) in the motor cortex, the anterior horn of the brainstem and spinal cord which eventually leads to death due to respiratory failure¹. Next to being third most common neurodegenerative disease with a worldwide prevalence of 4-6 per 100.000²², ALS is also one of the most common causes of adult mortality in this disease group. The peak age of onset is between 50 and 70 years².

The broad variety of disease manifestations in ALS appears to be caused by a complex interaction among genetics and environmental factors. Approximately 10% of ALS cases are familial, while the remaining cases are believed to be sporadic²³. Although the enormous efforts put into unravelling the underlying disease mechanism, no efficacious drug or therapy has been developed yet²⁴. Numerous hypotheses have been proposed to underlie ALS, but none of them is able to fully explain the mechanism of pathogenesis.

AIM OF THIS THESIS

Within this thesis I will review the proposed disease mechanisms of ALS by discussing the current hypotheses and models that are used to investigate it. Furthermore, I will propose a new concept with respect to ALS pathology and outline a research proposal for future investigation.

ALS PATHOLOGY AND DIAGNOSIS

ALS pathology is characterized by loss of motor neurons in the anterior horns of the spinal cord, motor nuclei of the brain stem and motor cortex, which together make up the corticospinal tract (with a relative sparing of sensory neurons)^{25,23}. Signals are transmitted from motor neurons in the brain (upper motor neurons, UMN) to motor neurons in the spinal cord (lower motor neurons, LMN). From there they are passed through to particular muscles. Degeneration of these motor neurons ceases signal transmission, which result in muscle weakness, spasticity, fasciculation's and atrophy throughout the body²⁶. Eventually progressively loss of voluntary movement is observed²⁷. In the contrary to other anterior horn diseases and motor neuropathology's, such as spinal muscular atrophy or primary lateral sclerosis, ALS shows degeneration of both upper and lower motor tracts²⁸. The primary clinical symptoms and signs of ALS include impairment affecting limb, bulbar, axial and respiratory function. Specific UMN signs are hyperreflexia, spasticity, extensor plantar response (up going toes), and positive jaw jerk, while LMN signs include weakness, muscle atrophy, cramps and fasciculation's. However, differences in site of onset, pattern and speed of spread as well as the degree of upper motor neuron (UMN) and/or lower motor neuron (LMN) dysfunction produce a remarkably variable disorder²⁹. Notably, also frontotemporal dementia (FTLD) is seen in 5% of the cases³⁰.

Although, the clinical diagnosis is primarily based on clinical manifestations, it is supported by laboratory studies, electrodiagnostics and imaging (Supplementary Table.1). At molecular and cellular level, a wide range of processes are associated with ALS as well, such as protein misfolding and aggregation (ubiquitinated and neurofilamental inclusions) and mitochondrial damage. Also energy deficit, ER stress³¹, changes in calcium homeostasis³² and excitotoxicity are seen³³. Furthermore, specific vulnerability of motor neuron degeneration is determined by its characteristics (for a overview about motor neuron vulnerability features, see supplementary table.2). Additionally, degeneration of motor neurons is often accompanied by reactive gliosis³⁴.

These processes have all been intensively investigated and will be shortly reviewed in the next sections.

CAUSES OF ALS

ALS seems to be a multifactorial disease, which appears in a broad variety of disease manifestations caused by a complex interaction among genetics and environmental factors. People of all races and backgrounds are affected³⁵, but there appears to be a slight male predominance³⁶. Although the exact working mechanism remains unclear, ALS is most often seen in elderly individuals, which indicates that aging might be an important risk factor⁵. Furthermore, multiple external risk factors have been associated with ALS, such as: ingestion of high concentrations of β -methylamino- L-alanine³⁷, use of cholesterol-lowering medication³⁸, intensive physical exercise³⁹ including football playing^{13,40} and service in the USA Army⁴¹. The latter possibly links to intermittent work-related hypoxia⁴² or to head injury^{43, 44}. Other environmental factors include exposure to heavy metals⁴⁵, cigarette smoking⁴⁶, and pesticides or herbicides⁴⁷. Although exposure to toxics initially seemed to be important risk factors, no additional evidence has been found in the last 25 years⁴⁸. Lastly, associations are found between ALS, viral infections (HIV)⁴⁹ and prion disease⁵⁰.

GENETICS

Besides environmental factors, several genes have been associated with ALS as well. Approximately 90% of patients⁵¹ suffering from ALS don't have a family history, which is defined as sporadic ALS (sALS)²³. The residual cases are inherited termed familial ALS (fALS). Whereas fALS is known to be caused by mutations in at least 10 different genes⁵², sALS do not show a defined genetic profile. Genes which are associated with ALS include a wide range of cellular processes, namely: involvement into antioxidant response⁵³, axonal and vesicular transport⁵⁴, angiogenesis, endoplasmic reticulum (ER) stress and unfolded protein response (UPR) and RNA metabolism³³.

fALS is inherited as a dominant trait with different penetrance and expressivities among ALS patients²³. Mutations in the SOD-1 gene are the most common cause of fALS (20%), followed by mutations in TDP-43 (5%), FUS (5%), and ANG (<1%). Due to recent developments, more genes become associated with this disease. Mutant SOD-1 was the first gene associated with fALS and many animal models have been developed to further investigate the working mechanism of ALS. However, since this account only for about 20% of the familial cases, other genes are still under intense investigation. However, the cause of sporadic ALS (sALS), in addition to fALS, remains unsolved⁵⁵. Therefore, environmental causes are thought to play an important role. However, many genes, which are found in fALS, are also affected in sALS, such as SOD1 mutations (5%). Also, similar conformational changes within the wild-typ SOD-1 and TDP-43/FUS gene have been discovered within sALS patients compared to their mutants. This suggests that modified wild-type SOD-1 and TDP-43/FUS are able to contribute to the same disease mechanisms.

Moreover, fALS is clinical indistinguishable to sporadic sALS⁵⁶, since they share the same symptoms and pathological phenotype, what highlights the similarity between both forms of the disease⁵⁷. Even survival of affected family members which harbours the same mutation in the same gene have showed to be highly variable⁵⁸. This variability is thought to be caused by genetic modifying factors. However, further research is needed to elucidate the role of genetics in ALS.

ALS DISEASE MECHANISM AND MODELS

This section will discuss the disease mechanism of ALS and the most commonly used models with their application.

DISEASE MECHANISMS

The exact working mechanism underlying the selective motor neuron death in ALS is not resolved yet, although numerous hypothesizes have been proposed. These hypothesizes include proteotoxicity, glutamate excitotoxicity, oxidative stress (mitochondrial dysfunction), proteasome inhibition (aggregate formation and ER stress), non-cell autonomous function mediated by astrocytes and glial cells, compromised axonal transport, inflammation, neurovascular and RNA metabolism. The SOD-1 transgenic animal model has been most widely used in ALS research, since this gene was first identified to be associated with significant subpopulation of both fALS as sALS.

This section will briefly outline the core principles of each hypothesizes. Interestingly, although they all seem plausible, either one of them can be the primary cause of the development of ALS, but also be a consequence of an earlier underlying event. People have always considered ALS as a pure motor disorder. However, subsets of patients have emphasizes the involvement of other cell types and working mechanisms as well. The multitude of contributing factors does underline the complexity of the disease. Nowadays, ALS is more seen as a multisystem disorder in which motor neurons appear to be the most severely affected⁵⁹.

PROTEOTOXICITY

Accumulation of aggregated proteins is the major phenotypical hallmark of ALS pathology^{60, 61}. Interestingly, this is seen in almost all neurodegenerative diseases, including Alzheimer's disease⁶², Parkinson's disease⁶², frontotemporal dementia (FTLD) and Huntington's disease⁶³, but also in prion disease⁶⁴. These protein aggregates consist of damaged and misfolded proteins that cannot be removed by the normal protein degradation mechanisms⁶⁵. TDP-43 was identified as the major component of these insoluble inclusions in both sALS as fALS. After this discovery, FUS has also been identified to be components of protein aggregations. TDP-43 and FUS are both primarily nuclear proteins that participate in common heteromultimeric complexes which are enrolled in RNA transcription, translation, splicing, nucleo-cytoplasmic shuttling, transport for local translation, and stress granule formation^{16,10}. Research suggests that they interact with other proteins by binding glycine-rich domains. Under pathological conditions, they are redistributed to the cytosol where they form abnormal protein-protein associations⁶⁶. Notably, SOD-1, another well known ALS causing gene, has also been identified in protein aggregations⁵⁰. However, toxicity caused by this gene seems not correlated to its aggregation potential. Therefore, further research proceeded on the function of aggregate prone proteins and their proteotoxicity in relation to ALS.

Proper protein folding has shown to be crucial for cellular function and viability⁶⁷. During cell division, damaged and oxidized proteins are sequestered and incorporated by the mother cells. This mechanism enables daughter cells to consist of an undamaged proteome⁶⁸. However, neurons, which are post-mitotic cells, do need another mechanism to ensure protein quality⁶⁷. Many proteins do fold spontaneously but they do need assistance to correct their structures by a specialized set of chaperones, which perform a quality-control process as well. Nonetheless, aggregation-prone proteins challenge these systems, by making them fail to handle them. Also, protein homeostatic system becomes less efficient and less effective during aging⁶⁹.

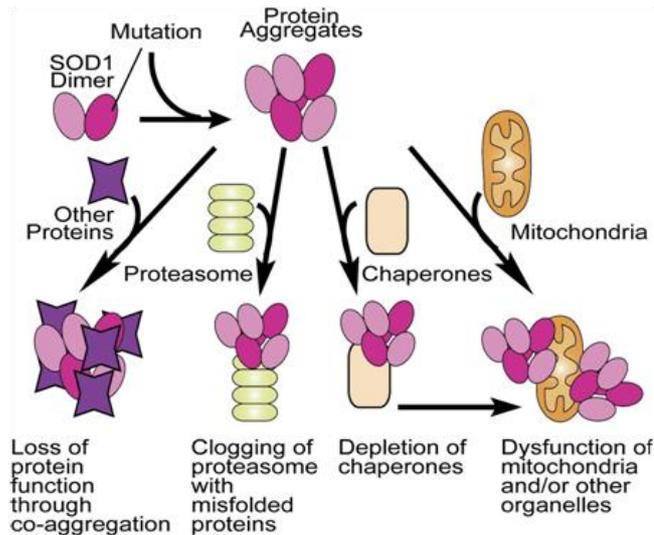


Fig.1: Supposed mechanism of protein aggregation. Aggregation prone proteins causes co-aggregation of other proteins and impairment of proteasome activity. This lead to distortion of other cellular proteins, such as for chaperones and mitochondrial dysfunction. (Image adapted from Boillée et al, 2006).

Unfortunately, a failure to properly fold one protein may result in protein aggregations, which can destabilize the proteome even further. This may lead to uncontrolled aggregation of other proteins as well and causes proteotoxicity⁷⁰. Moreover, when misfolded or damaged proteins are not removed, this disturbs cellular process and can trigger mitochondria dysfunction and the unfolded protein response (UPR), which lead to endoplasmatic reticulum (ER) stress. Beside, studies have shown that mutant SOD-1 activates components which mediate ER stress³¹. Eventually ER stress can cause cell death through caspases-mediated apoptosis. Furthermore, protein aggregation have also shown to cause mitochondrial dysfunction (Fig.1)⁷¹. Nonetheless, there is no universal agreement as to what function they have in disease pathology.

Moreover it is still under debate if protein aggregates could either directly cause neuronal dysfunction and death, or that they arise as a harmless by-product, or if they arise to protect cells by sequestering of toxic proteins. Major evidence for protein aggregation as primary cause is given by the fact that almost all ALS models feature prominent, cytoplasmic, intracellular inclusions in motor neurons as well as in the astrocytes surrounding them⁷². They are highly immunoreactive and develop often before onset of clinical symptoms, as an early sign of ALS⁷³. Aggregative proteins have showed to behave in a dose dependent manner, with enhanced aggregations during disease progression, and to generate motor neuron degeneration¹¹.

Several mechanisms have been suggested to cause toxicity in motor neurons. On the one hand, mutant SOD-1 aggregates may assemble other proteins, which are normally required for neuronal function⁷⁴. On the other hand, aggregations are caused by a reduction of protein-folding chaperones, which are needed to catalyze folding of other proteins as well⁷⁵. Therefore, mutant SOD-1 aggregates causes proteosomal dysfunction, which lead to disrupted protein turnover. Furthermore, imbalance of protein synthesis and degradation compromises its ability to degrade other critical components as well. Finally, almost all aggregates seems to co-express ubiquitin. This was an interestingly finding, since ubiquitin targets proteins to the proteasome degradation pathway for clearance⁷¹. This implies that accumulation of ubiquitinated, misfolded proteins, may affect the proteasome and impair normal protein degradation of proteins, such as SOD-1.

However, recent research has provided evidence consistent with each of the above-mentioned mechanisms. While it was previously assumed that high molecular mass aggregates are the cause of ALS, recent data in this field implies that small oligomeric aggregative structures, which are highly toxic to the cell, may underlie pathology^{76,77}. Therefore the production of less-toxic large aggregates seems to be a protective, compensatory mechanism to detoxify the cell as much as possible^{78,79}. It has now been suggested that during lifespan, the cell's capacity to remove these small toxic aggregates become reduced. This could be either the result of an increased amount of small toxic aggregations during lifetime or to a diminished clearance capacity of the cell during disease progression. In either way, more toxic aggregations appear, which causes damage that may lead to neuronal death. This mechanism seems especially important in low turnover cells (i.g. motorneurons), since protein aggregates are not diluted as in high turnover cells due to cell divisions. Moreover, mother cells retain aggregates during early divisions and budding, which protect new cells from damage.

Taken together, aggregates are seen in almost all neurodegenerative diseases. This accumulation of misfolded proteins appears to disrupt the proteasome degradation pathways and induces ER-stress. The selective vulnerability of aggregates on motor neurons in ALS has to be elucidated.

Current literature has linked multiple protein aggregation to ALS pathology^{9,62,80}. However, until now, only SOD-1 and TDP-43/FUS protein aggregations have been found to contribute to ALS pathology. Although proteotoxicity shows clear indications for the development of ALS pathogenesis, it could be argued if this is the primary cause of ALS, or that these proteins form aggregates upon stimulation of another mechanism. Initiation of protein misfolding includes random processes, caused by a reduction of protein folding chaperones, proteosomal dysfunction or by the formation of abnormal protein-protein interactions^{80, 81}. Therefore, alterations in other proteins are expected to be involved into ALS pathology as well. Questions arise about the working mechanism of proteotoxicity in ALS, since protein aggregations of only certain proteins has been observed to cause ALS⁸².

Protein aggregations have been associated in numerous neurodegenerative diseases. For instance, the accumulation of hyperphosphorylated tau protein, which is involved into the pathogenesis of Alzheimer Diseases (AD)⁸³. Interestingly, certain tau isoforms have been reported in other neurodegenerative diseases as well, such as in ALS⁸⁴. However, not all patients that expresses tau protein aggregates develop ALS pathology⁸⁵. This creates doubts about the specificity of the aggregate protein and may suggests that proteotoxicity is mediated by another mechanism rather than causing ALS itself. In this case, protein inclusions and aggregates may represent an end stage of a molecular cascade, while earlier steps in this cascade may be more directly involved into pathogenesis.

Nevertheless, studies support the idea that at least one aspect of toxicity may arise by processes mediated by misfolded aggregated proteins or by loss of essential components as consequence of protein aggregate formation⁷².

GLUTAMATE EXCITOTOXICITY

Glutamate is the main excitotoxin in the brain, albeit being major excitatory neurotransmitter in the mammalian CNS⁵⁷. During normal homeostasis, glutamate can be increased in the synaptic cleft, but this level will rapidly decrease. In ALS pathology, high glutamate levels are maintained due to increased glutamate release or an impaired re-uptake mechanisms (Fig.2)³². These high levels of glutamate stimulate presynaptic glutamate receptors, which subsequently enhance the release of additional glutamate via a positive feed forward mechanism. This causes increased intracellular Ca²⁺ levels, which lead to excitotoxicity and eventually cell death. Interestingly, excitatory amino-acid transporters (EAAT2) are often down regulated in the motor cortex and spinal cord of ALS patients⁷¹. These EAAT2 receptors are the major terminator of excitatory signals through re-uptake of glutamate from the synapse into glial cells and neurons. Eventually opening of mitochondrial permeability pores will occur due to high calcium concentrations⁸⁶. Consequently mitochondria swell and release reactive oxygen species (ROS) along with other proteins that stimulate apoptosis.

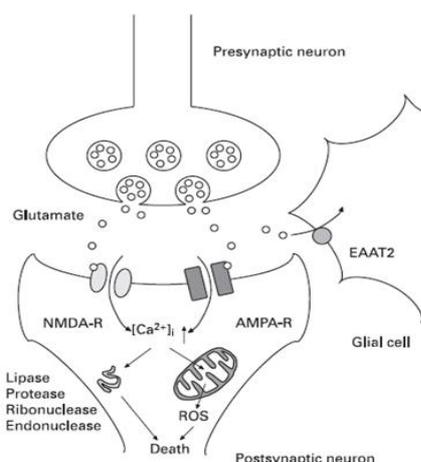


Fig.2: Excitotoxicity in ALS. Excessive glutamate lead to enhanced receptor stimulation. This causes increased glutamate release and inadequate removal and re-uptake. Therefore, electrochemical gradients change and the receptor function converse. Increased levels of calcium activated enzymes that are toxic to the cell. Subsequently, reactive oxygen species (ROS) is produced, which impairs mitochondrial function. This vicious circle eventually lead to excitotoxicity and degeneration of motor neurons in ALS. (Image adapted from Van dan Bosch, et al., 2006)

The pore can also stimulate the mitochondria to release even more calcium, to inhibit the production of adenosine triphosphate (ATP) and to activate ATP synthases, which hydrolyses ATP instead of producing it. Elimination of energy metabolism may distores the electrochemical gradients of certain ions. These gradients are needed to remove glutamate from the extracellular space by activation of glutamate transporters. However, when the electrochemical gradient changes, this causes a reversal in transporter function: instead of removing glutamate they now act as active secreting glutamate channels, which enhance excitotoxicity even more(fig.2)⁸⁷. Moreover, besides the influence of calcium influx on excitotoxicity, it has recently been noted that extra-synaptic NMDA receptor activation stimulates CREB protein inactivation. This results in loss of mitochondrial membrane potential and apoptosis. Additional research on selective motor neuron vulnerability, showed extremely sensibility of motor neurons to excitotoxicity. This selective sensibility is caused by a low Ca²⁺ buffering capacity and a high number of Ca²⁺ permeable AMPA receptors³².

All in all, this hypothesis shows one of the most robust mechanisms and is supported by much evidence. Moreover, the only drug available on the market to slow disease progression is riluzole, which has anti-excitotoxic properties. However, how this excitotoxicity arises remains to be questioned.

It could be argued that if protein aggregates, formed by mutant gene products (as discussed in the previous paragraph), might assemble in motor neurons and/or decrease glutamate uptake in the surrounding glial cells⁵⁷. Interestingly, recent research has found that glutamate hyperstimulation leads to the formation of microaggregates in iPSC-derived neurons. However, further research is needed to investigate if excitotoxicity may underly proteotoxicity as well. Also in this context, it remains unclear why this process would especially involve SOD-1, TDP-43/FUS protein aggregation.

OXIDATIVE STRESS

Oxidative stress derives from an accumulation of reactive oxygen species (ROS), which arise as by-products of aerobic metabolism^{88,87}. With aging, oxidative stress accumulates within the neuron and may cause a reduction in the ability of the biological system to eliminate or to repair ROS-induced damage.

Cellular ROS is caused by a leakage of electrons from the mitochondrial respiratory chain, due to an imbalance between the production and the removal of reactive oxygen. Particularly, the early components from the electron transport chain, complexes I and III, have showed to be prone to leak electrons to molecular oxygen⁸⁹. Therefore mitochondria are the major source of ROS production, followed by the endoplasmic reticulum.

Under normal conditions, the cell has multiple anti-oxidant defence mechanisms to remove ROS and to prevent oxidative stress. One of these mechanisms includes the catalytic removal of ROS by superoxide dismutase, which is mainly regulated by SOD-1. However, mutations in SOD-1 disrupt enzyme function and leads to increased ROS production, which results in enhanced oxidative stress (Fig.3)^{90,73}. Moreover, research showed reduced proteasomal activity in both, mutant SOD1 and TDP43/FUS expressing mice, which is caused by aggregations that disrupt the ubiquitin-proteasome system⁹¹. Under normal conditions this degradation pathway protects the cell by taking up damaged, misfolded and unfunctional proteins. Dysfunction of the proteasome and lysosome systems eventually lead to mitochondrial dysfunction with increased oxidative stress⁹². The process by which ROS creates oxidative stress is as followed: Firstly, reactive oxygen causes incomplete reduction of molecular oxygen during oxidative phosphorylation and lead to the production of hydrogen peroxide (H₂O₂) and the superoxide radical anion (O₂⁻). Subsequently, ROS will react with radical nitric oxide (NO) and will form the oxidant peroxynitrite (ONNOO⁻)^{93,94}, and H₂O₂ will eventually form the highly reactive hydroxyl radical, OH. These radicals causes damage to proteins, lipids and DNA⁵³. This can alter protein conformations and change the active site of the enzymes, change membrane properties by oxidation, and introduce mutations into DNA/RNA. Notably, some components of the electron transport chain are encoded by mitochondrial DNA. Mitochondrial DNA is already exposed to a higher mutation rate compared to nuclear DNA⁹². However, once oxidative stress is present, other mechanisms become involved as well, such as excitotoxicity. Excitotoxicity causes increased intracellular calcium levels, which enhance ROS production. Nonetheless, ROS seems to inhibit glutamate uptake through loss of EAAT2 transporter expression and enhanced expression of calcium permeable AMPA receptors in glial cells. Furthermore, activation of microglial results in secretion of cytokines and further ROS. Lastly, mutant protein aggregates inhibit neurofilament assembly and cytoskeletal transport.

Thereby, the central nervous system appears to be more susceptible to oxidative damage than other tissues. This is caused by the accumulation of metal ions throughout the CNS, which favor ROS production³³. The selective vulnerability of motor neurons may be caused by their high

metabolic input. Motor neurons have a relative big size and long axon length. This high-energy demand goes along with many mitochondria, with the side effect of increased oxidative stress.

Taken together, oxidative stress regulates multiple mechanisms, but in light of ALS, neuronal degeneration may result from a complex interaction of ROS, excitotoxic stimulation, genetics and dysfunction of proteins and organelles (e.g. mitochondria, ER)⁵³. Furthermore, the neurotoxic effects of ROS are not only limited to neurons, but could also affect glial cells. However, further research has to be performed to determine the causative role of oxidative stress in ALS manifestation.

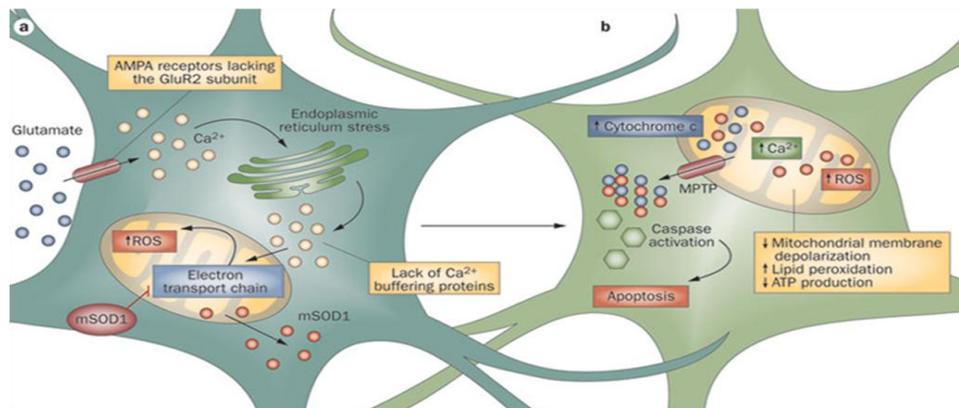


Fig.3: Molecular pathways involved in oxidative stress. Oxidative stress is caused by the enhancement of ROS, excitotoxicity and protein aggregates. (Image adapted from Ferraiuolo, L., et al. 2011)⁵⁷.

COMPROMISED AXONAL TRANSPORT

While previous stated mechanisms share many aspects and seem to be related to each other, compromised axonal transport includes a distinct mechanism.

Since motor neurons have extending axons that can be more than a meter in length, efficient transportation systems are needed. In both, patients and animal models of ALS, alternations in large (myelinated) axonal structure by misaccumulation of neurofilaments have been reported. Also the two forms of axonal transport, kinesin-mediated (anterograde) and dynein-mediated (retrograde), are decreased⁹⁰. Notably, transgenic mice which express a mutation in the NF-L subunit of neurofilament, which is similar in human, develop motor neuron degeneration⁹⁵. However, deletion of this mutation reversed the phenotype partly and caused prolonged survival^{96,97}. Consistently, presymptomatic deficits of slow axonal transport has been described in mutant SOD1 mice⁹⁸, which showed similar misaccumulation of neurofilament.

Furthermore, genetic evidence implicates axonal transport defects as an important mechanism in the development of motor neuron degeneration. Altered gene and protein expressions have been found in multiple genes associated with transport, like: loss of kinesin motor proteins KIF1B β ⁹⁹, KIF5a¹⁰⁰, KIF21A¹⁰¹, VAPB⁵⁴, Rab7¹⁰², and Vps54¹⁰³. Also, mutant SOD-1 mice showed significant inhibition of retrograde axonal transport before onset of the disease⁹⁰.

Although many of these features are similar to human ALS pathology, abnormalities in neurofilaments could be either causal or just being a harmless byproduct of neuronal degeneration of mutant SOD1 protein. Moreover, similar as with other protein aggregations,

accumulation of neurofilaments in motor neuron cells could also arise from a protective view of perspective: to protect the neuron against SOD1-mediated damage by buffering diverse components. Nonetheless, it has been shown that particular motor neurons, are highly sensitive to defects in axonal transport⁹⁰. This may be due to their extreme polarization. So while protein aggregation, glutamate hyperstimulation and oxidative stress are linked, axonal transport seems to be a separate potential cause.

ASTROCYTES AND GLIAL: NON-CELL AUTONOMOUS DEATH OF MOTOR NEURONS

Although most research is performed on previous outlined hypothesizes, there is growing evidence for a non-cell autonomous death of motor neurons. This means that the degeneration is not caused by the motor neurons themselves but rather through extracellular influences. Research showed that glial cells play a particular important role in this. On the one hand, astrocytes support neurons by providing them with nutritional factors and regulate neurotransmitter concentrations, metabolic or ionic homeostasis¹⁰⁴. While on the other hand microglial cells clear damaged and dead cells from the environment. Gliosis is an early process seen in pathogenesis in both the brain and the spinal cord¹⁰⁵. Although ALS is caused by motor neuron degeneration, neighbouring cells may mediate this in a non-cell autonomous manner. Consistently, all known familial forms are caused by mutations in genes that are ubiquitously expressed (i.g. *SOD1* and *VAPB*) throughout the body or at least expressed in multiple cell types (i.g. *VEGF* and *ANG*). Therefore, researchers started to examine the role of glial cells.

To investigate this in detail, chimeric animals were developed that express mutant SOD-1 selectively in motor neurons, microglial cells or astrocytes¹⁰⁶. Results showed that expression in all three cell types were needed to cause ALS. After this, they removed 30-50% of the mutant SOD-1 gene in motor neurons and found a delayed onset while disease progression remained the same¹⁰⁷. Animals who expressed a combination of mutant and wild type SOD-1 showed a positive correlation with the proportion of surviving mutant SOD-1 expressing motor neurons and lifespan of the mice. In addition, research with *C.elegans* has confirmed this. Glial cell activation also increases after motor neuron injury. After damage, motor neurons seem to release mutant proteins, fragments or distressed signals to the extracellular environment. It has been proposed that glial cells then start to secrete protective components. However, if the stimulation becomes chronically, toxic substances are released instead. These released proteins are involved in the production of nitric oxide or inflammation, to activate other glial cells. However, it is not known how they become activated in the first place, neither how this especially involves motor neurons. It could be that this results from specific signals released from the motor neurons after injury as stated above. Nonetheless, it seems that this active dialogue between motor neurons and glial cells contributes to disease progression.

Since fALS and sALS show similar disease manifestation, it is suggested that a more general principle is causing the disease. A non-cell autonomous mechanism might underlie this principle¹⁰⁷. However, existing disease models do not give the possibility to research a general mechanism for fALS and sALS. Therefore, induced Pluripotent Stem Cells (iPSCs), an upcoming in vitro technique, used more and more to further decipher the influence of astrocytes and glia on disease progression¹⁰⁷. Evidence deriving from this novel technique will be discussed in more detail in a later chapter.

INFLAMMATION: VIRAL INFECTION AND IMMUNE IMBALANCE

A common feature of ALS is the presence of neuroinflammatory reactions consisting of activated glial and T cells. Until recently, this feature has been viewed as a consequence rather than a possible cause of motor neuron degeneration¹⁰⁸. However, focus has now been put on inflammation as a target to treat ALS¹⁰⁹. Thus, instead of non-cell mediated motor neuron degeneration caused by intrinsic properties of glial cells, a primary role of inflammation has also been proposed.

Upon injury, microglia accumulate. Investigators have discovered a positive correlation between this microgliosis and disease progression¹¹⁰. Although it was already known that microglia becomes increasingly activated after motor neuron injury, research now also showed activation of microglia before motor neuron degeneration and thus before clinical disease-onset¹⁰⁹. Animal models were used to further investigate the potential link between inflammation and motor neuron degeneration.

Microglia are the macrophages in the nervous system. Next to monitor the extracellular environment, they closely interact with astrocytes and neurons. This is mediated by the M1 activation pathway. They can be identified as CD11b expressing cells and are activated by a range of different signals. Furthermore, they act as the first line of defence against infection or injury. When activated, they acquire an amoeboid like appearance and start to secrete proinflammatory molecules, like: tumour necrosis factor TNF α , interferon- γ , and interleukin 1- β . Furthermore they also upregulate NO and O₂. This mechanism protect against invading organisms and clears hazards. Besides this, activated microglia also mediate T-cells infiltration into the CNS and modulate the neuroinflammatory reactions. To regulate the extant of the immune response neighbouring astrocytes and inflammatory T-cells release anti-inflammatory molecules via the M2 activation pathway, e.g. insulin-like growth factor 1 (IGF1), interleukin 4 and 10.

In more detail, T-cells seemed to be the critical components in mediating disease progression. They are protective prior to onset of disease, but when the disease proceeds, T-cell number increase and their cytotoxic influence outweigh their neuroprotective effect. It has been suggested that the absence of this neuroprotective effect, mediated by the immune system, determines disease onset¹¹¹. Research have found aberrant proinflammatory cytokines and an increased number of proinflammatory mediators in patients with ALS. Moreover, anti-inflammatory drugs have showed to be efficacious on several aspects of ALS in mutant SOD-1 mice. Thereby, inflammation also mediates exitotoxicity, which causes increased Ca²⁺ into motor neurons, which lead to neuronal death. In healthy conditions, astrocytes reduces the Ca²⁺ permeability of AMPA-type glutamate receptors by enhanced expression of Glur2 in motor neurons¹¹².

Although there seems to be a concededly effect of gliosis on ALS onset and disease progression, the influence could be mediated by other processes as well. So are oligodendrocytes, ependymal and subependymal cells proposed to involved in neuroinflammation as well. However, their function in ALS is poorly understood¹¹³

NEURO-VASCULAR SYSTEM: A NEUROVASCULAR DISEASE

After the discovery of the influence of local inflammation, researchers started to investigate systemic regulated inflammation¹¹⁴. As discussed above, glial cells seem to be activated in patients with ALS. Recent research has discovered that neuroinflammation is regulated by the peripheral immune system, rather than through local responses. Particularly T-cells seem to be prone to be transferred from the systemic regulation into the brain and spinal cord parenchyma.

However, this field is still standing in its infancy. Therefore, it is important to further investigate the systemic influence on changes that occur in the brain during disease progression and to unravel the exact interaction between the peripheral and the local immune responses. Up-regulation of 32 cellular adhesion molecules has already been found. Adhesion molecules may be responsible for the arrest of circulating leukocytes and diapedesis into the brain parenchyma¹¹⁴. Controversially, tight junction proteins emerge to be down-regulated, which leads to increased vascular permeability¹¹⁵. From this point of view, it is interesting to further focus on the role of the blood brain barrier (BBB) and the blood spinal cord barrier (BSCB), as these are the separations between systemic and local immune responses connected by the vascular system. Various structural and functional alterations of the BBB/BSCB have been found in ALS patients (Fig.4)¹¹⁴. Major hallmarks of BBB/BSCB impairment are endothelial cell and astrocyte end-feet degeneration, modified basement membrane composition, tight junction and transporter system impairment, vascular protein leakage, extensive extracellular edema, as well as myelin protein (MP2) and matrix metalloproteinase 9 (MMP9) activation¹¹⁶. These processes eventually allow entry of blood borne substances into the brain, which might set up a cascade which may lead to direct and/or indirect motor neuron degeneration. Furthermore, high levels of IgG, albumin and complement C3a deposits have been found in the spinal cord and motor cortex, which implies vascular ruptures. These deficits appear to occur before expression of inflammatory molecules (i.e. before macrophage activation) in the brain and before motor neuron degeneration¹¹⁷. Interestingly, pre- and post- symptomatic mutant SOD-1 mice have shown decreased capillary length, capillary diameter and capillary¹¹⁵. This may imply ischemia or a similar event, that underlie this pathology. However, inconsistencies have been found between studies in both human as in mice.

In conclusion, motor neuron degeneration does not appear to be intrinsic. Systemic regulation seems to mediate neuronal survival and death, by regulation of various cell types (i.e. glial cells) and components within the CNS. Impairment of the BBB/BSCB may be the primary event prior to ALS onset. Furthermore, local BBB/BSCB impairment at the spinal cord or brainstem may cause the selective motor neuron degeneration in ALS. Nonetheless, it remains unclear how big the systemic influences are with respect to the severity of changes in glial cells (M1 and M2 activity) and their influence on disease progression. However, some controversial data has been found. Therefore, additional research is needed to unravel these mechanisms further. Though, this field of research showed promising results and may lead to future therapy on maintaining, repairing, strengthening the BBB/BSCB and reducing inflammation¹¹⁴.

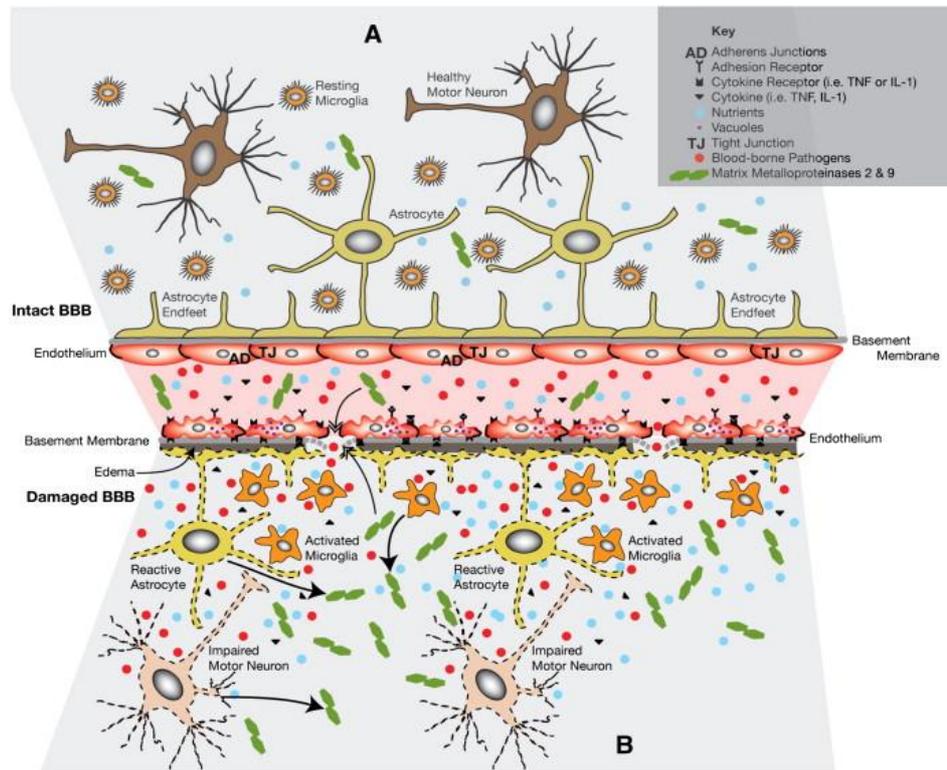


Fig.4: ALS a neurovascular mechanism. **A.** Represent a healthy condition; the BBB/BSCB maintains homeostasis through selective regulation of the blood circulation within the spinal cord and brain. This process includes endothelial cells and their tight/adherens junctions, astrocyte end-feet, perivascular macrophages, pericytes, and the basement membrane. They are all needed for providing a barrier. **B** Structural and functional impairment of the BBB/BSCB allows entry of blood-borne substances, by endothelial cell and astrocyte end-feet degeneration, modified basement membrane composition, tight junction and transporter system impairment, vascular protein leakage, extensive extracellular edema, and MMP-2 and MMP-9 activation. Additionally, cytokines (i.e. TNF and IL-1) are released from activated microglia and reactive astrocytes. (Image is adapted from Garbuzova-Davisa, S., et al., 2011) ¹⁰⁷.

RNA METABOLISM

Recently, another disease mechanism has been suggested to play an important role in ALS as well. Currently, an increasing number of mutations, which affect proteins involved in RNA processing, have been identified in ALS. These mutations imply disrupted RNA dependent mechanisms (Fig.5). Evidence for these modifications in epigenetics, transcriptomics, and proteomics have been found in both animal models as in humans suffering from ALS³³. On the one hand modified expression of genes involved in cytoskeletal dynamics, protein degradation system and mitochondrial dysfunction has been found in motor neurons¹¹⁸. Similar altered gene expression have found in muscle. On the other hand alternations in the insulin-like growth factor-1 and the RNA-binding protein ROD1¹¹⁹ occur in glial cells. Therefore, altered control of gene expression might be the overarching mechanism of ALS²⁴,

Epigenetic modifications showed to underlie altered gene expression, due to a disparity between histone acetyl transferases (HATs) and histone deacetylases (HDACs). The function of HATs is to modify core histone tails, which enhance DNA accessibility to transcription factors (TFs). In addition, HDACs activity results in transcriptional repression and gene silencing. However, many of the transcription initiation complex TF are themselves substrates to HATs and HDACs. Interestingly, ALS linked proteins, TDP-43 and FUS/TLS, control the expression of certain HDACs ¹²⁰.

Another way in which epigenetic control of transcription may be modified is by methylation of DNA methyltransferases (DNMTs) or histone methyltransferases (HMTs). Both use S-adenosyl-methionine (SAM) as methyl donor. In eukaryotes, DNA methylation occurs through covalent modifications of cytosine residues in CpG dinucleotides leading to gene silencing. Histones or TF can be methylated by lysine or guanidiny residues according to their function (i.g. activate or repress transcription). These processes may have enormous impact on ALS.

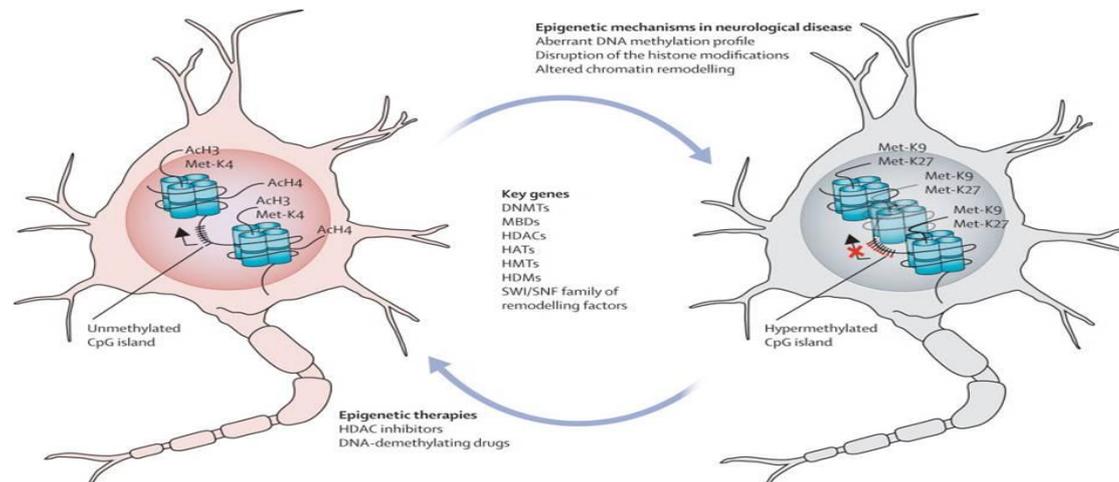


Fig.5: Epigenetic mechanisms. Healthy neurons or glial cells (left) express mRNA of a gene in occupancy from an unmethylated promoter, CpG island and a set of histone modifications. During ALS development altered DNA methylation, histone modifications and chromatin remodeling appear. (Image adapted from Urdinguio, R.G, et al. 2009)¹¹⁴.

Another hypothesis is that RNA dysmetabolism could underlie the altered gene expression as well (Supplementary fig.1)¹²⁰. Genes, which have already been identified in patients with ALS, appeared to be involved in the following processes: pre-mRNA splicing, mRNA transport, translational regulation and mRNA decay. Nonetheless, additional research has to be performed to investigate the specific motor neuron vulnerability to mutations in RNA-binding proteins.

In general, the function of a RNA protein is dependent on its associated proteins (i.e. RNA binding proteins, RBP) that together forms the ribonuclear particle (RNP). These complexes regulate RNA processes, such as alternative splicing. Alternative splicing is responsible for the generation of multiple mRNAs from a single transcript. Changes in the secondary structure of a protein subsequently lead to functional alterations. Moreover, various genes have already been found to express multiple splice variants.

Although the protein is still unknown, the C90RF72 gene is most likely involved in RNA processing. Recently it has been associated with an extension of a noncoding GGGGCC hexanucleotide repeat in fALS patients. When transcribed, this repeat forms intracellular accumulations of RNA fragments in cells¹²¹. This sequence may represent a potential binding site for RNA-binding proteins as hnRNP A2/B1 (TDP-43 interactor). To date, it has identified as most common genetic defect found in ALS patients¹²².

Another interesting player in RNA processing is TDP-43. Next to the fact that it has already been identified as a major component in ubiquitinated inclusions¹¹, it showed to be an

ubiquitously expressed RNA-DNA binding protein. Research found that TDP-43 is implicated in multiple aspects of RNA processing, including: transcriptional regulation, alternative splicing and microRNA processing. Pathologic cytoplasmic TDP-43 inclusions are seen in both motor neuron and glial cells in various animal and human ALS patients⁵⁹. This could imply different cascades in these genetic different phenotypes. Most mutations in TARDBP are located on a glycine-rich domain involved in protein-protein interactions and nuclear transport.

Additional evidence that dysregulated RNA processing may contribute to motor neuron injury in ALS arises from the detection of biomarkers of RNA oxidation³⁴. Furthermore transcriptional repression also occurs within motor neurons in the presence of mSOD1¹²³. This may provide a possible link between epigenetic modifications with mitochondrial damage and oxidative stress in ALS.

Oxidative stress and mitochondrial dysfunction often occur together. Excessive ROS production negatively influences the functionality of the organelles that in turn would produce more ROS. Presumably, an interaction between oxidative stress, redox signalling and epigenetic modulations may cause ALS outcome. In this vicious circle, oxidative stress modulates gene expression by alternations of DNA accessibility. Additionally it is also a modulator of TFs. The latter implies that ROS and HDACs may coincide within the same mechanism. Next to being involved in regulation of proteins in redox reactions, HDACs also modulate alternative splicing and the activity of TFs. Furthermore, mitochondrial damage causes also alternations in selected splicing variants and RNA metabolism. This is associated with SOD-1 linked ALS as a consequence of mitochondrial stress¹²³. Eventually, this may cause proinflammatory responses.

Overall, ALS may arise from RNA dysmetabolism, which explains the broad diversity of modified processing within the brain and muscle. Furthermore, altered expression of genes involved into RNA metabolism has been found in motor neurons. But besides mutations, which affect RNA transcription or processing, RNA proteins, also need to be correctly transported along the axon to the neuromuscular junction (NMJ). Axonal inclusions may block axonal transport, which may underlie ALS disease onset. Therefore, the primary site of damage in an ALS patient might determine the severity, age of onset and progression³³. Moreover, different alternations of gene and/or RNA expression may cause the different outcomes on disease pathology. However, RNA dysmetabolism may also be caused by oxidative stress. For the future, this field of research should definitely be further investigated.

DISEASE MODELS

Since the wide variety of disease manifestations, research is performed on many different aspects of ALS. To investigate the cellular and molecular basis of ALS, a wide variety of model organisms have been used, including nematodes (*Caenorhabditis elegans*), arthropods (*Drosophila melanogaster*), fish (zebrafish, *Danio rerio*), rodents (mouse, *Mus musculus* and rat, *Rattus norvegicus*) as well as non-human primates (rhesus monkey, *Macaca mulatta*) (See the supplementary table.3 for an overview)¹⁶. These animal models are particularly important for certain pathological and therapeutic studies which are impossible to perform in human ALS patients. Nonetheless, postmortem tissue remains also very important for investigation of the disease.

To establish a good working model, much research has been performed on human genetics. To mimic the disease in model organisms, a pattern of inheritance is important. Linkage studies have provided knowledge of chromosomal loci and sequencing of putative loci has identified multiple

genetic changes¹⁶. With this knowledge, transgenic systems are generated to model the disease or aspects of the altered gene function. These systems can be used to further confirm the genetic basis and aid to further investigation of the cellular and molecular disease phenotypes.

Nowadays, the most widely used model includes the SOD-1 transgenic animal model³. Much knowledge is obtained from this model about the ALS disease mechanism (Supplementary Fig.2). However, the utility of the (mutant) SOD-1 model is questioned due to recent controversial results.

There is growing evidence that overexpressed wild-type SOD-1 can also be neurotoxic. Research has showed that overexpression of wild-type SOD-1 result in SOD-1 misfolding and aggregate formation that lead to motor neuron degeneration^{72,124}. This neurotoxicity seems to result from a gain of function, rather than through loss of SOD-1 activity^{125,34}. It might be that conformational changes in wild type SOD-1 mimic structural features of fALS mutant forms of this gene (13-15). These conformation changes might either be caused by altered post-translational modifications or through induction of covalent aberrant modification to wild-type SOD-1. Interestingly, similar results were recently been found in the TDP-43/FUS gene in *Drosophila* and transgenic mice¹²⁶. However, another hypothesis states that environmental and lifestyle related factors are able to induce the wild-type SOD-1 or TDP-43/FUS to cause ALS⁹.

However, alterations within this genes may also be more universally involved in ALS pathology⁶⁰. These data might imply that aberrantly modified wild-type SOD-1 and TDP-43/FUS share a similar mechanism as fALS. Furthermore it would also be important to investigate whether overexpression of these proteins occurs in human ALS patients, too. Perhaps these mechanisms would underlie sALS patients.

If SOD-1 and TDP-43/FUS are involved into a more general mechanism that causes ALS, more research should be done to reveal the exact role or function of these genes. Therefore, previous experiments on various processes may have to be repeated as well. This may lead to new insights about ALS. Nonetheless, these observations suggest that sALS and fALS might share common pathogenic mechanisms.

In general, our lack of knowledge about ALS disease mechanisms and the absence of drugs or therapies reflects the limitations of existing model systems²⁴. It has now been recognized that the biology and progression of ALS differs in humans compared to animal models. Also, positive effects, which are observed in animal models, seem to have little or no effects on the disease in humans. Therefore, evolutionary conservation on anatomical as well as cellular levels has to be taken into account to improve experimental studies.

Anatomical divergence is most notably in the rostral regions, such as the cerebellum and the forebrain, but also the position of the corticospinal tract¹²⁷. This would argue for a different evolutionary process that may also govern patterning of cortical motor neuron precursors¹²⁸. This has also been supported by the observation of more subtle differences between human and lower primates or vertebrates, like the direct synaptic contact between the lower and upper MN and the more complex axonal projections of the upper motor neurons^{129,25}.

There are multiple differences at cellular level as well. Firstly, human transcription factors binding sites have showed to be functional different from rodents¹³⁰. Secondly, loss or gain of function of genes has been observed. For instance, the expression of certain genes, like the recently identified gene sulfiredoxin, contributes to increased anti-oxidant defence in rodents. However, this function has been lost in the primate lineage. To overcome these differences, newly humanized

models have to be developed. Currently, in vitro based technology, with special emphasizes on induced pluripotent stem cells (iPSCs), seems to be the solution^{131,132}. This technique uses patient derived somatic cells that can easily be reprogrammed to pluripotent cells and then differentiated to specific cell types.

In conclusion, therapeutic strategies that were thought to be efficacious in animal disease models have proved to be unsuccessful when translated to human trials²⁴. Our lack of knowledge about the disease mechanism and the evolutionary differences between animal and human hinder the process. Therefore, the development of new humanized models is needed to overcome these differences. iPSC technology may provide the solution¹³¹.

DISCUSSION

After decades of research on ALS pathology, various disease mechanisms have been postulated (Fig.6)^{4,31,53,120,71,114,71}. Interestingly, much coherence is seen between these mechanisms in a complex integrative manner.

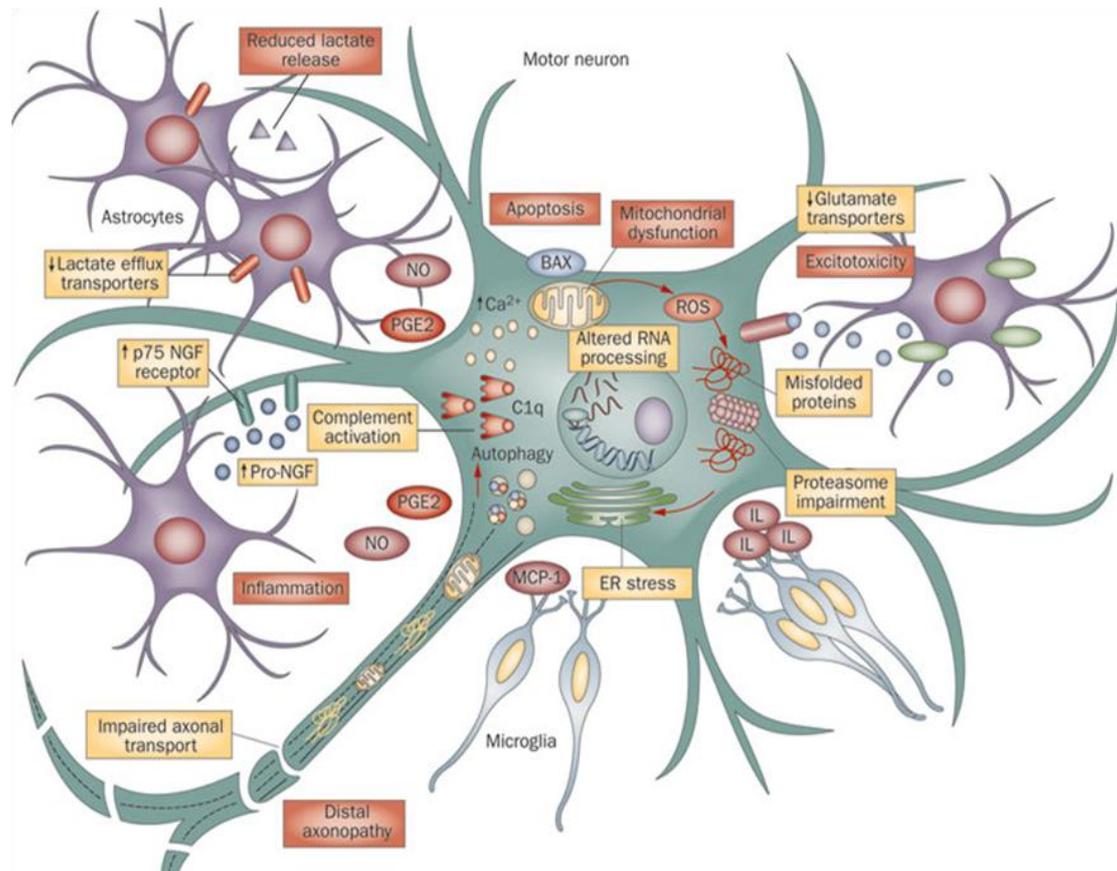


Fig.6: All mechanisms that are involved in ALS pathology: proteotoxicity, excitotoxicity, oxidative stress, impaired axonal transport, non-cell autonomous influences of glial cells and RNA metabolism. Only with exception of inflammation and immunity and neurovascular regulation. (Image adapted from Ferraiuolo, L. et al. 2011)³⁷.

Considering that the age of onset, start of deterioration and disease progression differ greatly among patients, it is an apparent question whether ALS might not be one disease¹³³. Instead it could be that what we call ALS, might be an overarching term for diseases where motor neurons degenerate due to different causes¹³³. This leads to three hypothesises. Firstly, it may be that different mechanisms can lead to ALS. Secondly, ALS may be caused by different mechanisms

that all results into the same outcome. Or finally, knowing that all the suggested disease mechanisms are related to the vulnerability of motor neurons, it raises the question which comprehensive cause might explain all hypotheses, and therefore, be the most likely cause of ALS.

Considering all aspects of the disease, it is striking that many of these take also place in “healthy” aging, albeit later and in a less pronounced manifestations ^{134,135}. A more general mechanism, such as aging, explains the natural variation in symptoms, even as a variation in disease onset. In the next chapter, this novel hypothesis will be described and discussed in greater detail.

A NEW CONCEPT: AGING

The incidence of ALS is highest among the elderly and occurs most often in individuals between 50 and 70 years². Therefore, aging was already seen as a risk factor. Though there was no knowledge about the function of aging itself. For decades, aging was thought to be a stochastic, uncontrolled degradation process independent of gene regulation¹³⁶. But this thought has been changed in the past ten years and much knowledge is obtained about the aging process. Therefore, it is tempting to further investigate aging as a potential cause of ALS.

Interesting findings on the role of aging have been identified by the use of parabiotic mice models^{137,138}. Parabiotic mice are mice that have been sutured together, side-by-side, along the lateral aspect of the body. Once blood vessels have connected with each other, the mice will share a common blood supply. This model can be used to determine whether the effect of aging in one, is affected by a blood-borne mediator in the other. Therefore, heterochronic (Young and old) pairs were compared with isochronic (young to young or old to old) pairs. These data show that exposure of young serum to old mice increase the rejuvenation and regeneration capacity of the old mice, which also showed that the effects of aging on regeneration capacity are reversible¹³⁷. These Age-related changes seemed to be determined by the age of the systemic environment, rather than by the cell-autonomous age. Notably, when young serum became mixed with old serum, the regenerative potential was inhibited, suggesting a dominant influence of old serum factors.

Moreover, aging has showed to be dependent on the regulation of classical signaling pathways, which are influenced by environmental and physiological cues. It has now been revealed that at least three independent metabolic pathways are involved in regulation of the aging process. Firstly, dietary restriction showed to extend the lifespan in multiple animal models^{136,139}. Secondly, the insulin/insulin-like growth factor 1 (IGF-1) signaling (IIS) pathway has been associated to be an active regulator of longevity in worms¹⁴⁰, flies^{141,142} and mice^{143,144}. Lastly, also

the rate of mitochondrial respiration was found to influence lifespan. Among these pathways, the insulin/insulin-like growth factor 1 signaling (IIS) pathway is the most well-known pathway and has already been intensively studied. Therefore, we will focus on how this pathway contributes to aging in relation with ALS pathology. But first, the function of aging during the evolution will be shortly discussed.

EVOLUTION

Research has been performed on mammalian species to investigate to which extend normal brain aging is conserved through evolution. Knowledge about its conservation has important implications for the use and/or the development of animal models of ALS. Genome-wide gene expression studies in *C. Elegans*, *Drosophila melanogaster*, mice, rat, chimpanzees and humans have revealed functional conserved genes with age-dependent expression changes⁵. Notably, most genes are associated with reduced mitochondrial functioning. Furthermore, an increased expression of genes that are involved into the stress-response pathway have been identified. Resistance of stress during aging may be effective to protect the brain against pathology. However, research on evolutionary conservation of aging genes has showed a shift between rodent and human lineages, as rodents showed repression rather than activation of many of these age associated genes during aging (Fig.7). When working with animal models, this should be taken into account.

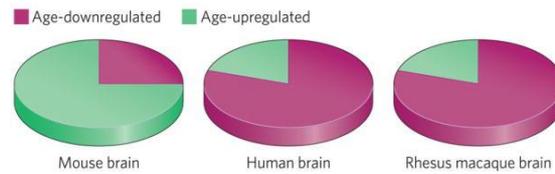


Fig.7: Evolutionary age-related gene expression shift in the primate lineage. Age-related changes are downregulated (green) in Rhesus macaque and human in contrast to downregulation (green) in the mouse. (Image adapted from Bishop, N.A., 2010)¹³⁸.

THE IIS PATHWAY

The strongly conserved Insulin/IGF signaling (IIS) pathway was the first pathway discovered to be involved into aging. This pathway showed to be a major regulator in numerous processes including lifespan, metabolism and stress resistance. Moreover, has recently been associated with neurodegenerative diseases as well⁹. On the one hand, reduced insulin/IGF-1 signaling seems to correlate with lifespan extension⁸ and thereby decrease risk to develop ALS. On the other hand altered expression of these factors may result in a broad cascade of cell-protective mechanisms, which extend or shortens lifespan. Multiple polymorphisms within genes involved into this signaling pathway, have been observed in ALS models of worms, flies and mammals, which are associated with longevity¹⁴⁵.

While invertebrates seem to have only one receptor that binds components like insulin and IGF-1, mammals contain different kind of receptors for these factors. IGF-1 mainly controls growth, whereas insulin regulates metabolism. Defective signaling of the latter results in insulin resistance and diabetes, while defective signaling of the former leads to protein breakdown and muscle degeneration. IGF-1 overexpression reduces various, age-associated organ dysfunction and increase muscle reparation. Controversially, reduced insulin seems to extend lifespan, particularly in tissue which expresses low IGF-1¹⁴⁶. Therefore, specific tissue modulation by different signaling pathways might obstruct aging in humans. This contradiction between their neuroprotective effect versus their deleterious effect resembles the complexity of this signaling pathway.

In general, this pathway becomes activated when a insulin like ligand binds to the insulin/IGF-1 receptor DAF-2^{140,147}. Subsequently, DAF-2 recruits insulin receptor substrates and the phosphoinositide 3-kinase (PI3K) adaptor/regulatory subunit (the human orthologues of IST-1 and AGE-1 in *C.elegans*)¹⁴⁸. This mediates the generation of phosphatidylinositol-3,4,5-trisphosphate (PtdIns(3,4,5)P3). Next, members of the AKT kinase family in a pyruvate dehydrogenase kinase (PDK)-dependent way (37,38) are activated. However, dissociation of PtdIns(3,4,5)P3 and reduction of AKT activation is negatively regulated by PTEN (the phosphatase DAF-18 in *C. elegans*)¹⁴⁹. Upon activation of AKT, the downstream protein DAF-16 (a FOXO forkhead transcription factor)¹⁵⁰ becomes phosphorylated. This protein in turn is not able to enter the nucleus. Therefore DAF-16 is negatively regulated by the IIS pathway. Nonetheless, this system becomes even more complex when taking the other FOXO transcription factors, the silent information regulator (SIRT) protein deacetylases (sirtuins) and heat-shock factor 1 (HSF-1) into account.

On the one hand FOXO transcription factors initiate stress-induced cell death (apoptosis) in damaged cells. On the other hand they can also upregulate antioxidant defence and DNA repair facilitating genes⁷⁰. Sirtuins belong to a class of proteins that are involved in aging, stress resistance, apoptosis and energy metabolism. SIRT-1 is able to deacetylate FOXO transcription factors. This enhances FOXO dependent resistance to oxidative stress and cell cycle arrest, but inhibits FOXO regulated apoptosis¹⁵¹. SIRT1 can also deacetylate the p53 tumour suppressor protein, which diminishes its transcriptional activity and inhibits stress-induced apoptosis and cellular senescence (irreversible cell cycle arrest)⁶³. However, it seems to be difficult to decide whether enhanced FOXO or SIRT1 would increase or decrease longevity. The complexity of this system is probably due to protection of the organism against cancer. That would also explain why yeast, flies and nematodes, which don't develop cancer, contain only one kind of receptor in this signalling pathway. Notably, many malignancies are caused by Igf-1 and Igf-2 signalling, which supports the dual role of this pathway. Moreover, research has shown that certain mutations in insulin, IGF-1 or their receptors greatly extend lifespan. This has been clearly depicted in research in *C.elegans*, in which the IIS pathway has been shown to play an important role on development¹⁵². IIS is postulated to regulate the developmental switch, which determine whether the development of the larva will arrest, a condition called dauer, or if the larva will become adult. Notably, regulation of lifespan in *C.elegans* occurs during the reproductive adulthood, which is distinct from the dauer switch during development. Inhibition of DAF-2 (the IIS receptor) in worms prolonged lifespan and protected them from protein aggregation(Fig.8)^{82,153}.

Next to FOXO and SIRT1, also HSF-1 has showed to be crucial in longevity¹⁵⁴. HSF-1 regulates multiple processes, ranging from development¹⁵⁵, the stress response⁴, the circadian rhythmicity, innate immunity, to hypoxia. Recently it has been associated with the IIS pathway as well⁴. Although the exact working mechanism remains unclear, it has been suggested that apart from DAF-16, reduced IIS promotes enhanced stress resistance by activation of HSF-1. Remarkably, Sirt-1 has also been identified to activate HSF-1 in mammals^{156,69}.

Its target genes include additional small HSPs and chaperone proteins. Interestingly, a link between aging and ALS-like protein aggregations has been found in *C.elegans*. In this model, PolyQ aggregations showed to be reduced by DAF-16 and HSF-1 expression, which is regulated by reduced IIS. Additional evidence was found by a worm model of Alzheimer Diseases which showed that reduced IIS protects against proteotoxicity⁸². Also knockdown of hsf-1 led to a huge increase of soluble toxic aggregates and increased high molecular mass aggregates⁸². This may suggest that the IIS pathway has two distinct counter activities, namely: protective aggregation of high molecular mass aggregates regulated by DAF-16 and disaggregation of the more toxic oligomers by HSF-1. In general, reduced IIS signalling has been associated with longevity and stress resistance in mice¹⁴³.

However, in contradiction with previous findings, increased IIS appears to be protective against proteotoxicity and polyQ-mediated toxicity¹⁵⁷. On the one hand, stimulation of the IIS pathway by injection of IGF-1 results in decreased A β aggregates in Alzheimer's disease (AD) models. On the other hand activation of this pathway also leads to phosphorylation of the polyO gene, dependent on the IGF/AKT pathway. Eventually it will be cleared via autophagy, which has previously been implicated as a crucial regulator of aging¹⁵⁸.

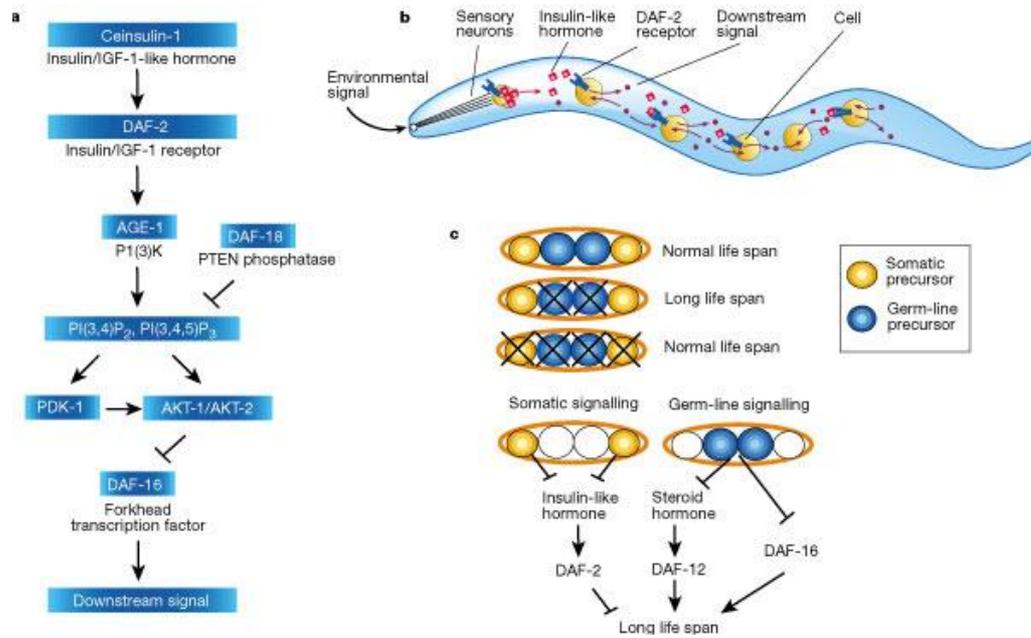


Fig.8: Involvement of the IIS pathway in longevity in *C.elegans*. a, The insulin/IGF-1-like signalling pathway regulates ageing in *C. elegans* (further information can be found into this chapter above). b, c The lifespan of *C. elegans* has showed to be regulated by an elaborate endocrine system that involves at least four hormones. Surprisingly, lifespan seems to be very plastic. This process is modulated by two systems: the sensory system (registers environmental conditions) and its reproductive system. Both systems affect lifespan by activation of components of the DAF-2 insulin/IGF-1 pathway. b, Modulation of lifespan regulated by the sensory system. Environmental signals such as food or a pheromones causes sensory neurons to release insulin/IGF-1-like DAF-2 ligands. Rather, release of DAF-2 ligands could also be regulated by other cells. Upon activation of DAF-2, cells either produce (as shown in the picture) or repress production of downstream signals or hormones, which acts through an unidentified pathway to aging. c, Modulation of lifespan regulated by reproductive cells. DAF-2 has found to function in somatic gonad signaling. Furthermore, both DAF-12 and DAF-16 has been found to be required for germ-line ablation to extend lifespan. However, it remains unclear if these proteins function in the same pathway or in parallel (as shown in the picture). Assuredly, sensory neurons, the insulin/IGF-1-like hormone, germ cells or somatic gonad cells could activate or repress hormone production. For instance, somatic gonad cells may produce an insulin/IGF-1 like hormone, which in turn may act as an antagonist. (Image adapted from Guarente L., and Kenyon C. (2000))

Autophagy deficient mice show neuron degeneration with accumulation of ubiquitylated protein aggregates, similar as in human ALS patients¹⁵⁹. Downregulation of multiple genes involved in autophagy has been observed during aging and may explain enhanced susceptibility to the toxic effects of protein aggregates¹⁵³. Interestingly, autophagy can also be promoted by increased IIS through FOXO-mediated factors.

Nonetheless, more research is needed to fully unravel these mechanisms. Above stated contradictions emphasise the complexity of the IIS pathway. However, it seems plausible that signals downstream of IIS diverge and might counter proteotoxicity through multiple mechanisms.

TDP-43/FUS AS LINK BETWEEN AGING AND THE CELLULAR STRESS RESPONSE

Another very recent breakthrough came from the result of a research in which TDP1, the *C.elegans* orthologue of TDP43, showed to have a complex role in lifespan and the cellular stress-response via the IIS pathway⁹. TDP43 seems to regulate longevity and the oxidative stress response downstream the forkhead transcription factor DAF16/FOXO3a through IIS receptor activation. In normal conditions, activated DAF-16 stimulates multiple genes that are involved in

prolonged lifespan and enhanced stress resistance¹⁵⁴. This makes them resistant to oxidative, osmotic and thermal stress as well as proteotoxicity¹⁶⁰. However, in response to stress there is DAF-16-dependent upregulation of TDP-43. This initially protects the cell by upregulation of various genes, which combat the effects of stress. Only, prolonged cellular stress showed to cause chronic upregulation of TDP-43, which is neurotoxic and decreases lifespan. Controversially, deletion of TDP-1 has shown to rescue mutant TDP-43 and FUS proteotoxicity in *C.elegans* after a prolonged period of stress. These results imply that even chronic induction of wild-type TDP-1/TDP-43 may stimulate neurodegeneration and reduce lifespan. Whereas TDP-1 has already been associated with proteotoxicity and endoplasmic reticulum (ER) stress, involvement of the IIS pathway may be the missing part to connect them to each other. However, little is known about the separate functions of the components involved into the IIS pathway.

TDP-43 is a constituent of stress granules. This protein arises in response to environmental stress, such as oxidative and osmotic stress, heat shock, oxidative and osmotic stress^{136,139}. Previous research on *C.elegans* investigated if TDP-43 participates in the cellular stress response and longevity pathways. Furthermore, it was researched if TDP-43 controls age-dependent proteotoxicity. Results showed that *tdp-1* is required for the expression of DAF-16 under low IIS conditions (Fig.9)⁹. In case of the presence of oxidative stress, increased TDP-1 protein levels were observed which is dependent on DAF-16. These processes are mediated by a significant altered cellular distribution of TDP-43 upon stress and/or low IIS. These findings suggest that TDP-43 toxicity may activate the ER unfolded protein response (UPR). In turn, experimentally induced ER stress in *C.elegans* increased expression of TDP-1 compared to their untreated controls⁹¹. However, this was only the case in worm expressing wild-type TDP-1 and not in mutants. Similar findings were seen when they investigated FUS⁹, the counterpart of TDP-43 with similar function. Overall, TDP-1 mutants showed the same age-dependent degeneration of motor neurons as TDP-43/FUS mutants. Therefore, a possible loss of function of TDP-43/FUS can cause protein aggregation and cellular stress. This is mediated by the IIS pathway and results in reduced lifespan.

The ER stress response functions through clearance of misfolded proteins, mediated by ER-associated degradation (ERAD). This facilitates transportation of proteins from the ER lumen towards the cytoplasm to become degraded by the ubiquitin proteasome. Only, this mechanism is redox intense and causes a high production of oxidative stress⁹². Moreover, ER stress causes DAF-16 dependent upregulation of TDP-43 expression, which further reduce neuronal function and lifespan. If protein misfolding would be the primary step in neurodegeneration, this may lead to toxicity by: 1) primary toxicity from damaged or misfolded proteins; 2) secondary toxicity from enhanced oxidative stress or lastly 3) toxicity mediated by stress induced TDP-43 expression. Notably, overexpression of wild-type TDP-43 has also been observed in both animals as human ALS patients. Accumulation of this protein in general may therefore actively contribute to neurodegeneration.

Moreover, this model represent the two hit hypothesis for ALS disease in which TDP-43 represents the trigger for the development of pathogenesis⁸². Remarkably, fALS patients mostly harbor mutations within genes that results in aggregate prone proteins. In the contrary, patients with sporadic ALS don't carry those mutant genes and produce less toxic aggregates. This would explain why fALS patients develop the disease earlier in life. Interestingly, the ER stress response showed to be activated in a cell non-autonomous manner. Research found that heat shock

protein-4p (HSP-4p) was mainly expressed in the worms intestinal cells, while mutant TDP-43/FUS appears to be particular expressed in motor neurons of the central nervous system⁹. Therefore, it has been suggested that ER stress is able to signal to other cells and/or tissue types by functioning in a coordinated-organism wide response. Moreover, such a coordinated response has already been identified for mitochondrial stress and the heat shock reponse¹⁶¹.

In conclusion, proteostasis is crucial for survival, but protein control systems fail during aging what lead to accumulation of TDP-43^{154,147}. Thereby, this mechanism becomes positively regulated upon stressful conditions by expression of TDP-43. This results in pathogenesis. However, more research is needed to further elucidate the role of proteotoxicity in longevity mediated by the IIS pathway. This model shows that alternations in signaling can have multiple effects and implies a very complicated role of IIS in ALS. However, the involvement of this pathway in ALS pathology has found strong scientific support, as this will become emphasized in the proceeding chapters.

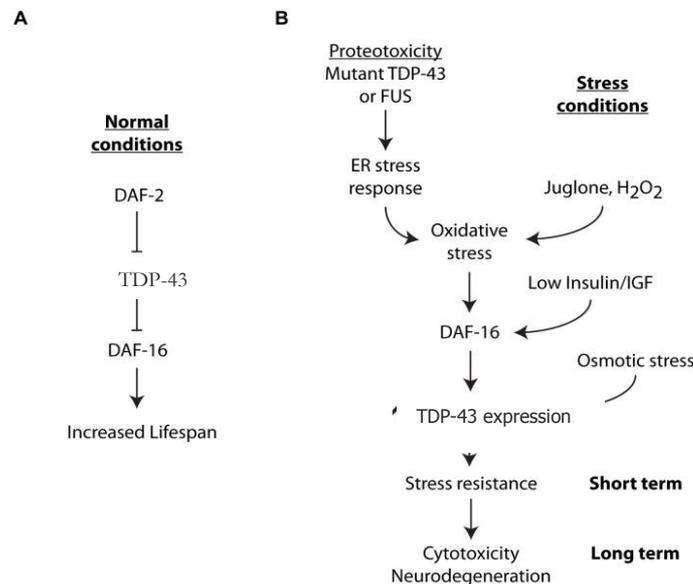


Fig.9: The role of TDP-43 in the IIS pathway. TDP-43 showed to have multiple contributions to lifespan. A. Under normal conditions TDP-43 might function upstream from DAF-16 to regulate longevity (right). B. However, under stressful conditions, TDP43 expression is further induced by oxidative stress and/ or through activation of the IIS pathway. This induction is dependend on DAF-16 and is then regulated in a vicious circle. Higher expression of TDP-43 causes aggregates, which activate the unfolded protein response (UPR). As a consequence, generation of oxidative stress appears. This in turn enhance TDP-43 expression. On the one hand, this mechanism provide stress resistance in acute stress conditions. On the other hand, chronic induction of TDP-43 has negative consequences for the cell, such as neurodegeneration. (Image adapted from Vaccaro, A., et al. 2012)⁹.

ATHLETES

Heavy exercise has also been supposed to be a strong risk factor for ALS since the seventies or eighties¹⁶². Early case control studies tried to unravel the relationship between ALS and sports. However, no clear results were found¹⁶³. Interestingly, Mr. Raffaele Guariniello analysed the cause of mortality of 24000 Italian football players within the period of 1960 till 1996¹⁶⁴. During the course of this study, 375 individuals died, of which 8 due to ALS. The expected number of ALS-caused deaths was 0.69. This indicates a ten-fold higher risk for ALS in soccer players¹³. Typically, these individuals suffered from early onset ALS (in the 40'ties), while mean age of

onset for sALS is around 60-70 years¹⁶⁴. Interestingly, although the study was completed, more participants developed ALS. Nevertheless, this stimulated researchers to further investigate the relationship between heavy exercise on ALS development. In 2005, Chio et al, analysed a large Italian cohort of 7325 male professional football players in the period 1970 till 2001¹³. Since those men were engaged by teams from the First or Second Division they were subject to very intense exercise. During this period five ALS patients were identified with a mean age of onset of 43.4 years. Again this exceeded the expected number of 0.77 patients¹³. In fact, the percentage of ALS cases would have been even higher if 13 participants hadn't been excluded, due to ethnical origin or above threshold exercise. Furthermore, a dose-response relationship has been found between the duration of professional football activity and the risk to develop ALS¹³.

Direct support of the involvement of excessive exercise also derives from the following case. Three men that next to playing soccer on a high level, also grew up under comparable conditions in the same village, developed ALS in the same year. Interestingly, between two of them, the disease progression was similar as well⁴⁰. Therefore, a high level of exercise is most likely the overarching factor, together with the other environmental influences they shared.

Taken all possible contributing factors together, this lead to the following hypotheses: (1) ALS is simply related to participation in sport or heavy physical exercise, and therefore not specifically limited to soccer, (2) Micro trauma, caused by i.g. heading the ball, micro trauma of the legs or muscle lead to an increased change to develop ALS; (3) ALS is related to either the use of illegal drugs or toxic substances to improve performance or to higher dosages of therapeutic drugs; (4) ALS is associated with environmental toxins used on soccer fields. Studies on athletics show an abnormal high amount of individuals with bulbar onset ALS¹³. This could be linked to headers^{13,165}. Though, most individuals develop early onset, which suggest that one or more toxic substances may have triggered ALS in patients with a susceptible genotype⁴⁰. However, it is not known whether and how genetics can interact with environmental toxicants contributing to ALS. Furthermore, no substances have been identified to be involved^{166,167,168}.

Interestingly, recent research on *C.elegans* has discovered involvement of the IIS pathway in ALS, which links the insulin pathway to longevity in ALS. Remarkably, two of the three soccer playing friends, even as a significant amount of football players, showed to suffer from diabetes⁴⁰. Prior to the development of diabetes, high levels of insulin are present due to insulin resistance (IR). Because of IR, the capacity to respond to a given amount of insulin drops. Therefore, higher levels of insulin are necessarily to create the same effects. However, this still needs to be investigated.

To further unravel the direct role of exercise on ALS, follow-up studies have been performed in transgenic mice. However, these studies found contradictory results. Mice which over express wild-type SOD-1 show no clear contribution of exercise on ALS¹⁶⁹, while mutant SOD-1 mice showed that high-intensity endurance exercise accelerate motor degeneration and death, but seemed not to affect onset of ALS¹⁷⁰. In addition, other research showed that low intensity exercise may even is protective by a prolongation of survival in ALS mice. This suggests that the effects of exercise on ALS pathology are dependent on the training intensity¹⁷⁰. Intense physical exercise seems to facilitate molecular mechanisms of oxidative stress¹⁷¹ by the enormous generation of reactive oxygen species^{164,172}. Research found that 15 minutes of high intensity exercise is already enough to cause an increased oxidant production. This causes a rapid, transient reduction in muscle protein thiol content that lead to increased amounts of SOD-1, catalase and heat shock proteins¹⁷². It is suggested that high respiratory activity drives the high production of

ROS during intense activity¹⁷³. Additionally, research on *C.elegans* with knockout of the *clk-1* gene showed reduced respiratory rates and longer lifespan. The *clk-1* gene is required for ubiquinone, which is involved in mitochondrial respiration by affecting the electron transport chain^{174,160,160}. However, this seems to be strictly dose dependent, because severe reduction of respiration appears to shorten lifespan¹⁷⁵. Moreover, reduced expression of strongly conserved mitochondrial genes is observed in organisms ranging from *C. elegans* to humans^{5,176,177}. Research showed that diverse mitochondrial DNA mutations, which causes impaired electron transport chain function (and enhanced ROS), accelerate aging and shorten lifespan¹⁷⁸. Decreased activity within complex I, III, IV and V in of the electron transport chain showed a 40% life extension in *C.elegans*¹⁶⁰. Besides, an increased anti-oxidant function, caused by catalase, seems to prolong lifespan¹⁷³. Although these studies don't reveal the underlying pathways, they do show the importance of mitochondrial function on aging. Motor neurons have a high bio-energetic demand and mitochondrial dysfunction makes them vulnerable to degeneration (as outlined in previous section; oxidative stress). Very intensive exercise may accelerate mitochondrial dysfunction and thereby motor neuron degeneration.

It has now been suggested that a modestly amount of ROS acts as signaling molecule to activate survival pathways, which promote longevity through the induction of stress resistance by regulation of other genes¹⁷⁹. The initial decline of mitochondrial gene expression during brain aging may therefore be part of an compensatory mechanism that increases stress resistance. Additionally, chronic stress may act as an positive feedback cycle stimulating further decline in mitochondrial function which eventually leads to irreversible damage within the cell (Fig.)⁵³.

Besides, excessive ROS also results in a disturbed epigenetic modifications, for instance by the silencing of specific gene-promoters¹⁴. Gene silencing includes a protective mechanism to exclude the expression of DNA damaged regions, rather than undergoing apoptosis. This mechanism may be a transition towards a more repressive transcriptional state¹⁸⁰ in which gene expression becomes totally altered through changes in histone modification patters. SIRT-1 is known to regulate epigenetic gene silencing and has found to be a mediator of the IIS pathway, by regulation of FOXO transcription factors and HSF-1 in response to cellular stress¹⁸¹. SIR1 is able to deacetylate FOXO, which enhances FOXO dependent resistance to oxidative stress and cell cycle arrest, but inhibits FOXO regulated apoptosis¹⁵¹. Therefore, increased SIRT-1 activity seems initially to have a protective role against cellular damage. However, SIRT-1 contributes to the cellular stress response and chronic activation negatively regulates lifespan. Overall, it seems that epigenomic dynamics are conserved during aging and are controlled by oxidative stress. Interestingly, yet another lifespan-regulating transcription factor similar to DAF-16, SKN-1, have been found to modulate longevity by regulation of genes involved in protection against oxidative stress¹⁸². Yet these mechanisms have to be further investigated.

ASSOCIATION BETWEEN ATAXIN-2 AND TDP-43/FUS

As previous outlined, the TDP-43/FUS gene have been suggested to play a crucial role in disease pathogenesis, by regulating longevity and oxidative stress⁹. Moreover, these genes have showed to be the major proteins found in ubiquitinated cytoplasmic inclusions in neurons of patients with ALS and frontotemporal lobar degeneration with ubiquitinated inclusions (FTLD-TDP)^{183,11,44}. However, still little is known about how loss of function or how a (toxic) gain of function, of these proteins would lead to ALS. Until recently, genetic modifiers of TDP-43/FUS were unknown, Therefore, a genetic screen in yeast was performed to define modifiers of TDP-43 /FUS toxicity¹⁸⁴. Ataxin (*ATXN2*), a polyglutamine (polyQ) protein was identified. Notably, it is a dose-sensitive modulator of TDP-43 toxicity within *Drosophila*, Yeast and human^{185,186}.

Furthermore, ataxin-2 has already been associated in different kinds of neurodegenerative diseases¹⁸⁷. Depending on the different polyQ expansions of ataxin-2, ultimately distinct phenotypes develop (Fig.10)¹⁸⁷. For instance, long-length polyQ expansions in this gene causes spinocerebellar ataxia type 2 (SCA2)^{188,189}. Follow-up investigation of polyQ repeats within 900 sporadic and familial ALS patients showed a significant association between intermediate-length polyQ expansions in the ATAXIN-2 gene and ALS pathology¹⁸⁴. Subsequently, the mechanism by which these polyQ expansions might confer pathology became investigated. ATAXIN-2 is normally localised in a diffuse pattern throughout the cytoplasm of spinal cord cells. However, in ALS its localization is altered and moved towards cytoplasmic accumulations.

Both ATAXIN-2 and TDP-43/FUS are RNA binding proteins and showed to be associated in a complex that depends on RNA¹⁹⁰. The intermediate-length polyQ expansion within ATAXIN-2 may enhance ataxin-2 stability or prevent its degradation¹⁹¹. This could cause increased ataxin-2 concentrations, which further stimulate TDP-43 pathology. In turn TDP-43 regulates longevity in interaction with the IIS pathway^{190,184}. Additionally, overexpression of Ataxin-2 lead to increased caspase activity and apoptosis, which causes increased cell death^{192,157}. Upon cellular stress, ataxin-2 enhance caspase activation. In particular caspase 3 showed to cleave TDP-43, which then facilitates and enhances the sequestrates within cytoplasmic stress granules¹⁸⁴. Stress granules contain ubiquitin-modifying enzymes, kinases, proteases and phosphatases^{193,194,195}. Moreover, they seem to be crucial in making the decision whether or not to enter apoptosis as response to extended stress. Under stressful conditions, ataxin-2 is proposed to lower the threshold for TDP-43 phosphorylation^{12,196} and thereby increases ALS pathology.

Although there will certainly be other routes that lead to TDP-43 pathology as well, these data provide strong support for its causative role in ALS pathology. Recent research reported that ALS cases with intermediate-length ataxin 2 polyQ expansions are characterized by distinct TDP-43 pathological features compared to TDP-43 mutations¹⁹¹. Perhaps this distinct pathology is the result of a slightly different mechanism that both result in TDP-43 pathology. Therefore, it is likely that there are multiple independent mechanisms (i.e. mutations in TDP-43, FUS, SOD-1, polyQ repeats), which includes the same target pathways leading to motorneuron dysfunction and eventual degeneration and death¹⁸⁷.

These data have provided important information about the ALS disease mechanism and implies that TDP-43-ATAXIN-2 may be an important genetic risk factor for ALS in both sALS and fALS. Furthermore, this provides crucial knowledge and indications with respect to previous findings about TDP-43 and the IIS pathway.

Effect of ataxin 2 intermediate-length polyQ expansions

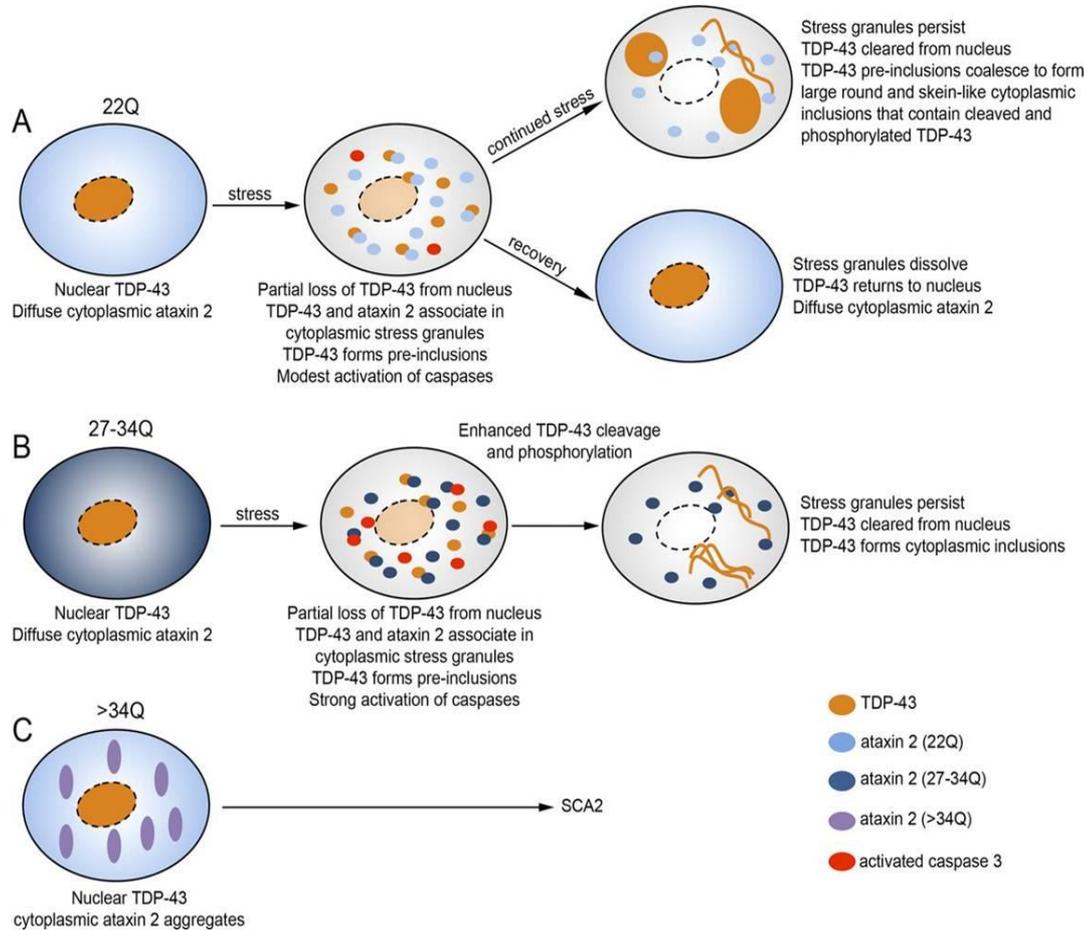


Fig. 10: Effect of ataxin 2 intermediate-length polyQ expansions on TDP-43. This model explains the effect of ataxin-2 intermediate length polyQ expansions on TDP-43. A. During healthy conditions, ataxin-2 is diffusely in the cytoplasm, while TDP-43 is located in the nucleus. Upon cellular stress, TDP-43 is transported into the cytoplasm. TDP-43 and Ataxin-2 both associated within stress granules by activation of caspase 3. After stressful conditions, the cell recovers, stress granules are dissolved and TDP-43 returns to the nucleus again. However, chronic stress results in persistent stress granules and increased TDP-43 clearance from the nucleus leading to large cytoplasmic aggregates. B. Cells that express Ataxin-2 probably have a lower threshold for activation of caspases upon stress. Therefore, increased caspase activity results in increased TDP-43 which eventually lead to enhanced cytoplasmic aggregation. Notably, since the pathology of TDP-43 aggregation may be slightly different than ALS pathology caused without ataxin-2. C. Other evidence from a pathological function of Ataxin-2 resulted from patients with spinocerebellar ataxia 2 (SCA2), which harbour long ataxin 2 polyQ expansions. Aggregation of ataxin-2 in the cytoplasm causes a toxic-gain of function effect that might be unrelated from the normal function. (*This image is adapted from Hart, M.P., 2010*)¹⁸⁵.

DISCUSSION

Research on aging mechanisms have revealed new insights into ALS pathology. Many features which have previously been associated with ALS have showed to be involved into 'healthy' aging processes, albeit in less extreme form (Fig11)^{4,187,154,9}. These features includes, proteotoxicity, oxidative stress, ER stress and RNA metabolism (Fig.12 and Table1; overview). Together, these findings support the idea that premature motor neuron aging, is the common mechanism, underlying ALS

Interestingly, ALS most often occurs in relative healthy individuals: Athletes or individuals without a disease or extraordinary disease history. . Because of the high prevalence among athletics that develop ALS, the influence of heavy exercise on ALS pathology has been investigated. Interestingly, these studies have found a link between heavy exercise and ALS, which is mediated by the aging process^{179,181}. High intensity exercise strongly increases the production of ROS and thereby oxidative stress^{170,173}. This mechanism has previously been associated with the development of ALS^{9,33,9}. Moreover, high cellular ROS concentrations seems to mediate the IIS pathway as well^{197,124}. Besides that heavy exercise is supposed to be a strong risk factor for ALS, these results also validate a causal role of both the IIS pathway and oxidative stress into ALS development^{40,13}.

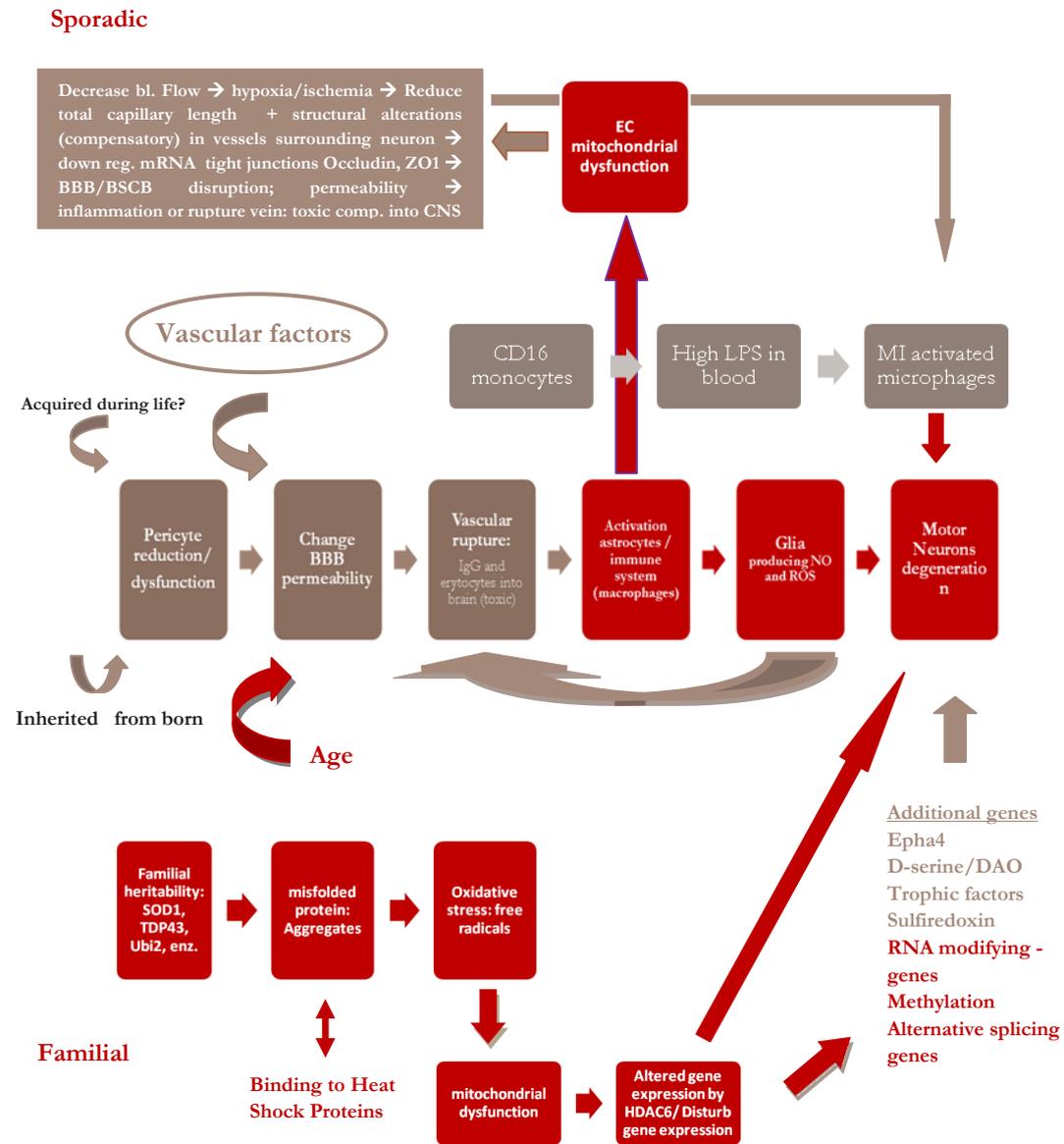
It may be hypothesizes that mutant or excessive SOD-1 in motor neurons cause ALS through a vicious circle. Motor neurons do already have increased levels of SOD-1 to protect them against their own high metabolic demands (see supplementary table.2)¹⁹⁸. It might be that exercise further stimulates the generation of SOD-1. This toxic gain of function negatively impacts motor neuron function, causing further deterioration.

Another remarkable fact is that a significant number of ALS patients have showed to suffer from diabetes prior to onset of ALS^{4,199,40}. Especially diabetes type 2, which is caused by insulin resistance, would be interesting to investigate. Insulin resistance causes the circulation of high levels of insulin in blood. Obviously this can stimulate the IIS pathway excessively. Therefore, this might include the same mechanism in which other components, like TDP-43 or exercise, causes cellular stress and eventually lead to ALS⁹. Consequently, it would be interesting to investigate the occurrence of ALS in diabetes type 1 vs. type 2 patients.

In spite of the evidence that excessive exercise might cause higher prevalence of ALS in athletics, it has also been suggested that trauma underlies the disease^{165,200,201,44}. Although numerous reports have showed evidence of trauma in physically active patients with ALS, the association between physical exercise, trauma and the disease itself is undefined²⁰². Nevertheless recent research showed for the first time that ALS patients show a more frequent history of repeater or severe compared to the normal population²⁰⁰. In particular this is found in headers.

Nonetheless, research on aging, exercise and head trauma is still in its infancy. Detailed investigation into this relationship might provide crucial evidence on the pathology of ALS. Also the specific role soccer and football need further research, since other sports are not (yet) implicated as risk factors.

An overview of features which has been associated with ALS



Disease mechanisms involved into ALS which are associated with aging:	Disease mechanisms involved into ALS which aren't associated
<p>Immune system</p> <p>astrocyte-glia function</p> <p>mitochondrial dysfunction</p> <p>Altered gene expression</p> <p>fALS</p> <p>age</p> <p>RNA metabolism</p>	<p>Pericyte dysfunction</p> <p>neurovascular: BBB and vascular rupture</p>

Figure 11: Overview of mechanisms which have already been identified by the use of iPSCs (red) compared to all mechanisms which has previous been proposed by the use of animal models (gray and red).

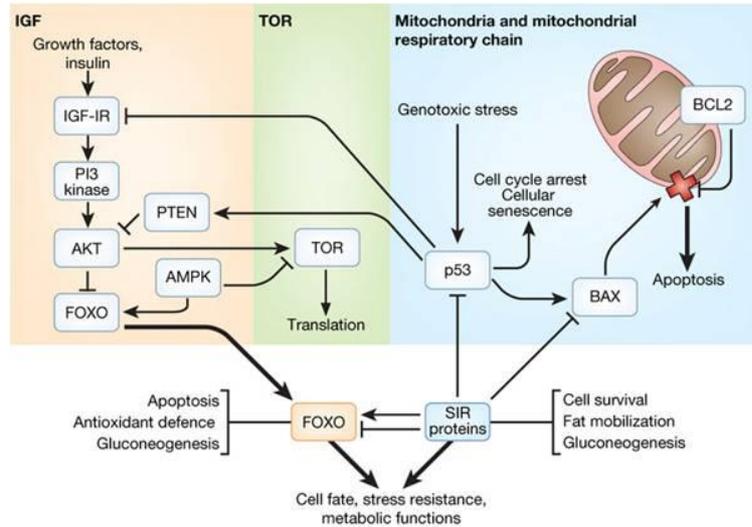


Fig. 12: Integrative aging pathways. These include the three major signalling pathways that reflects the major interactions which are involved into the aging: IIS Pathway (right), TOR (mid) and mitochondrial function (left). Together they regulate the complex interconnections between the IIS – TOR – oxidative stress signalling pathways. Notably, TDP-43 is known to activate FOXO transcription factors , with Ataxin-2 as a possible mediator. (Image adapted from Vijg, J. & Campisi, J., 2008) ¹⁶.

Pathway	Effects on ageing of model organism	Effects on mammalian brain ageing
Insulin/IGF-1 signalling	Decreased signalling promotes increased stress resistance and lifespan	Decreased signalling promotes decreased Alzheimer's disease pathology; paradoxically, increased signalling may be neuroprotective
TOR signalling	Decreased signalling causes increased lifespan, increased autophagy and decreased protein translation	Regulation of autophagy and protein homeostasis may modulate toxic-protein aggregation in neurodegenerative disease
Mitochondrial function	Severely decreased function causes decreased lifespan, but modestly decreased function can cause increased lifespan	Progressively decreasing function during ageing contributes to decline and pathology. However, preliminary evidence suggests that modestly decreased function may engage beneficial pathways
Sirtuins	Can increase or decrease lifespan in different contexts	Can be neuroprotective or detrimental to neurons, depending on context
Caloric restriction	Optimal caloric restriction causes increased lifespan	Increased preservation of cognitive function during ageing

Table 4: Signaling pathways that influence ageing in model organisms and brain ageing in mammals (adapted from Bishop, N.A. 2010)²⁰³.

AGING AND STEM CELLS

In the previous chapter we hypothesized that accelerated motor neuron aging may be the underlying cause of ALS pathology. The IIS pathway showed to be a crucial determinant of lifespan and seemed to be regulated by TDP-43 and (oxidative) stress⁹. To further investigate the influence of aging, there has been a lot of research on stem cells. It has already been shown that tissue and organ aging and/or rejuvenation are closely linked to stem cell function⁷. Aging in stem cells is reflected in their self-renewal capacity. It has therefore been suggested that the process of aging impairs self renewal and differentiation of stem cells, either through alternations that affect them directly or through alternations of their niches²⁰⁴. In this chapter we will investigate the influence of stem cells on aging and rejuvenation by discussing adult stem cells, the stem cell niche and intrinsic aging. Finally, the influence of stem cells on ALS will be discussed.

INTRINSIC AND EXTRINSIC STEM CELL PROPERTIES

An important question when talking about stem cell is which are their properties and how do they change during aging? Knowledge about both intrinsic and extrinsic properties of stem cells contribute to our understanding of how stem cells are able to regenerate and rejuvenate tissues and organs

EXTRINSIC PROPERTIES IN GENERAL

Recent data suggest that extrinsic cues, as the systemic and local organ environment, causes multiple changes during aging, such as decreased activation of organ stem cells, inhibition of repair-specific molecular signaling which together lead to diminished tissue repair¹⁵. Interestingly, there is now evidence that suggests that stem cell based tissue replacement therapies might be rejected in older people. Replaced cells seem to age very rapidly and fail to contribute to organ repair after transplantation of the cells in vivo. Research in animal models has confirmed that the age of the host environment regulates the regenerative capacity¹³⁷. For instance, when skeletal muscle explants were injected into animals, they were only effectively regenerated in young animals²⁰⁵. Furthermore, parabiotically paired young and old mice showed that the regenerative potential of muscle and liver was affected by the age of the systemic environment¹⁵. These data suggest that external factors are regulating the regenerative capacity of stem cells, rather than losing their regenerative capacity irreversibly¹³⁸. Nonetheless, stem cells also showed to be important in the regulation of the replacement of cells that are lost during exercise and aging¹²⁸.

However, during all kinds of regeneration, their response includes a rapidly increased cellular turnover. During aging, this regenerative capacity gradually declines. In muscles, this may be witnessed by muscle fiber atrophy and a general loss of muscle bulk and strength¹⁵. On the one hand, it has been implied that declined stem cell function might be due to intrinsic aging of the resident stem cells. This makes them less responsive to environmental cues. On the other hand, this may be due to an age-dependent decline of environmental signals that regulate these cells²⁰⁶.

Various researches have showed changes in developmental regulatory pathways, which are thought to regulate stem cell function. Pathways that are identified to promote the regenerative responses include: Activation of IGF-1, Notch, or MAPK/pERK pathways and reduction of TGF- β and Wnt signalling (recently reviewed by Conboy, I.A., 2011)^{137,207,208,209}. Most knowledge about stem cell functioning has been gained from the MAPK/pERK pathway. This pathway showed to upregulate Delta and to stimulate Notch signalling. During development, Notch signaling stimulates proliferation and tissue renewal. After stem cell activation, the Notch ligand, Delta²⁰⁷, showed to be upregulated, which lead to increased cell proliferation. Additional research

showed that when Notch inhibitors were injected into damaged muscle, this causes a dramatic impairment of muscle regeneration similar to the declined response seen in aged muscle^{207, 207}. These results showed to be reversed by injection of pseudo-ligands that activate Notch signalling.

In conclusion, recent researchers have found supportive data about the influence of altered environmental cues as cause of declined stem cell regenerative capacity²¹⁰. However, further identification of these cues and their function are needed.

STEM CELL NICHE:

The stem cell niche is the microenvironment in which the stem cells reside. It varies in nature and location depending on the tissue type. Stem cells have their own intrinsic properties, though environmental regulation has shown to be important to direct stem cell proliferation and differentiation²⁰⁴. Studies in *C.elegans* and *Drosophila* have provided important knowledge into the molecular pathways which controls the niche of a stem cell²¹¹. Both, invertebrates and mammals display an asymmetric niche structure. Among cell divisions, one daughter cell stays in the niche as a stem cell, where the other leaves the niche to proliferate and differentiate²¹⁰. The niche is responsible for maintaining homeostasis and for anchoring stem cells, regulated by evolutionary conserved pathways. These pathways include the Notch, Wnt, TGF-B/BMP and Shh pathways²⁰⁴. Evidence for involvement of these pathways shows that activation of developmental regulatory pathways, e.g. IGF-1, Notch or MAPK/pERK pathways, and reduced TGF- β and Wnt signalling, are involved into the regenerative responses in aged tissue (as outlined into the previous chapter).

Interestingly neural stem cells (NSCs) were found in various regions in the adult brain and peripheral nervous system. Here, they regulate the generation of neurons during adulthood. This so called neurogenesis seems to occur in direct contact with blood vessels and astrocytes(Fig.13)²⁰⁴. These stem cells undergo self-renewal and generate daughter cells which become granule neurons²¹⁰. Furthermore, endothelial cells seems to play a crucial role in the NSC niche with respect to the generation of signals that control stem cell self-renewal and lineage commitment²¹⁰. Moreover, various external signals regulate neuronal fate and number, such as BMP and Noggin²⁰⁴. In general, the BMP and Wnt signal pathways seems most involved in self-renewal properties and lineage fate, while activation of Notch seems to be crucial for tissue genesis and for control and maintenance of stem Cells. Recent work suggests that some of these pathways are controlled by micro-RNAs, with special emphasizes on IGF-1 and FoxO3.

Altogether, developmental regulatory pathways seems to regulate stem cell functioning²¹². There seems to be a correlation between stem cell maintenance and general tissue aging. Molecular identification of molecules, which regulates the formation of the aging niches, will broaden our knowledge about the process of aging itself. Interestingly, the neural stem cell niche is in close proximity of blood vessels. If there would be a systemic influence on aging, this might explain the special vulnerability of neurons to aging. Moreover, there are less stem cells in the adult brain compared to other tissues. This might explain why aging-mediated decline in stem cell function damages first the brain. In addition, since NSC niches reside next to capillaries, neurons and glial cells are directly exposed to high insulin and IGF levels. This may lead to enhanced stimulation of the stem cell, which results in earlier senescence. However, it could also be that neuronal stem cells are more sensitive to regulatory signals as a result of increased receptors or an altered binding affinity to those components. However, further research is needed to further unravel this working mechanism.

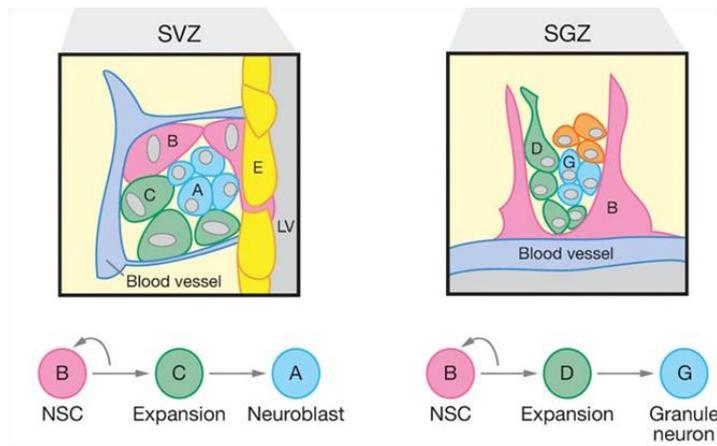


Fig.13: Structural and functional morphology of the neuronal stem Cell Niche. Neuronal stem Cells are always positioned next to a neighboring blood vessel. This is needed to maintain their homeostasis. The numbers in the image each represent: A. neuroblasts, B. Neuronal stem cells, C. expansion of NSCs, E. endothelial cells and LV. the lumen of the vascular blood vessel. (Image adapted from Li, L., & Xie, T., 2005) ¹⁹⁴.

INTRINSIC AGING

As above mentioned, stem cells are pluripotent, which means that they resist factors that do causes age-related changes in replicative or postmitotic progeny in adult stem cells¹²⁸. They behave as a self-renewing population, which may have sustainable periods of quiescence²¹³. Furthermore, they have defense and repair mechanisms²¹³. Continuously renewing tissues, like skin cells, undergo a high number of cell divisions to maintain normal tissue homeostasis²¹⁴. However, each replication cycle contributes to intrinsic aging: shortening of the telomere²¹⁵, chromosomal rearrangements and mutations.²¹⁶ Eventually this may results in cellular senescence²¹⁴. Studies have shown that adult stem cells do have a finite replicative lifespan²⁰⁶. However, this was investigated using experimental manipulations and it remains to be questioned whether replicative aging is enough to cause declined stem cell function in long-lived animals²⁰⁶. Besides, adult stem cells are also subject to chronological aging (when being in quiescence), which takes place in non-dividing cells, such as neurons²⁰⁶. Chronological aging is caused by the accumulation of damaged macromolecules, like proteins, lipids and nucleic acids that cause cellular toxicity²¹⁷.

CELL FATE

Another intrinsic feature of stem cells is their cell fate (Fig.14)²⁰⁶. Based on the tissue, an adult stem cell could be unipotent, bipotent or multipotent²¹⁸. Functional tissue homeostasis and regeneration are regulated by the ability of stem cells to produce a relevant repertoire of tissue specific progenitors. Cell fate is mainly determined by their epigenome²¹⁹. This epigenomic program is influenced by environmental factors and regulated by developmental signaling pathways²²⁰. These pathways seemed to be involved in organogenesis during development and include Wnt, Notch, Hedgehog pathways and TGF β (see for more information the following reviews)^{10,221,209,206}. For instance, a research by Carlson et al., (2009) showed that Wnt signalling restored the youthfulness of cells, whereas increased TGF- β cytokine levels are associated with aging²²². Interestingly, in a study by Conboy, et al., (2005), researchers demonstrate that premature activation, which occurs when young stem cells are placed in an old environment, results in aberrant stem cell progeny¹⁵. Exceedingly, this age related interruption of lineage fidelity can be restored if stem cells are placed in a youthful environment²⁰⁶.

Notably, when stem cells lose their ability to self-renew and/or if senescence pathways become activated, this can lead to the depletion of the stem cell pool²²³. However, this decline could also be a result from loss of self-renewal activity which seems to be regulated by Bmi-1, HMGA2 and the longevity associated transcription factor FoxO3 in mice²²⁴. Bmi-1 and HMGA2 proteins regulate NSC self-renewal by inhibition of the expression of the cell cycle inhibitors, p16INK4a

and p19ARF²²⁵. Interestingly, FOXO proteins, which has previous been associated with ALS, seemed to regulate NSC homeostasis by regulating the expression of cell cycle regulatory proteins as cyclin D1, inhibitor of DNA binding 1 and polo-like kinase 2.

Furthermore, an increased amount of stem cells undergo senescence (growth arrest) during aging, which has a negative impact on tissue homeostasis and regenerative capacity^{226,225}. This seems to be caused by genomic instabilities and/or loss of transcriptional regulation, which affect the accessibility of large chromosomal regions during aging²²⁰. DNA damage is caused by changes in chromatin-remodelling proteins, loss of nucleosome occupancy and redistribution of histone modifying enzymes on the chromatin in old cells compared to young. Moreover, it would be interestingly to investigate whether and to what degree, rejuvenation of old stem Cells¹³⁷ is mediated by restoration of the balance among various chromatin remodelling complexes. This could be performed using the parabiotic mice model. Furthermore, even as in somatic cells, alternations of the proteome can lead to aging as well.

Early removal of improperly folded or damaged proteins is required to maintain normal cellular function⁶⁹. Therefore, autophagosomes, chaperones, lysosomes and the ubiquitin-proteasome system are needed to sense and remove misfolded or damaged proteins in cells⁸².

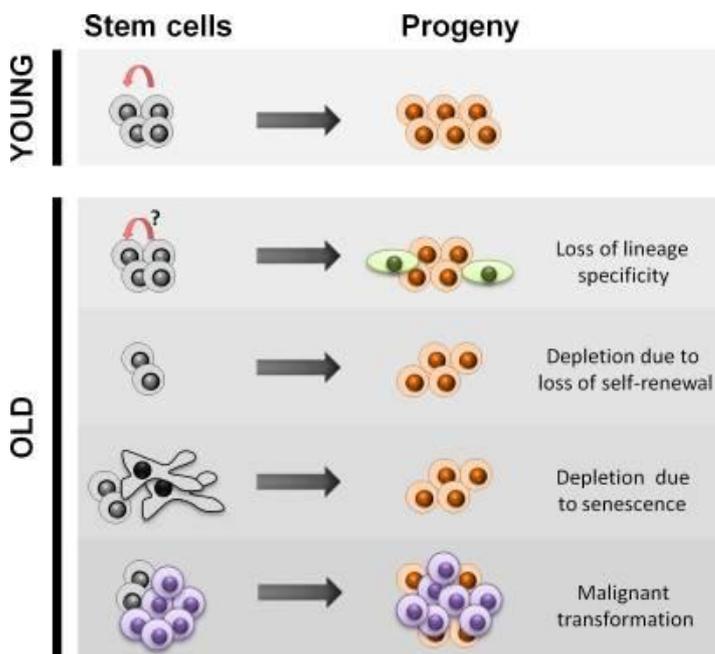


Fig.14: Cell fate. In young organisms, stem cells behave like a indefinitely dividing, self-renewal population. They give rise to lineage specific differentiation during tissue homeostasis or regeneration. However, during lifespan stem cells lose their lineage specificity and contributes to nonfunctional progeny. This results in loss of tissue integrity and a decline of physiological functions. (Image adapted from Liu, L., 2010) ¹⁹⁶.

EMBRYONIC STEM CELLS

Embryonic stem cells (ESCs) are pluripotent stem cells, derived from the inner cell mass of the blastocyst and contain the ability to replicate indefinitely. They are able to differentiate into any other cell type by asymmetric cell divisions. This means that one daughter cell will maintain stemness, while another differentiates into a particular tissue type⁷. Because of their tremendous replication potential, much research is focused on the replacement of damaged tissues and organs. This replacement is mediated by their regenerative capacity and rejuvenation. Cell replacement approaches expand human ESCs and then differentiate them (in vitro). This way patient specific cells can be produced and used to replace their dysfunctional counterparts in vivo. The extent of this method is huge and includes many age-related disorders that are currently incurable. More details about ESCs, and the use of these cells to model ALS can be found in the final chapter; In vitro cell based techniques.

LINK BETWEEN STEM CELL FUNCTION, AGING AND ALS.

Although there are already multiple pathways identified in stem cells and/or in their environment that are associated with aging, these factors do not link the influence of embryonic cells directly to ALS.

It could also be hypothesised that a decreased regenerative capacity of stem cells during aging may be the underlining reason for neurodegenerative late-onset diseases, such as ALS. In this perspective, it may be that the aging process itself causes damage to motor neurons mediated by the IIS pathway. Than it could be a matter of time before the damage will outweigh the protective and regenerative capacity of the motor neurons itself. More damage will accumulate within cells during aging and whenever normal functioning becomes severely impaired this could be the onset of the disease. From that point, a rapid progressive degeneration takes place. The amount of damage increases, while protective and regenerative mechanisms are totally diminished.

This hypothesis has been supported by the discovery of very small embryonic-like stem cells (VSELs) in adult murine tissues²²⁷. Their proliferations seems to be negatively regulated by epigenetic changes of certain imprinted genes, namely *Igf2-H19* locus, *Igf2R* and *RasGRF1*²²⁰. These genes are involved in the IIS pathway. These data support previous findings, in which increased insulin-like growth factor-1 (IGF-1) has been associated with reduced lifespan. Furthermore, a direct link between IGF-1 plasma concentrations, aging and the number of VSELs, residing in adult organ tissues, was found²²⁸. Depletion of these pluripotent stem cells in an IIS-dependent manner causes a decreased number of VSELs and a reduced regenerative potential that directs rejuvenation of tissues²²⁸. Moreover, stressful conditions, such as organ injury, increase VSELs number and enhance their cellular mobility in mice as humans²²⁸. This supports the assumption that they have a potential role in rejuvenation and tissue regeneration. Additional experiments showed that isolated VSELs from young mice were able to expand to all three germ-cell layers both in vitro as in vivo²²⁷, while older mice (>2 years) failed^{229,230}. This observation could either be the result of different paracrine influences or due to epigenetic changes in imprinted genes related to the insulin factor signalling in adult tissues²²⁰.

Adult tissue seems to be able to activate a quiescent VSEL state during aging. This cellular mechanism may have been evolved as protection against spontaneous growth of teratomas²²⁰. Research has found that VSELs downregulate Insulin growth factor-2 (*Igf2*) expression. *Igf2* is highly expressed during the embryonic development and causes upregulation of the IGF-2 receptor (IGF-2R)²³¹. Subsequently IGF-2 is able to bind Insulin growth factor-1-receptor (IGF-

1R). However, as the result of epigenetic alterations during aging, VSELs show a decrease in IGF-2 and RasGRF1 expression and an overexpression of IGF-2R²³². This causes quiescent in adult tissues. Nonetheless, chronic exposure to IGF-2 or insulin seems to accelerate premature depletion of VSELs. To further investigate involvement of the IIS pathway in stem cell functioning, more research was performed using the dwarf mice as model²³³. Dwarf mice harbor a deficiency of growth hormone (GH) receptor, and as a consequence of this deficiency they show a reduction in IGF-1 plasma levels as well. Their number of VSELs showed to be 3-4 fold higher than in normal mice during aging even as their average lifespan²²⁸. This confirms the influence of the IIS pathway on VSELs appearance.

Interestingly, also calorie intake has previous been associated with the IIS pathway, by regulation of IGF-1 levels in serum¹⁴⁵. Now, it has been found that high calorie intake causes a reduced number of VSELs during aging²²⁸. This confirmed that high IGF-1 plasma levels (either autonomous regulated or influenced by calorie intake)²³⁴ may cause premature depletion of stem cells and may thus be the cause of premature aging.

Above findings show, that the rejuvenation and the regenerative capacity of VSELs can be influenced by IGF concentration. Interestingly, chronic exposure to it causes premature depletion in these stem cells²³². During aging exactly this happens. IGF concentrations are high and therefore can directly influence the regenerative capacity of VSELs. Since the same pathway is implicated in ALS, it might be possible that the loss of VSEL regeneration diminishes the regeneration of MN as well. Therefore it might influence the pathology observed in these patients.

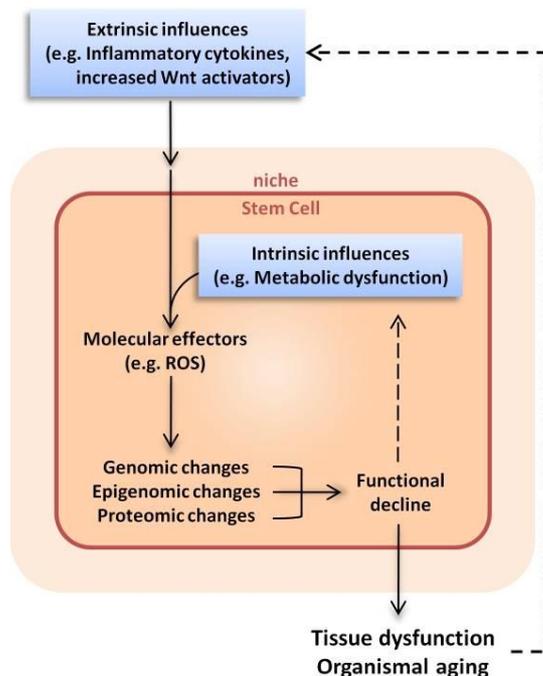


Fig.15: Extrinsic and intrinsic influences on stem cell function. Age-related changes in the systemic milieu might result in altered signaling cascades or molecular changes within the cell. This can be caused by a change in the systemic blood circulation or by changes within the stem cell niche. In contrast, intrinsic changes do also appear during aging, such as mitochondrial dysfunction. These signals might integrate and influence downstream molecular effectors. This causes genomic, epigenomic and proteomic changes in the cell, and subsequently lead to decreased cellular functioning. Decreased stem cell function exacerbates the extrinsic influences of aging. Thereby they contribute to aging. (Image adapted from Liu, L., 2011) ¹⁹⁶.

DISCUSSION

Stem cell research has provided important knowledge about aging and rejuvenation¹³⁷. Diverse studies have found an association between the IIS pathway, aging and stem cell function^{228,7}. This again confirmed involvement of the IIS pathway in aging. These findings might have important implications for ALS research.

Recent studies suggest that neurodegenerative diseases may be dependent of the rejuvenation and regenerative capacity of stem cells^{15,138,206}. During aging, this rejuvenation and regenerative capacity of stem cells declines²⁰³. This decline may be the result of environmental changes²¹². Research on parabiotic mice showed that the regenerative potential of organ function was affected by the age of the systemic environment¹⁵. These data suggest that external factors are regulating the regenerative capacity of stem cells, rather than losing their regenerative capacity irreversibly¹³⁸. This implies a systemic role on the regulation of stem cell function²³⁵.

Parabiotic studies have showed the need of increased IGF-1, Sonic Hedgehock, Notch, MAPK/pERK pathway signalling, and a reduced TGF- β and Wnt signalling, during regenerative responses^{212,222,236,237,238}. Moreover, researchers have found atrophy of the motor cortex, termed motor cortex thinning, in ALS patients²³⁹ (validated by recent research; preliminary data). Interestingly, a fundamental study into the working mechanism of TGF- β 1 by Piras F. et al. (2012), has discovered a correlation between TGF- β 1 serum concentrations and cortical thickness. In healthy individuals, high serum levels of TGF- β 1 has been associated with increased cortical thickness in the cingulate and right frontal cortex²⁴⁰. This implies that a reduced serum level of TGF- β 1 might be involved into ALS pathogenesis. However, this result is in contradiction with the above mentioned studies, in which reduced TGF- β seems to be required for regeneration of cells and tissues^{222,209}. Therefore, more research is needed to unravel the mechanism behind motor cortex thinning and its development, to exclude if this is a cause or a consequence of ALS.

Interestingly, recent research discovered that signals from the systemic environment drive age-related, intrinsic changes in oligodendrocyte precursor cells that cause demyelination of neurons which leads to demyelinating diseases such as multiple sclerosis (MS)¹³⁷. Although this finding involves another type of neurodegeneration, it supports the hypothesis that the systemic environment might be able to drive age related changes leading to ALS. Moreover, previous research has observed increased mobility of stem cells in response to damage²⁴¹. This makes us questioning if the presence of stem cells in the skin or intestine could have an influence on ALS pathology (e.g. damage) in the brain? Besides the question about the intrinsic function of stem cells, it would therefore be interesting to further investigate to what extend environmental factors contribute to ALS pathology. Consequently, if systemic factors would be able to influence the regenerative capacity of stem cells, they might enhance the regenerative capacity of somatic cells as well. Nevertheless, additional investigation have to be done to identify the exact location of embryonic stem cells in the brain, since their effect may only affect the local environment.

To further investigate stem cell function in relation to aging and ALS, various aspects have to be considered. For instance: What is the role of stem cell rejuvenation and regenerative capacity on damage induced by oxidative stress and proteotoxicity as caused by ALS? In which way does oxidative stress and proteotoxicity interfere or reinforce stem cell function in relation to the aging process? Oxidative stress and proteotoxicity can either change environmental influences or they can cause direct damage to stem cells or their niche (Fig.15)²⁰⁶. To summarize, it is not clear yet to which extend stem cell function is involved in ALS onset and disease progression.

In conclusion, age-related changes seem to be determined by the age of the systemic environment, rather than by the cell-autonomous age²⁰⁶. To further investigate these systemic influences and or environmental influences that contribute to a declined stem cell function and/or ALS pathology during aging, a new promising technique, the iPSC model, could be used to obtain a deeper understanding¹³¹. In addition, these models could then also be used to investigate, whether and to what degree, rejuvenation of old stem cells is mediated by restoration of the balance among various chromatin remodelling complexes, since they have showed to regulate activation of the IIS pathway¹³⁷. In the next section a new ALS model will be introduced, which can be used to investigate the influence of aging. Consequently, the final chapter will suppose a future research suggestion.

IN VITRO BASED THERAPEUTIC STRATEGIES

Previous chapters have suggested a causative role of aging on the development of ALS. To confirm this, more research is needed. Yet, tractable models to test this hypothesis in human neural system are scarce. Much knowledge into the pathophysiology of ALS is obtained from animal models, which displays familial disease forms of ALS. They have been implemented by the use of highly overexpression of mutant genes, such as SOD-1⁶⁰. However, more than 90% of ALS patients suffer from a sporadic forms²³ due to complex interaction between genetics and environmental factors. Therefore, these models may not completely mimic the actual disease¹⁰⁷. Moreover, a complete recapitulation of ALS is not possible, because of fundamental genetic and anatomical differences between animals and human. This became particularly clear after repetitive failure of translating animal data to the clinical settings²⁴. Therefore, new models are needed.

Novel stem cell technologies offer exciting new opportunities to study human neural development and disease in the laboratory. They may provide the first feasible models available to investigate the disease mechanism for sALS by utilizing induced pluripotent stem cells (iPSCs)¹⁸. Nowadays, the challenge is to evaluate differences and similarities between fALS and sALS. This chapter will introduce the concept of human embryonic stem cells, induced pluripotent stem cells and their purposes.

HUMAN EMBRYONIC STEM CELLS (hESCs)

Human embryonic stem cells were discovered in 1998 and afterwards used for various purposes^{242,243}. They have the advantage that a human-situation can be investigated directly, without further manipulation. Therefore, hESCs would be the perfect model to investigate diseases. However, this technique uses human embryos. Of course this raises several ethical concerns and hinder their application. iPSCs resemble the use of embryonic stem cells (ESCs) in that they are both pluripotent and share the ability to replicate indefinitely. Since there are no ethical concerns with respect to iPSC, this method has obtained increased interest over the past ten year. Though hESCs are still important to investigate fundamental questions and basic biology, but also to further optimize iPSC modelling by unravelling the intrinsic properties of pluripotency and differentiation²⁴⁴. Therefore, this model is needed for the development and optimization of other in vitro cell based methods, like iPSCs, for good disease modeling.

HUMAN INDUCED PLURIPOTENT STEM CELLS (HUMAN iPSCs)

iPSCs are adult somatic cells that have been genetically reprogrammed to an embryonic stem cell¹⁹. They provide enormous potential to model ALS because of their competence to developmental signals that permits their specification of functional, regional subtype-specific neurons²⁴. In the case of ALS, this includes the directed differentiation of spinal cord, midbrain and motor cortex neurons²³⁷. Studies have showed that this technique reliable recapitulates the spatio-temporally regulated, developmental responsiveness to specific extrinsic morphogenetic signals. Subsequently, the next challenge is to understand how these refined subregion specific human neuronal and glial diversity is produced. There are multiple motor neuron subtypes, each with different disease vulnerability²⁴⁵. In addition, currently in vitro based stem cell models are already quite competent to do this²⁴⁶. Furthermore, iPSCs enables the investigation of patient specific lines that includes disease causing mutations, which is not able with hESCs²⁴⁷.

The first iPSC, generated from fibroblasts, has been discovered in mice in 2006 and in human in late 2007^{248,249,249}. These patients'-derived iPSCs carry the exact package of genetic information that is associated with patients' pathology. Therefore iPSCs allows the study of living cells and

provides insight into their intrinsic properties, their susceptibility to environmental influences and their interactions with other cell types²⁵⁰.

However, the process of reprogramming is low (typically 1% of transfected fibroblasts change into a iPSC) and often incomplete^{248, 248}. It has been suggested that numerous cells undergo the process of reframing, but not all finish the process completely. However, it is beyond the scope of this review to provide an overview of the many studies which have investigated this (more details are provided by Papapatrou, E., 2009)²⁵⁰. In contrast, one of the main advantages of the use of iPSC is its simplicity and reproducibility²⁴⁸. It has been found, that only four transcription factors (TF), namely Oct4, Sox2, Klf4 and c-Myc, are needed to induce pluripotency²⁵¹. This can be performed on both terminally differentiated or postmitotic cells^{250, 252}. Furthermore, it has been hypothesized that all somatic cells have a potential to become iPSCs, although with different efficiencies²⁵³.

Another point of debate is whether differentiation of iPSCs are functionally different from ESCs²⁵⁴. In vitro directed neural differentiation showed that more than 90% of ESCs became successfully differentiated, compared to 10-50% of iPSCs²⁵⁵. Though, other research found much higher differentiation efficacies in iPSCs, which again emphasizes the conflicting results regarding the similarity between those cells¹⁹. Besides reprogramming efficacy, the reprogrammed cells of iPSC have showed to be similar to ESC in their morphology, expression of marker genes and their ability to form teratomas (tumours comprised of cells from all three germ layers)²⁵⁴. Transcriptomics have shown a more fetal state of the neurons rather than an adult one²⁵⁶. This may limit the modelling of the disease. However, research confirmed that human iPSCs use the same transcriptional network to obtain specific cell types. This takes place over the same developmental time period as hESCs, mediated by the same morphogenesis (i.g. morphogenetic protein (BMP), Sonic Hedgehog (Shh) and Notch)²⁵⁴.

Moreover, the biggest question still remains if iPSCs, besides the above-mentioned features, are exactly similar to ESCs with respect to epigenetic reprogramming and cellular memory? Research have showed contradictory results, some have found persistent expression of donor cell genes, while others found differences in DNA methylation pattern and or epigenetic memory, while others haven't found any differences at all^{257, 258}.

IPSC DISEASE MODELLING OF ALS PATHOLOGY

IPSC technology is an interesting technique to further investigate the ALS pathology. The recent discoveries that emphasize a major role for IIS-pathway-mediated aging and rejuvenation on motor neuron degeneration had made the use of former disease models difficult. Nonetheless, iPSC technology can be used to investigate these disease mechanisms²⁵⁹. Although, the question if induced pluripotent stem cells can be produced directly from elderly patients with ALS remains. Their cells have been exposed to disease causing components all their life. However, recent data showed successful results of motor neurons, generated from iPSCs from skin fibroblasts from a 89 and 82-years old patient who suffered from fALS²⁶⁰. These patients were among the oldest living individuals with mutant SOD1 alleles. This makes us questioning about the role of SOD-1 in disease pathology, since inherited mutations usual causes early onset of disease. However, more research is needed to further investigate this.

One of the key challenges of iPSC disease modelling is assess to what extend fALS and sALS differ from each other¹⁰⁷. For instance, recent research has investigated if astrocytes release neurotoxic factors which affect motor neuron degeneration by the use of post mortem spinal

cord neural progenitor cells of both sALS and fALS patients¹⁰⁷. Motor neurons which were treated with sALS and fALS conditioned media showed a 50% higher reduction of motor neurons compared to non-ALS controls²⁶¹. This glial mediated toxicity has showed to be similar in sALS and fALS and again confirm a systemic or environmental role on motor neuron degeneration. Furthermore, a human iPSC model of TDP-43 proteinopathy in motor neurons has showed that mutant TDP-43 causes increased soluble and detergent resistant cytoplasmic TDP-43 levels, which led to decreased lifespan under basal conditions. Interestingly, mutant TDP-43 motor neurons seems to be more susceptible to PI3K inhibition, which is a crucial signalling pathway in neurons²⁶². However, this susceptibility showed to be independent of the MAPK/ERK pathway or the induction of ER stress. Notably, both pathways are part of the IIS pathway, which may link this observation to aging⁹. Yet another study has showed that TDP-43 protein mediated toxicity appears in both sALS and fALS²⁶³. ALS patients specific iPSCs which harbour mutant TDP-43 have showed to be more vulnerable to cellular stressors compared to non-ALS control patients²⁶³. They also turned to be more sensitive to oxidative stress which resulted in an increased amount of insoluble TDP-43 and thus enhance aggregation²⁶⁴. Moreover, they found various altered gene expressions.

iPSCs could also be used to obtain insight into both development and disease of motor neurons at the RNA level, with high throughput approaches, such as exome- and RNA-sequencing^{120,24}. These techniques have showed various changes in gene expression and alternative splicing events (i.e. C90RF72 gene)²⁴. In addition, another research created iPSC lines from patients with mutations in the VAPB gene²⁶⁵, another gene that has recently been associated with ALS, as well as from non-ALS controls. Reduced level of VAPB protein were observed in patients with mutant VAPB gene compared to controls⁵⁴. Subsequently, they showed a gradual increase of VAPB during cellular differentiation in controls, but this was not found in patients with mutant VAPB. This implies that mutant VAPB causes a failure of VAPB protein upregulation during the induction of motor neurons²⁶⁶. This is likely due to posttranslational processing of VAPB, since there has no mRNA level differences been found between control and mutant VAPB patients²⁶⁷. Moreover, posttranslational processes have already been identified, though this has not been intensively investigated yet.

In general, iPSCs have showed major potential for disease modelling. Therefore, further research has to be performed to further identify ALS disease mechanisms and to discover candidate drugs for future therapy(For an overview see Table.1) ²⁶⁴.

Disease	Genetic defects	iPSC derived cell types	Disease phenocopied in iPSCs or differentiated cells	Drug or functional tests	Reference
ALS	Mutation in SOD-1	Motor Neurons	NA	No	268, 269, 261
	Mutation in VAPB (vesicle-associated membrane protein)-associated protein B	Motor neurons	Yes	NO	266
	Mutation in TDP-43/FUS	Neurons and motor neurons	Yes	Yes	270, 263

Table.1: Overview of all mechanisms which has already been found in ALS using iPSCs

DRUG SCREENING AND DEVELOPMENT

Besides elucidation of ALS disease pathogenesis, motor neurons produced from patient derived iPSCs may provide also an important tool for screening of drug candidates^{264,270}. iPSC serves as a platform to discover novel compounds by making use of molecular dissection of the disease process²¹⁸. Currently, this technique enables to test for efficacy and toxicity of small compounds in patient specific populations. It can now exactly be determined which drug or which drug combinations are effective in human patients^{218,132,21}. However, the molecular and cellular phenotypes first have to be identified. Subsequently, the generation of high quantities of specific neurons or glial cells enables high throughput drug screening²⁷¹. On this moment, ALS patients specific iPSCs which harbour mutant TDP-43 have showed to be more vulnerable to cellular stressors compared to non-ALS control patients²⁶³. They also turned to be more sensitive to oxidative stress which led to an increased amount of insoluble TDP-43 and thus the formation of aggregates²⁶⁴. Moreover, they found various altered gene expressions. However, further research has to be performed to further identify candidate drugs for future therapy²⁶⁴.

CELL THERAPY

Generation of iPSCs from ALS patients allow the production of large numbers of immune matched cells. This technique enables cell replacement and neuroprotection strategies in patients with ALS. Major results have already been observed with rodent and human iPSC derived neurons in various neurodegenerative diseases, such as in Parkinson's Diseases^{272,273}. Besides, glial cells could be used to support cells, rather than as replacement of motor neurons^{274,218}. However, in spite of the major potential of iPSCs, several challenges have to be overcome before cell therapy could be clinically applied²¹⁸.

Firstly, integrating viral vectors, which carry oncogenic genes, like retroviruses, are used to reprogram cells. However, this causes an increased risk to develop cancer. However, currently research has already developed new mechanisms to reprogram cells, without the use of viral vectors^{238,275}, like the use of microRNAs or transcription activator-like effector nucleases (TALENs).

Secondly, purification of the desired cells, by differentiation of iPSCs to specific cell types is required for objective measurements. This will be established by increasing the understanding of directed differentiation and to optimise or develop improved protocols, which eventually enables highest possible cell specificity. High cell specificity is needed to unravel the exact disease mechanism, but this also lead to a better immune matched cell for future cell transplantations²⁷⁴.

Thirdly, the exact disease mechanism, of each genetic defect, has to be unravelled before they can be used for cell therapy. This is the hardest point to challenge with respect to ALS. Intrinsic pathogenic mechanisms of ALS may be delayed, due to the relative short lifetime of iPSCs. Since ALS is the result of a lifetime accumulation of disease pathology, iPSC derived motor neurons and glial cells are conducted to external stressors to facilitate late-onset ALS phenotype²⁷⁶. As stressor, l-glutamate-induced excitation of iPSCs derived neurons are often used to generate the ALS phenotype^{264,277}. This causes Ca^{2+} dependent proteolysis, which results in micro aggregate formation. However, the question remains how relevant these iPSC neurons actually are for the investigation of ALS pathology. Notably, heavy exercise has been supposed to be a strong risk factor for ALS by facilitation of oxidative stress and ROS production^{171,164,172}. A similar principle could underlie the effect of l-glutamate-induced excitation of iPSC-derived neurons.

Finally, the main goal of cell replacement therapy in ALS disease is to replace degenerated (motor) neurons. Yet, to make them functionally similar to autonomous (motor) neurons, they have to acquire the ability to extend their projections to the proper target structures. This is often mediated by non-cell autonomous signalling regulated by glial cells²¹⁸. Therefore, glial cells need to be taken into account as well. However, further research should be done to further investigate this complexity and to overcome these challenges.

DISCUSSION

The discovery of iPSCs offers very exciting and unprecedented opportunities for the use of ALS disease modelling^{264,132,263}. This technique has to potentially decipher the molecular and cellular mechanisms underlying ALS pathology. We hypothesised that accelerated motor neuron aging may be the underlying cause of ALS pathology. iPSCs now enables investigation of aging on the development of ALS in humans.

This technique has several important advantages. Firstly, iPSC technology is the first method, which enables investigation of sALS instead of fALS (Fig.16). Besides, iPSCs derived motor neurons could be co-cultured with human derived glial cells as well^{278,269}. Secondly, iPSCs are efficiently converged to neural lineages by the establishment of different protocols. This allows the investigation of early progenitors that were first inaccessible in patients. Lastly, this technique enables the production of a big number of cells that can be used for cell-based toxicity screening and future therapy. This can close the gap between animal studies and clinical settings²⁷⁹. iPSCs may even have the potential to establish personalized drug development using high-throughput screening²⁸⁰. Though, several challenges have to be overcome before iPSCs can be clinically applied, which mainly includes safety issues and differentiation specificity. However, due to the many developments in this field in recent years, these challenge will certainly be overcome in the near future.

In conclusion, iPSC technology offers huge potential for the investigation of ALS pathology. iPSCs are live functioning, human, patient derived specific cells, which will give us insights into disease mechanisms, drug discovery, cell therapy and a potential new diagnostic method for patients with ALS^{19,20,21}. It would be a very tempting purpose to further investigate the influence of IIS-pathway-mediated aging and rejuvenation on motor neuron degeneration.

Experimental set-up

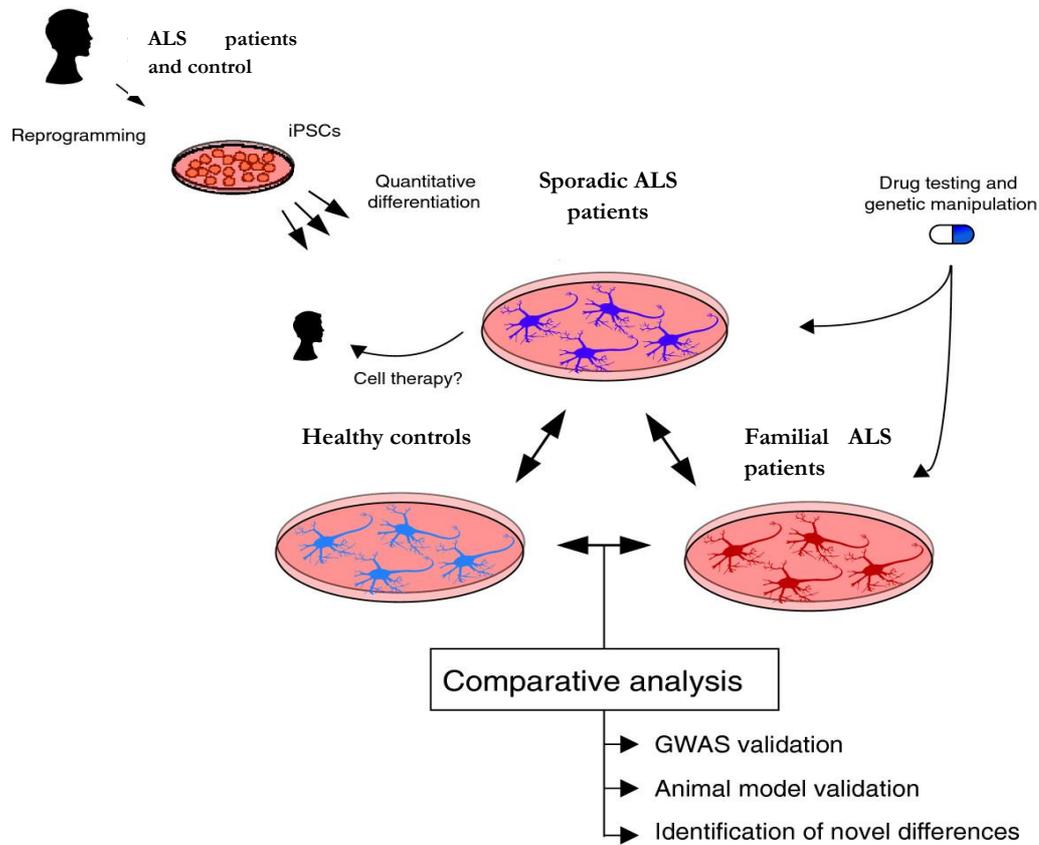


Fig.16: iPSC potential in ALS disease. Patient derived iPSCs could be differentiated in motor neurons and used for disease modeling and drug screening. This technique enables the investigation of disease pathology of sALS in comparison to the disease pathology of fALS. These results could be verified by the use of Genome Wide Association Studies (GWAS) and animal models.

PROPOSAL FOR FURTHER RESEARCH

Recent research strongly supports that accelerated motor neuron aging may be the underlying cause of ALS pathology⁹. Data have showed that age-related changes seemed to be determined by the age of the systemic environment, rather than by the cell-autonomous age¹⁵. Therefore, it has been suggested that signals from the systemic environment drive age-related, intrinsic changes, mediated by the IIS pathway. This activates a cascade of events that eventually can lead to ALS pathology. However, current animal models are not suitable to further investigate this postulation. Not only did the obtained results fail when translated to the human situation, also are fundamental genetic and anatomical differences between both species in age-related pathways found^{16,17}. Therefore, new and improved models are needed.

As stated in the previous chapter, *in vitro* cell based therapies seems to be the first feasible model to investigate the disease mechanism for both sALS and fALS by utilizing induced pluripotent stem cells (iPSCs)^{18,19,20,21}. Therefore, it would be a very tempting purpose to use this iPSCs to investigate the influence of IIS-pathway-mediated aging and rejuvenation on motor neuron degeneration.

Studies on parabiotic mice, have already indicated a major influence of the environment on aging^{7,137,138,212}. Parabiotic mice are mice that have been sutured together, side-by-side, along the lateral aspect of the body. Once blood vessels have connected with each other, the mice will share a common blood supply. This model has been used to determine whether a blood-borne mediator from one, affects the effect of aging of the other. Therefore, heterochronic (young and old) pairs were compared with isochronic (young to young or old to old) pairs. In the study of Conboy, et al. (2005), researchers demonstrated that exposure of young serum to old mice increase the rejuvenation and regeneration capacity of the old mice. Likewise, this research showed that effects of aging on regeneration capacity are reversible. Moreover, these age-related changes seemed to be determined by the age of the systemic environment, rather than by the cell-autonomous age. Notably, when young serum became mixed with old serum, the regenerative potential was inhibited, suggesting a dominant influence of old serum factors⁷. Further research on which systemic factors might influence aging, found that increased IGF-1, Notch, MAPK/pERK pathway signaling or reduced TGF- β or Wnt signaling, increases the regenerative responses in aged tissue^{222,207,206}. Interestingly, recent research by Conboy et al. (2011) on multiple sclerosis showed that signals from the systemic environment drive age-related, intrinsic changes in oligodendrocyte precursor cells⁶. This process seems to underlie demyelination of neurons, which leads to multiple sclerosis (MS)¹³⁷. This supports the above mentioned hypothesis, suggesting that the systemic environment is able to drive age related neurodegenerative diseases. Hence, it would be very interesting to identify which systemic factors might contribute to the influence of aging on ALS pathology.

To accomplish this, iPSCs-derived motor neurons from sALS and fALS patients can be cultured in serum from younger non-ALS individuals (Fig.17A). As a control 'old' non-ALS patient derived iPSCs can be used. On the other hand, old serum from both sALS and fALS patients will be exposed to motor neurons, which are derived from non-ALS individuals. This will be compared with iPSCs from sALS and fALS patients as a control (Fig.17B). Since a non-autonomous cause has been proposed, the same experiments can also be performed with iPSCs-derived motor neuron and glial cell co cultures. Additionally, to further investigate the influence of the IIS pathway, an experiment that uses insulin/IGF blockers and agonists can be performed (Fig.17C). The same approach can also be used to investigate the effect of oxidative stressors or if being a diabetic functions as a risk factor.

Furthermore, this model can be used to investigate the role of stem cell functioning on the development of ALS as well. Research showed that human stem cells are responsible for rejuvenation and regeneration of tissues cells. It could be hypothesised that a decreased regenerative capacity of stem cells during aging may be the underlining reason for neurodegenerative late-onset diseases, such as ALS. In this perspective, it may be that the aging process itself causes damage to motor neurons mediated by the IIS pathway. To investigate this, co-cultures of postnatal stem cells with motor neurons of different maturity from the above-mentioned patients would be exposed to the different sera (Fig.17D).

Considering these aspects, it would be a unique approach to investigate the influence of aging on ALS, since this would be the first experiment, on this subject, that uses human derived cells. Importantly, research into the effect of aging on human neurodegenerative disease would have clear implications for future therapies as well.



Experimental set-up

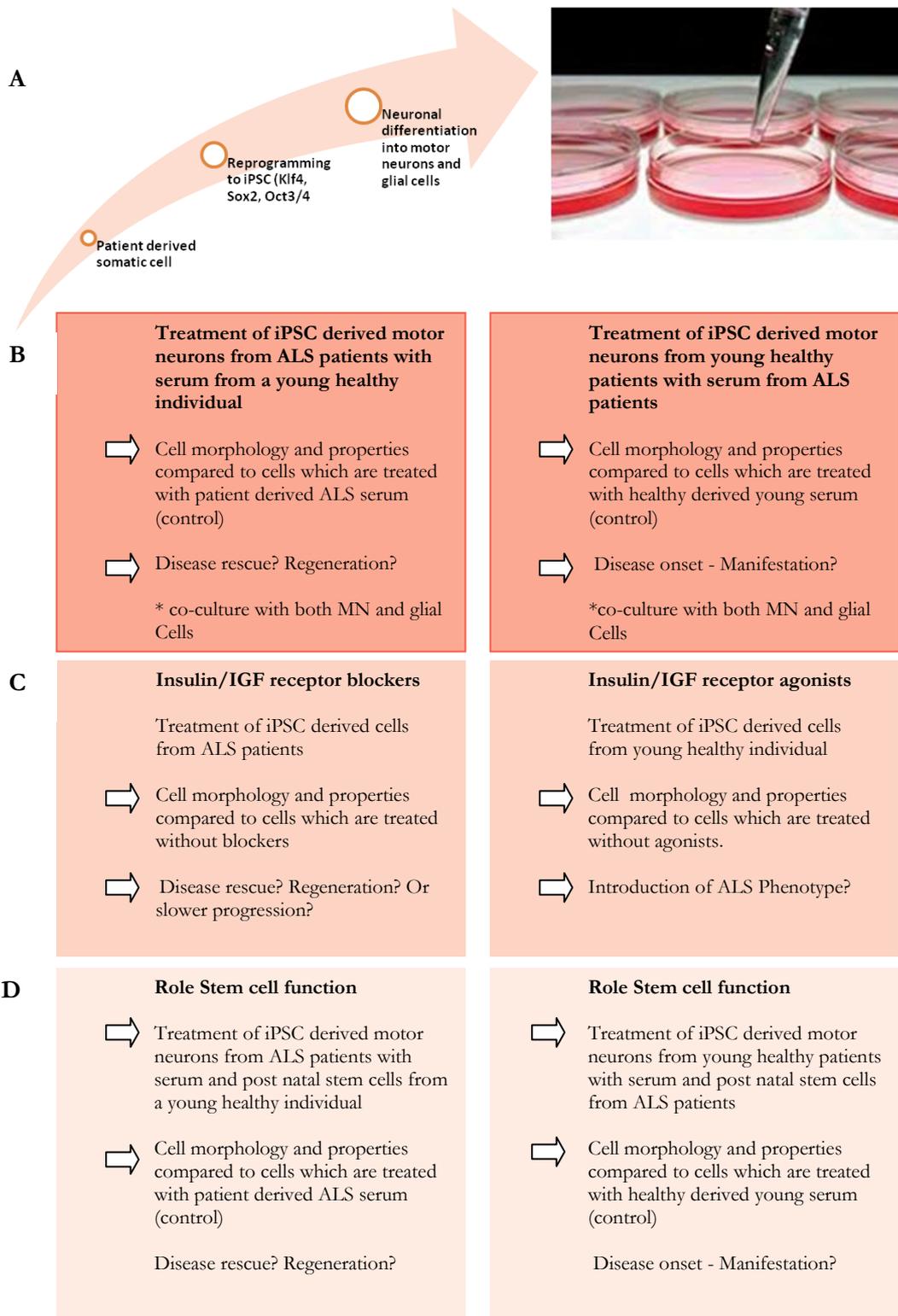


Fig.17: Proposed future experiments

BIBLIOGRAPHY

1. Venkova-Hristova, K., Christov, A., Kamaluddin, Z., Kobalka, P. & Hensley, K. Progress in therapy development for amyotrophic lateral sclerosis. *Neurology research international* **2012**, 187234 (2012).
2. Andrew Eisen MD, F. Amyotrophic lateral sclerosis: A review. *BCMj*, Vol. 44, No. 7 at <<http://www.bcmj.org/article/amyotrophic-lateral-sclerosis-review>>
3. Acevedo-Arozena, A. *et al.* A comprehensive assessment of the SOD1G93A low-copy transgenic mouse, which models human amyotrophic lateral sclerosis. *Disease models & mechanisms* **4**, 686–700 (2011).
4. Cohen, E. & Dillin, A. The insulin paradox: aging, proteotoxicity and neurodegeneration. *Nature reviews. Neuroscience* **9**, 759–67 (2008).
5. Yankner, B. A., Lu, T. & Loerch, P. The aging brain. *Annual review of pathology* **3**, 41–66 (2008).
6. Ruckh, J. M. *et al.* Rejuvenation of regeneration in the aging central nervous system. *Cell stem cell* **10**, 96–103 (2012).
7. Carlson, M. E. & Conboy, I. M. Loss of stem cell regenerative capacity within aged niches. *Aging cell* **6**, 371–82 (2007).
8. Kenyon, C. The first long-lived mutants: discovery of the insulin/IGF-1 pathway for ageing. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences* **366**, 9–16 (2011).
9. Vaccaro, A. *et al.* TDP-1/TDP-43 regulates stress signaling and age-dependent proteotoxicity in *Caenorhabditis elegans*. *PLoS genetics* **8**, e1002806 (2012).
10. Kim, S. H., Shanware, N. P., Bowler, M. J. & Tibbetts, R. S. Amyotrophic lateral sclerosis-associated proteins TDP-43 and FUS/TLS function in a common biochemical complex to co-regulate HDAC6 mRNA. *The Journal of biological chemistry* **285**, 34097–105 (2010).
11. Baloh, R. H. TDP-43: the relationship between protein aggregation and neurodegeneration in amyotrophic lateral sclerosis and frontotemporal lobar degeneration. *The FEBS journal* **278**, 3539–49 (2011).
12. Zhang, Y.-J. *et al.* Progranulin mediates caspase-dependent cleavage of TAR DNA binding protein-43. *The Journal of neuroscience : the official journal of the Society for Neuroscience* **27**, 10530–4 (2007).
13. Chiò, A., Benzi, G., Dossena, M., Mutani, R. & Mora, G. Severely increased risk of amyotrophic lateral sclerosis among Italian professional football players. *Brain : a journal of neurology* **128**, 472–6 (2005).
14. Lu, T. *et al.* Gene regulation and DNA damage in the ageing human brain. *Nature* **429**, 883–91 (2004).

-
15. Conboy, I. M. & Rando, T. A. Aging, Stem Cells and Tissue Regeneration: Lessons from Muscle. *Cell Cycle* **4**, 407–410 (2005).
 16. Gama Sosa, M. a, De Gasperi, R. & Elder, G. a Modeling human neurodegenerative diseases in transgenic systems. *Human genetics* **131**, 535–63 (2012).
 17. Vijg, J. & Campisi, J. Puzzles, promises and a cure for ageing. *Nature* **454**, 1065–71 (2008).
 18. Park, I.-H. *et al.* Reprogramming of human somatic cells to pluripotency with defined factors. *Nature* **451**, 141–6 (2008).
 19. Dolmetsch, R. & Geschwind, D. H. The human brain in a dish: the promise of iPSC-derived neurons. *Cell* **145**, 831–4 (2011).
 20. Cundiff, P. E. & Anderson, S. A. Impact of induced pluripotent stem cells on the study of central nervous system disease. *Current opinion in genetics & development* **21**, 354–61 (2011).
 21. Ming, G.-L. *et al.* Cellular reprogramming: recent advances in modeling neurological diseases. *The Journal of neuroscience : the official journal of the Society for Neuroscience* **31**, 16070–5 (2011).
 22. H., P. P. and R. & Brown Molecular biology of amyotrophic lateral sclerosis: insights from genetics. at http://www.researchchals.org/uploaded_files/pasinelli_nns_rev_0906_844ub6.pdf
 23. Brown, R. H. Amyotrophic lateral sclerosis. Insights from genetics. *Archives of neurology* **54**, 1246–50 (1997).
 24. Patani, R., Sibley, C. R., Chandran, S. & Ule, J. Using human pluripotent stem cells to study post-transcriptional mechanisms of neurodegenerative diseases. *Brain research* **1462**, 129–38 (2012).
 25. Lemon, R. N. & Griffiths, J. Comparing the function of the corticospinal system in different species: organizational differences for motor specialization? *Muscle & nerve* **32**, 261–79 (2005).
 26. Dugdale DC, H. D. *A.D.A.M. Medical Encyclopedia. Amyotrophic lateral sclerosis* (2010).at <http://www.ncbi.nlm.nih.gov/pubmedhealth/PMH0001708/>
 27. Liachko, N. F., Guthrie, C. R. & Kraemer, B. C. Phosphorylation promotes neurotoxicity in a *Caenorhabditis elegans* model of TDP-43 proteinopathy. *The Journal of neuroscience : the official journal of the Society for Neuroscience* **30**, 16208–19 (2010).
 28. Lewis P. Rowland, M.D., and Neil A. Shneider, M.D., P. D. Amyotrophic Lateral Sclerosis. at <http://www.nejm.org.proxy.library.uu.nl/doi/full/10.1056/NEJM200105313442207>
 29. Hayashi, H. & Kato, S. Total manifestations of amyotrophic lateral sclerosis. *Journal of the Neurological Sciences* **93**, 19–35 (1989).
 30. Phukan, J., Pender, N. P. & Hardiman, O. Cognitive impairment in amyotrophic lateral sclerosis. *Lancet neurology* **6**, 994–1003 (2007).
-

31. Nishitoh, H. *et al.* ALS-linked mutant SOD1 induces ER stress- and ASK1-dependent motor neuron death by targeting Derlin-1. *Genes & development* **22**, 1451–64 (2008).
32. Camacho, A. & Massieu, L. Role of glutamate transporters in the clearance and release of glutamate during ischemia and its relation to neuronal death. *Archives of medical research* **37**, 11–8 (2006).
33. Rossi, L., Valle, C. & Carrì, M. T. Altered gene expression, mitochondrial damage and oxidative stress: converging routes in motor neuron degeneration. *International journal of cell biology* **2012**, 908724 (2012).
34. Acevedo-Arozena, A. *et al.* A comprehensive assessment of the SOD1G93A low-copy transgenic mouse, which models human amyotrophic lateral sclerosis. *Disease models & mechanisms* **4**, 686–700
35. Cronin, S., Hardiman, O. & Traynor, B. J. Ethnic variation in the incidence of ALS: a systematic review. *Neurology* **68**, 1002–7 (2007).
36. Wijesekera, L. C. & Leigh, P. N. Amyotrophic lateral sclerosis. *Orphanet journal of rare diseases* **4**, 3 (2009).
37. Cox, P. A. & Sacks, O. W. Cycad neurotoxins, consumption of flying foxes, and ALS-PDC disease in Guam. *Neurology* **58**, 956–9 (2002).
38. Golomb, B. A., Kwon, E. K., Koperski, S. & Evans, M. A. Amyotrophic lateral sclerosis-like conditions in possible association with cholesterol-lowering drugs: an analysis of patient reports to the University of California, San Diego (UCSD) Statin Effects Study. *Drug safety : an international journal of medical toxicology and drug experience* **32**, 649–61 (2009).
39. Harwood, C. A., McDermott, C. J. & Shaw, P. J. Physical activity as an exogenous risk factor in motor neuron disease (MND): a review of the evidence. *Amyotrophic lateral sclerosis : official publication of the World Federation of Neurology Research Group on Motor Neuron Diseases* **10**, 191–204 (2009).
40. Wicks, P. *et al.* Three soccer playing friends with simultaneous amyotrophic lateral sclerosis. *Amyotrophic lateral sclerosis : official publication of the World Federation of Neurology Research Group on Motor Neuron Diseases* **8**, 177–9 (2007).
41. Kasarskis, E. J. *et al.* Clinical aspects of ALS in Gulf War veterans. *Amyotrophic lateral sclerosis : official publication of the World Federation of Neurology Research Group on Motor Neuron Diseases* **10**, 35–41 (2009).
42. Vanacore, N., Cocco, P., Fadda, D. & Dosemeci, M. Job strain, hypoxia and risk of amyotrophic lateral sclerosis: Results from a death certificate study. *Amyotrophic lateral sclerosis : official publication of the World Federation of Neurology Research Group on Motor Neuron Diseases* **11**, 430–4 (2010).
43. Chen, H., Richard, M., Sandler, D. P., Umbach, D. M. & Kamel, F. Head injury and amyotrophic lateral sclerosis. *American journal of epidemiology* **166**, 810–6 (2007).
44. McKee, A. C. *et al.* TDP-43 proteinopathy and motor neuron disease in chronic traumatic encephalopathy. *Journal of neuropathology and experimental neurology* **69**, 918–29 (2010).

-
45. Kamel, F. *et al.* Lead exposure as a risk factor for amyotrophic lateral sclerosis. *Neuro-degenerative diseases* **2**, 195–201 (2005).
 46. Schmidt, S., Kwee, L. C., Allen, K. D. & Oddone, E. Z. Association of ALS with head injury, cigarette smoking and APOE genotypes. *Journal of the neurological sciences* **291**, 22–9 (2010).
 47. Brooks, B. R. Risk factors in the early diagnosis of ALS: North American epidemiological studies. ALS CARE Study Group. *Amyotrophic lateral sclerosis and other motor neuron disorders : official publication of the World Federation of Neurology, Research Group on Motor Neuron Diseases* **1 Suppl 1**, S19–26 (2000).
 48. ROWLAND L.P., S. H. N. . Amyotrophic Lateral Sclerosis. **344**, 1688–1700 (2001).
 49. MacGowan, D. J. L., Scelsa, S. N. & Waldron, M. An ALS-like syndrome with new HIV infection and complete response to antiretroviral therapy. *Neurology* **57**, 1094–1097 (2001).
 50. Münch, C., O'Brien, J. & Bertolotti, A. Prion-like propagation of mutant superoxide dismutase-1 misfolding in neuronal cells. *Proceedings of the National Academy of Sciences of the United States of America* **108**, 3548–53 (2011).
 51. Ramesh, T. *et al.* A genetic model of amyotrophic lateral sclerosis in zebrafish displays phenotypic hallmarks of motoneuron disease. *Disease models & mechanisms* **3**, 652–62
 52. Graffmo, K. S. *et al.* zdfhxx. *Human molecular genetics* (2012).doi:10.1093/hmg/dds399
 53. Barber, S. C., Mead, R. J. & Shaw, P. J. Oxidative stress in ALS: a mechanism of neurodegeneration and a therapeutic target. *Biochimica et biophysica acta* **1762**, 1051–67
 54. Nishimura, A. L. *et al.* A mutation in the vesicle-trafficking protein VAPB causes late-onset spinal muscular atrophy and amyotrophic lateral sclerosis. *American journal of human genetics* **75**, 822–31 (2004).
 55. Deng, H.-X. *et al.* Mutations in UBQLN2 cause dominant X-linked juvenile and adult-onset ALS and ALS/dementia. *Nature* **477**, 211–5 (2011).
 56. Paul, P. & De Bellerche, J. The role of D-amino acids in amyotrophic lateral sclerosis pathogenesis: a review. *Amino acids* **43**, 1823–31 (2012).
 57. Van Damme, P., Dewil, M., Robberecht, W. & Van Den Bosch, L. Excitotoxicity and amyotrophic lateral sclerosis. *Neuro-degenerative diseases* **2**, 147–59 (2005).
 58. Van Hoecke, A. *et al.* EPHA4 is a disease modifier of amyotrophic lateral sclerosis in animal models and in humans. *Nature medicine* **18**, 1418–22 (2012).
 59. Ferraiuolo, L., Kirby, J., Grierson, A. J., Sendtner, M. & Shaw, P. J. Molecular pathways of motor neuron injury in amyotrophic lateral sclerosis. *Nature Reviews Neurology* **7**, 616–630 (2011).
 60. Bosco, D. A. *et al.* Wild-type and mutant SOD1 share an aberrant conformation and a common pathogenic pathway in ALS. *Nature neuroscience* **13**, 1396–403 (2010).
 61. Lagier-Tourenne, C. & Cleveland, D. W. Rethinking ALS: the FUS about TDP-43. *Cell* **136**, 1001–4 (2009).
-

-
62. Gozes, I. Parkinson's and Alzheimer's diseases: protein aggregations and neuroprotection. *Journal of molecular neuroscience : MN* **24**, 333–6 (2004).
63. Selkoe, D. J. Folding proteins in fatal ways. *Nature* **426**, 900–4 (2003).
64. Keohane, C. The human prion diseases. A review with special emphasis on new variant CJD and comments on surveillance. *Clinical and experimental pathology* **47**, 125–32 (1999).
65. Hebert, D. N. & Molinari, M. In and out of the ER: protein folding, quality control, degradation, and related human diseases. *Physiological reviews* **87**, 1377–408 (2007).
66. Moisse, K. *et al.* Divergent patterns of cytosolic TDP-43 and neuronal progranulin expression following axotomy: implications for TDP-43 in the physiological response to neuronal injury. *Brain research* **1249**, 202–11 (2009).
67. Balch, W. E., Morimoto, R. I., Dillin, A. & Kelly, J. W. Adapting proteostasis for disease intervention. *Science (New York, N.Y.)* **319**, 916–9 (2008).
68. Aguilaniu, H., Gustafsson, L., Rigoulet, M. & Nyström, T. Asymmetric inheritance of oxidatively damaged proteins during cytokinesis. *Science (New York, N.Y.)* **299**, 1751–3 (2003).
69. ScienceDirect.com - Ageing Research Reviews - Protein homeostasis and aging: The importance of exquisite quality control. at
<<http://www.sciencedirect.com.proxy.library.uu.nl/science/article/pii/S156816371000005X>>
70. Gidalevitz, T., Ben-Zvi, A., Ho, K. H., Brignull, H. R. & Morimoto, R. I. Progressive disruption of cellular protein folding in models of polyglutamine diseases. *Science (New York, N.Y.)* **311**, 1471–4 (2006).
71. Boillée, S., Vande Velde, C. & Cleveland, D. W. ALS: a disease of motor neurons and their nonneuronal neighbors. *Neuron* **52**, 39–59 (2006).
72. Bruijn, L. I. *et al.* Aggregation and motor neuron toxicity of an ALS-linked SOD1 mutant independent from wild-type SOD1. *Science (New York, N.Y.)* **281**, 1851–4 (1998).
73. Cleveland, D. W. & Liu, J. Oxidation versus aggregation - how do SOD1 mutants cause ALS? *Nature medicine* **6**, 1320–1 (2000).
74. Ripps, M. E., Huntley, G. W., Hof, P. R., Morrison, J. H. & Gordon, J. W. Transgenic mice expressing an altered murine superoxide dismutase gene provide an animal model of amyotrophic lateral sclerosis. *Proceedings of the National Academy of Sciences of the United States of America* **92**, 689–93 (1995).
75. Brown, R. H. SOD1 aggregates in ALS: cause, correlate or consequence? *Nature medicine* **4**, 1362–4 (1998).
76. Caughey, B. & Lansbury, P. T. Protofibrils, pores, fibrils, and neurodegeneration: separating the responsible protein aggregates from the innocent bystanders. *Annual review of neuroscience* **26**, 267–98 (2003).
-

77. Haass, C. & Selkoe, D. J. Soluble protein oligomers in neurodegeneration: lessons from the Alzheimer's amyloid beta-peptide. *Nature reviews. Molecular cell biology* **8**, 101–12 (2007).
78. Behrends, C. *et al.* Chaperonin TRiC promotes the assembly of polyQ expansion proteins into nontoxic oligomers. *Molecular cell* **23**, 887–97 (2006).
79. Shorter, J. & Lindquist, S. Hsp104 catalyzes formation and elimination of self-replicating Sup35 prion conformers. *Science (New York, N.Y.)* **304**, 1793–7 (2004).
80. Kryndushkin, D. & Shewmaker, F. Modeling ALS and FTL D proteinopathies in yeast: an efficient approach for studying protein aggregation and toxicity. *Prion* **5**, 250–7
81. Morley, J. F. & Morimoto, R. I. Regulation of longevity in *Caenorhabditis elegans* by heat shock factor and molecular chaperones. *Molecular biology of the cell* **15**, 657–64 (2004).
82. Cohen, E., Bieschke, J., Perciavalle, R. M., Kelly, J. W. & Dillin, A. Opposing activities protect against age-onset proteotoxicity. *Science (New York, N.Y.)* **313**, 1604–10 (2006).
83. Alonso, A., Zaidi, T., Novak, M., Grundke-Iqbal, I. & Iqbal, K. Hyperphosphorylation induces self-assembly of tau into tangles of paired helical filaments/straight filaments. *Proceedings of the National Academy of Sciences of the United States of America* **98**, 6923–8 (2001).
84. Tauopathies: A Distinct Class of Neurodegenerative Diseases. **10**, 3–14
85. Taes, I. *et al.* Tau levels do not influence human ALS or motor neuron degeneration in the SOD1G93A mouse. *Neurology* **74**, 1687–93 (2010).
86. Jaiswal, M. K. *et al.* Impairment of mitochondrial calcium handling in a mtSOD1 cell culture model of motoneuron disease. *BMC neuroscience* **10**, 64 (2009).
87. Coyle, J. T. & Puttfarcken, P. Oxidative stress, glutamate, and neurodegenerative disorders. *Science (New York, N.Y.)* **262**, 689–95 (1993).
88. Pervaiz, S. Redox pioneer: professor Barry Halliwell. *Antioxidants & redox signaling* **14**, 1761–6 (2011).
89. Cadenas, E., Boveris, A., Ragan, C. I. & Stoppani, A. O. Production of superoxide radicals and hydrogen peroxide by NADH-ubiquinone reductase and ubiquinol-cytochrome c reductase from beef-heart mitochondria. *Archives of biochemistry and biophysics* **180**, 248–57 (1977).
90. Lenzken, S. C. *et al.* Mutant SOD1 and mitochondrial damage alter expression and splicing of genes controlling neurogenesis in models of neurodegeneration. *Human mutation* **32**, 168–82 (2011).
91. Pokrishevsky, E. *et al.* Aberrant localization of FUS and TDP43 is associated with misfolding of SOD1 in amyotrophic lateral sclerosis. *PLoS one* **7**, e35050 (2012).
92. Mecocci, P. *et al.* Oxidative damage to mitochondrial DNA shows marked age-dependent increases in human brain. *Annals of neurology* **34**, 609–16 (1993).
93. Pryor, W. A. & Squadrito, G. L. The chemistry of peroxynitrite: a product from the reaction of nitric oxide with superoxide. *The American journal of physiology* **268**, L699–722 (1995).

94. Beckman, J. S. & Koppenol, W. H. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. *The American journal of physiology* **271**, C1424–37 (1996).
95. Lee, M. K., Marszalek, J. R. & Cleveland, D. W. A mutant neurofilament subunit causes massive, selective motor neuron death: implications for the pathogenesis of human motor neuron disease. *Neuron* **13**, 975–88 (1994).
96. Nguyen, M. D., Larivière, R. C. & Julien, J. P. Deregulation of Cdk5 in a mouse model of ALS: toxicity alleviated by perikaryal neurofilament inclusions. *Neuron* **30**, 135–47 (2001).
97. Williamson, T. L. *et al.* Absence of neurofilaments reduces the selective vulnerability of motor neurons and slows disease caused by a familial amyotrophic lateral sclerosis-linked superoxide dismutase 1 mutant. *Proceedings of the National Academy of Sciences of the United States of America* **95**, 9631–6 (1998).
98. Ackerley, S. *et al.* Neurofilament heavy chain side arm phosphorylation regulates axonal transport of neurofilaments. *The Journal of cell biology* **161**, 489–95 (2003).
99. Zhao, C. *et al.* Charcot-Marie-Tooth disease type 2A caused by mutation in a microtubule motor KIF1Bbeta. *Cell* **105**, 587–97 (2001).
100. Reid, E. *et al.* A kinesin heavy chain (KIF5A) mutation in hereditary spastic paraplegia (SPG10). *American journal of human genetics* **71**, 1189–94 (2002).
101. Yamada, K. *et al.* Heterozygous mutations of the kinesin KIF21A in congenital fibrosis of the extraocular muscles type 1 (CFEOM1). *Nature genetics* **35**, 318–21 (2003).
102. Verhoeven, K. *et al.* Mutations in the small GTP-ase late endosomal protein RAB7 cause Charcot-Marie-Tooth type 2B neuropathy. *American journal of human genetics* **72**, 722–7 (2003).
103. Schmitt-John, T. *et al.* Mutation of Vps54 causes motor neuron disease and defective spermiogenesis in the wobbler mouse. *Nature genetics* **37**, 1213–5 (2005).
104. Sofroniew, M. V & Vinters, H. V Astrocytes: biology and pathology. *Acta neuropathologica* **119**, 7–35 (2010).
105. ScienceDirect.com - Neurobiology of Aging - Neuron–glia interactions underlie ALS-like axonal cytoskeletal pathology. at
<<http://www.sciencedirect.com.proxy.library.uu.nl/science/article/pii/S0197458009001195>>
106. Clement, A. M. *et al.* Wild-type nonneuronal cells extend survival of SOD1 mutant motor neurons in ALS mice. *Science (New York, N.Y.)* **302**, 113–7 (2003).
107. Haidet-Phillips, A. M. *et al.* Astrocytes from familial and sporadic ALS patients are toxic to motor neurons. *Nature biotechnology* **29**, 824–8 (2011).
108. Hald, A. & Lotharius, J. Oxidative stress and inflammation in Parkinson’s disease: is there a causal link? *Experimental neurology* **193**, 279–90 (2005).

109. Lewis, C.-A., Manning, J., Rossi, F. & Krieger, C. The Neuroinflammatory Response in ALS: The Roles of Microglia and T Cells. *Neurology research international* **2012**, 803701 (2012).
110. Zhang, R. *et al.* Evidence for systemic immune system alterations in sporadic amyotrophic lateral sclerosis (sALS). *Journal of neuroimmunology* **159**, 215–24 (2005).
111. ScienceDirect.com - The Lancet Neurology - Neuroinflammation in amyotrophic lateral sclerosis: role of glial activation in motor neuron disease. at <<http://www.sciencedirect.com.proxy.library.uu.nl/science/article/pii/S1474442211700151>>
112. Van Damme, P. *et al.* Astrocytes regulate GluR2 expression in motor neurons and their vulnerability to excitotoxicity. *Proceedings of the National Academy of Sciences of the United States of America* **104**, 14825–30 (2007).
113. Chi, L. *et al.* Motor neuron degeneration promotes neural progenitor cell proliferation, migration, and neurogenesis in the spinal cords of amyotrophic lateral sclerosis mice. *Stem cells (Dayton, Ohio)* **24**, 34–43 (2006).
114. Svitlana Garbuzova-Davisa, b, c, d, Corresponding author contact information, E. the corresponding author *et al.* Amyotrophic lateral sclerosis: A neurovascular disease. at <<http://www.sciencedirect.com.proxy.library.uu.nl/science/article/pii/S0006899311008705>>
115. Svitlana Garbuzova-Davisa, b, c, d, Corresponding author contact information, E. the corresponding author *et al.* Disruption of neurovascular unit prior to motor neuron degeneration in amyotrophic lateral sclerosis. *Journal of neuroscience research* **89**, 718–28 (2011).
116. Garbuzova-Davis, S. *et al.* Ultrastructure of blood-brain barrier and blood-spinal cord barrier in SOD1 mice modeling ALS. *Brain research* **1157**, 126–37 (2007).
117. Nicaise, C. *et al.* Impaired blood-brain and blood-spinal cord barriers in mutant SOD1-linked ALS rat. *Brain research* **1301**, 152–62 (2009).
118. Tanaka, F., Ikenaka, K., Yamamoto, M. & Sobue, G. Neuropathology and omics in motor neuron diseases. *Neuropathology: official journal of the Japanese Society of Neuropathology* **32**, 458–62 (2012).
119. Vargas, M. R., Pehar, M., Díaz-Amarilla, P. J., Beckman, J. S. & Barbeito, L. Transcriptional profile of primary astrocytes expressing ALS-linked mutant SOD1. *Journal of neuroscience research* **86**, 3515–25 (2008).
120. Bäumer, D., Ansorge, O., Almeida, M. & Talbot, K. The role of RNA processing in the pathogenesis of motor neuron degeneration. *Expert reviews in molecular medicine* **12**, e21 (2010).
121. DeJesus-Hernandez, M. *et al.* Expanded GGGGCC hexanucleotide repeat in noncoding region of C9ORF72 causes chromosome 9p-linked FTD and ALS. *Neuron* **72**, 245–56 (2011).
122. Renton, A. E. *et al.* A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. *Neuron* **72**, 257–68 (2011).

123. Maracchioni, A. *et al.* Mitochondrial damage modulates alternative splicing in neuronal cells: implications for neurodegeneration. *Journal of neurochemistry* **100**, 142–53 (2007).
124. Yim, M. B. *et al.* A gain-of-function of an amyotrophic lateral sclerosis-associated Cu,Zn-superoxide dismutase mutant: An enhancement of free radical formation due to a decrease in Km for hydrogen peroxide. *Proceedings of the National Academy of Sciences of the United States of America* **93**, 5709–14 (1996).
125. Elia, A. J. *et al.* Expression of human FALS SOD in motoneurons of *Drosophila*. *Free radical biology & medicine* **26**, 1332–8 (1999).
126. Lee, E. B., Lee, V. M.-Y. & Trojanowski, J. Q. Gains or losses: molecular mechanisms of TDP43-mediated neurodegeneration. *Nature reviews. Neuroscience* **13**, 38–50 (2012).
127. Rouiller, E. M., Moret, V., Tanne, J. & Boussaoud, D. Evidence for direct connections between the hand region of the supplementary motor area and cervical motoneurons in the macaque monkey. *The European journal of neuroscience* **8**, 1055–9 (1996).
128. Zhang, S.-C. Neural subtype specification from embryonic stem cells. *Brain pathology (Zurich, Switzerland)* **16**, 132–42 (2006).
129. Eisen, A. Amyotrophic lateral sclerosis-Evolutionary and other perspectives. *Muscle & nerve* **40**, 297–304 (2009).
130. Dermitzakis, E. T. & Clark, A. G. Evolution of transcription factor binding sites in Mammalian gene regulatory regions: conservation and turnover. *Molecular biology and evolution* **19**, 1114–21 (2002).
131. Liu, G.-H., Ding, Z. & Izpisua Belmonte, J. C. iPSC technology to study human aging and aging-related disorders. *Current opinion in cell biology* **null**, (2012).
132. Grskovic, M., Javaherian, A., Strulovici, B. & Daley, G. Q. Induced pluripotent stem cells—opportunities for disease modelling and drug discovery. *Nature reviews. Drug discovery* **10**, 915–29 (2011).
133. Bastos, A. F. *et al.* Amyotrophic lateral sclerosis: one or multiple causes? *Neurology international* **3**, e4 (2011).
134. Morrison, S. J., Wandycz, A. M., Akashi, K., Globerson, A. & Weissman, I. L. The aging of hematopoietic stem cells. *Nature Medicine* **2**, 1011–1016 (1996).
135. Hasty, P., Campisi, J., Hoeijmakers, J., Van Steeg, H. & Vijg, J. Aging and genome maintenance: lessons from the mouse? *Science (New York, N.Y.)* **299**, 1355–9 (2003).
136. Bishop, N. A. & Guarente, L. Genetic links between diet and lifespan: shared mechanisms from yeast to humans. *Nature reviews. Genetics* **8**, 835–44 (2007).
137. Conboy, I. M. *et al.* Rejuvenation of aged progenitor cells by exposure to a young systemic environment. *Nature* **433**, 760–4 (2005).
138. Carlson, M. E. & Conboy, I. M. Loss of stem cell regenerative capacity within aged niches. *Aging Cell* **6**, 371–382 (2007).

139. Mair, W. & Dillin, A. Aging and survival: the genetics of life span extension by dietary restriction. *Annual review of biochemistry* **77**, 727–54 (2008).
140. Kenyon, C., Chang, J., Gensch, E., Rudner, A. & Tabtiang, R. A *C. elegans* mutant that lives twice as long as wild type. *Nature* **366**, 461–4 (1993).
141. Giannakou, M. E. & Partridge, L. Role of insulin-like signalling in *Drosophila* lifespan. *Trends in biochemical sciences* **32**, 180–8 (2007).
142. Tatar, M. *et al.* A mutant *Drosophila* insulin receptor homolog that extends life-span and impairs neuroendocrine function. *Science (New York, N.Y.)* **292**, 107–10 (2001).
143. Holzenberger, M. *et al.* IGF-1 receptor regulates lifespan and resistance to oxidative stress in mice. *Nature* **421**, 182–7 (2003).
144. Taguchi, A., Wartschow, L. M. & White, M. F. Brain IRS2 signaling coordinates life span and nutrient homeostasis. *Science (New York, N.Y.)* **317**, 369–72 (2007).
145. Wood, J. G. *et al.* Sirtuin activators mimic caloric restriction and delay ageing in metazoans. *Nature* **430**, 686–9 (2004).
146. Kimura, K. D. *daf-2*, an Insulin Receptor-Like Gene That Regulates Longevity and Diapause in *Caenorhabditis elegans*. *Science* **277**, 942–946 (1997).
147. Kimura, K. D., Tissenbaum, H. A., Liu, Y. & Ruvkun, G. *daf-2*, an insulin receptor-like gene that regulates longevity and diapause in *Caenorhabditis elegans*. *Science (New York, N.Y.)* **277**, 942–6 (1997).
148. Morris, J. Z., Tissenbaum, H. A. & Ruvkun, G. A phosphatidylinositol-3-OH kinase family member regulating longevity and diapause in *Caenorhabditis elegans*. *Nature* **382**, 536–9 (1996).
149. Ogg, S. & Ruvkun, G. The *C. elegans* PTEN homolog, DAF-18, acts in the insulin receptor-like metabolic signaling pathway. *Molecular cell* **2**, 887–93 (1998).
150. Henderson, S. T. & Johnson, T. E. *daf-16* integrates developmental and environmental inputs to mediate aging in the nematode *Caenorhabditis elegans*. *Current biology : CB* **11**, 1975–80 (2001).
151. Kopito, R. R. & Ron, D. Conformational disease. *Nature cell biology* **2**, E207–9 (2000).
152. Lithgow, G. J., White, T. M., Melov, S. & Johnson, T. E. Thermotolerance and extended life-span conferred by single-gene mutations and induced by thermal stress. *Proceedings of the National Academy of Sciences of the United States of America* **92**, 7540–4 (1995).
153. Morley, J. F., Brignull, H. R., Weyers, J. J. & Morimoto, R. I. The threshold for polyglutamine-expansion protein aggregation and cellular toxicity is dynamic and influenced by aging in *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences of the United States of America* **99**, 10417–22 (2002).
154. Hsu, A.-L., Murphy, C. T. & Kenyon, C. Regulation of aging and age-related disease by DAF-16 and heat-shock factor. *Science (New York, N.Y.)* **300**, 1142–5 (2003).

155. Walker, G. A., Thompson, F. J., Brawley, A., Scanlon, T. & Devaney, E. Heat shock factor functions at the convergence of the stress response and developmental pathways in *Caenorhabditis elegans*. *FASEB journal: official publication of the Federation of American Societies for Experimental Biology* **17**, 1960–2 (2003).
156. Sirt, D. *et al.* Stress-Inducible Regulation of Heat Shock Factor 1 by the Deacetylase SIRT1. **323**, 1063–1066 (2012).
157. Ross, C. A. Polyglutamine pathogenesis: emergence of unifying mechanisms for Huntington’s disease and related disorders. *Neuron* **35**, 819–22 (2002).
158. Meléndez, A. *et al.* Autophagy genes are essential for dauer development and life-span extension in *C. elegans*. *Science (New York, N.Y.)* **301**, 1387–91 (2003).
159. Hara, T. *et al.* Suppression of basal autophagy in neural cells causes neurodegenerative disease in mice. *Nature* **441**, 885–9 (2006).
160. Dillin, A. *et al.* Rates of behavior and aging specified by mitochondrial function during development. *Science (New York, N.Y.)* **298**, 2398–401 (2002).
161. Blüher, M., Kahn, B. B. & Kahn, C. R. Extended longevity in mice lacking the insulin receptor in adipose tissue. *Science (New York, N.Y.)* **299**, 572–4 (2003).
162. Kurtzke, J. F. & Beebe, G. W. Epidemiology of amyotrophic lateral sclerosis: 1. A case-control comparison based on ALS deaths. *Neurology* **30**, 453–62 (1980).
163. Longstreth, W. T., McGuire, V., Koepsell, T. D., Wang, Y. & Van Belle, G. Risk of amyotrophic lateral sclerosis and history of physical activity: a population-based case-control study. *Archives of neurology* **55**, 201–6 (1998).
164. Beretta, S., Carri, M. T., Beghi, E., Chiò, A. & Ferrarese, C. The sinister side of Italian soccer. *The Lancet Neurology* **2**, 656–657 (2003).
165. Armon, C. & Nelson, L. M. Is head trauma a risk factor for amyotrophic lateral sclerosis? An evidence based review. *Amyotrophic lateral sclerosis: official publication of the World Federation of Neurology Research Group on Motor Neuron Diseases* **13**, 351–6 (2012).
166. Snyder, E. M. *et al.* SCRAM: A scoring and ranking system for persistent, bioaccumulative, and toxic substances for the North American Great Lakes. Part II: Bioaccumulation potential and persistence. *Environmental science and pollution research international* **7**, 116–21 (2000).
167. Weisskopf, M. G. *et al.* Prospective study of chemical exposures and amyotrophic lateral sclerosis. *Journal of neurology, neurosurgery, and psychiatry* **80**, 558–61 (2009).
168. Mitchell, J. D. Amyotrophic lateral sclerosis: toxins and environment. *Amyotrophic lateral sclerosis and other motor neuron disorders: official publication of the World Federation of Neurology, Research Group on Motor Neuron Diseases* **1**, 235–50 (2000).
169. Veldink, J. H. *et al.* Sexual differences in onset of disease and response to exercise in a transgenic model of ALS. *Neuromuscular disorders: NMD* **13**, 737–43 (2003).

-
170. Mahoney, D. J., Rodriguez, C., Devries, M., Yasuda, N. & Tarnopolsky, M. A. Effects of high-intensity endurance exercise training in the G93A mouse model of amyotrophic lateral sclerosis. *Muscle & nerve* **29**, 656–62 (2004).
171. Trojsi, F. *et al.* Clinical features and lifestyle of patients with amyotrophic lateral sclerosis in Campania: brief overview of an Italian database. *Annali dell'Istituto superiore di sanità* **48**, 287–91 (2012).
172. McArdle, A., Pattwell, D., Vasilaki, A., Griffiths, R. D. & Jackson, M. J. Contractile activity-induced oxidative stress: cellular origin and adaptive responses. *American journal of physiology. Cell physiology* **280**, C621–7 (2001).
173. Kadenbach, B., Ramzan, R. & Vogt, S. High Efficiency versus Maximal Performance - The Cause of Oxidative Stress in Eukaryotes: A Hypothesis. *Mitochondrion* (2012).doi:10.1016/j.mito.2012.11.005
174. Lee, S. S. *et al.* A systematic RNAi screen identifies a critical role for mitochondria in *C. elegans* longevity. *Nature genetics* **33**, 40–8 (2003).
175. Rea, S. L., Ventura, N. & Johnson, T. E. Relationship between mitochondrial electron transport chain dysfunction, development, and life extension in *Caenorhabditis elegans*. *PLoS biology* **5**, e259 (2007).
176. Zahn, J. M. *et al.* AGEMAP: a gene expression database for aging in mice. *PLoS genetics* **3**, e201 (2007).
177. Loerch, P. M. *et al.* Evolution of the aging brain transcriptome and synaptic regulation. *PloS one* **3**, e3329 (2008).
178. Trifunovic, A. *et al.* Premature ageing in mice expressing defective mitochondrial DNA polymerase. *Nature* **429**, 417–23 (2004).
179. Cristina, D., Cary, M., Lunceford, A., Clarke, C. & Kenyon, C. A regulated response to impaired respiration slows behavioral rates and increases lifespan in *Caenorhabditis elegans*. *PLoS genetics* **5**, e1000450 (2009).
180. Goldberg, A. D., Allis, C. D. & Bernstein, E. Epigenetics: a landscape takes shape. *Cell* **128**, 635–8 (2007).
181. Sebastian, C., Satterstrom, K. F., Haigis, M. C. & Mostoslavsky, R. From Sirtuin biology to human diseases: an update. *The Journal of biological chemistry* (2012).doi:10.1074/jbc.R112.402768
182. Tullet, J. M. A. *et al.* Direct inhibition of the longevity-promoting factor SKN-1 by insulin-like signaling in *C. elegans*. *Cell* **132**, 1025–38 (2008).
183. Neumann, M. *et al.* Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science (New York, N.Y.)* **314**, 130–3 (2006).
184. Elden, A. C. *et al.* Ataxin-2 intermediate-length polyglutamine expansions are associated with increased risk for ALS. *Nature* **466**, 1069–75 (2010).
185. Lee, T. *et al.* Ataxin-2 intermediate-length polyglutamine expansions in European ALS patients. *Human molecular genetics* **20**, 1697–700 (2011).
-

-
186. Bonini, N. M. & Gitler, A. D. Model organisms reveal insight into human neurodegenerative disease: ataxin-2 intermediate-length polyglutamine expansions are a risk factor for ALS. *Journal of molecular neuroscience : MN* **45**, 676–83 (2011).
187. Van Langenhove, T. *et al.* Ataxin-2 polyQ expansions in FTLD-ALS spectrum disorders in Flanders-Belgian cohorts. *Neurobiology of aging* **33**, 1004.e17–20 (2012).
188. Imbert, G. *et al.* Cloning of the gene for spinocerebellar ataxia 2 reveals a locus with high sensitivity to expanded CAG/glutamine repeats. *Nature genetics* **14**, 285–91 (1996).
189. Sanpei, K. *et al.* Identification of the spinocerebellar ataxia type 2 gene using a direct identification of repeat expansion and cloning technique, DIRECT. *Nature genetics* **14**, 277–84 (1996).
190. Nihei, Y., Ito, D. & Suzuki, N. Roles of ataxin-2 in pathological cascades mediated by TAR DNA-binding Protein 43 (TDP-43) and Fused in Sarcoma (FUS). *The Journal of biological chemistry* **43**, (2012).
191. Hart, M. P. & Gitler, A. D. ALS-associated ataxin 2 polyQ expansions enhance stress-induced caspase 3 activation and increase TDP-43 pathological modifications. *The Journal of neuroscience : the official journal of the Society for Neuroscience* **32**, 9133–42 (2012).
192. Evert, B. O., Wüllner, U. & Klockgether, T. Cell death in polyglutamine diseases. *Cell and tissue research* **301**, 189–204 (2000).
193. Kwon, S., Zhang, Y. & Matthias, P. The deacetylase HDAC6 is a novel critical component of stress granules involved in the stress response. *Genes & development* **21**, 3381–94 (2007).
194. Khong, A. & Jan, E. Modulation of stress granules and P bodies during dicistrovirus infection. *Journal of virology* **85**, 1439–51 (2011).
195. White, J. P., Cardenas, A. M., Marissen, W. E. & Lloyd, R. E. Inhibition of cytoplasmic mRNA stress granule formation by a viral proteinase. *Cell host & microbe* **2**, 295–305 (2007).
196. Dormann, D. *et al.* ALS-associated fused in sarcoma (FUS) mutations disrupt Transportin-mediated nuclear import. *The EMBO journal* **29**, 2841–57 (2010).
197. Schieke, S. M. & Finkel, T. Mitochondrial signaling, TOR, and life span. *Biological chemistry* **387**, 1357–61
198. Ilieva, H., Polymenidou, M. & Cleveland, D. W. Non-cell autonomous toxicity in neurodegenerative disorders: ALS and beyond. *The Journal of cell biology* **187**, 761–72 (2009).
199. Johnson, J. D., Otani, K., Bell, G. I. & Polonsky, K. S. Impaired insulin secretion in transgenic mice over-expressing calpastatin in pancreatic β -cells. *Islets* **1**, 242–8
200. Pupillo, E. *et al.* Trauma and amyotrophic lateral sclerosis: a case-control study from a population-based registry. *European journal of neurology : the official journal of the European Federation of Neurological Societies* 1509–1517 (2012).doi:10.1111/j.1468-1331.2012.03723.x
-

201. Beghi, E. *et al.* Amyotrophic lateral sclerosis, physical exercise, trauma and sports: results of a population-based pilot case-control study. *Amyotrophic lateral sclerosis : official publication of the World Federation of Neurology Research Group on Motor Neuron Diseases* **11**, 289–92 (2010).
202. Turner, M. R., Abisgold, J., Yeates, D. G. R., Talbot, K. & Goldacre, M. J. Head and other physical trauma requiring hospitalisation is not a significant risk factor in the development of ALS. *Journal of the neurological sciences* **288**, 45–8 (2010).
203. Bishop, N. A., Lu, T. & Yankner, B. A. Neural mechanisms of ageing and cognitive decline. *Nature* **464**, 529–35 (2010).
204. Li, L. & Xie, T. Stem cell niche: structure and function. *Annual review of cell and developmental biology* **21**, 605–31 (2005).
205. Zacks, S. I. & Sheff, M. F. Age-related impeded regeneration of mouse minced anterior tibial muscle. *Muscle & nerve* **5**, 152–61 (1982).
206. Liu, L. & Rando, T. a Manifestations and mechanisms of stem cell aging. *The Journal of cell biology* **193**, 257–66 (2011).
207. Conboy, I. M., Conboy, M. J., Smythe, G. M. & Rando, T. A. Notch-mediated restoration of regenerative potential to aged muscle. *Science (New York, N.Y.)* **302**, 1575–7 (2003).
208. Delafontaine, P., Song, Y.-H. & Li, Y. Expression, regulation, and function of IGF-1, IGF-1R, and IGF-1 binding proteins in blood vessels. *Arteriosclerosis, thrombosis, and vascular biology* **24**, 435–44 (2004).
209. Carlson, M. E. *et al.* Relative roles of TGF-beta1 and Wnt in the systemic regulation and aging of satellite cell responses. *Aging cell* **8**, 676–89 (2009).
210. Doetsch, F. A niche for adult neural stem cells. *Current opinion in genetics & development* **13**, 543–50 (2003).
211. Lin, H. The stem-cell niche theory: lessons from flies. *Nature reviews. Genetics* **3**, 931–40 (2002).
212. Conboy, I. M., Yousef, H. & Conboy, M. J. Embryonic anti-aging niche. *Aging* **3**, 555–63 (2011).
213. Rando, T. A. Stem cells, ageing and the quest for immortality. *Nature* **441**, 1080–6 (2006).
214. ScienceDirect.com - Developmental Cell - At the Roots of a Never-Ending Cycle. at <<http://www.sciencedirect.com.proxy.library.uu.nl/science/article/pii/S1534580701000223>>
215. Anchelin, M., Murcia, L., Alcaraz-Pérez, F., García-Navarro, E. M. & Cayuela, M. L. Behaviour of telomere and telomerase during aging and regeneration in zebrafish. *PloS one* **6**, e16955 (2011).
216. Lee, H. W. *et al.* Essential role of mouse telomerase in highly proliferative organs. *Nature* **392**, 569–74 (1998).
217. ScienceDirect.com - Ageing Research Reviews - Aging: Central role for autophagy and the lysosomal degradative system. at

- <<http://www.sciencedirect.com.proxy.library.uu.nl/science/article/pii/S1568163709000257>>
218. Wichterle, H. & Przedborski, S. What can pluripotent stem cells teach us about neurodegenerative diseases? *Nature neuroscience* **13**, 800–4 (2010).
219. ScienceDirect.com - Cell - A Bivalent Chromatin Structure Marks Key Developmental Genes in Embryonic Stem Cells. at <<http://www.sciencedirect.com.proxy.library.uu.nl/science/article/pii/S0092867406003801>>
220. Shin, D. M. *et al.* Novel epigenetic mechanisms that control pluripotency and quiescence of adult bone marrow-derived Oct4(+) very small embryonic-like stem cells. *Leukemia : official journal of the Leukemia Society of America, Leukemia Research Fund, U.K* **23**, 2042–51 (2009).
221. Berger, S. L. The complex language of chromatin regulation during transcription. *Nature* **447**, 407–12 (2007).
222. Carlson, M. E. *et al.* Relative roles of TGF-beta1 and Wnt in the systemic regulation and aging of satellite cell responses. *Aging cell* **8**, 676–89 (2009).
223. Kuhn, H. G., Dickinson-Anson, H. & Gage, F. H. Neurogenesis in the dentate gyrus of the adult rat: age-related decrease of neuronal progenitor proliferation. *The Journal of neuroscience : the official journal of the Society for Neuroscience* **16**, 2027–33 (1996).
224. Molofsky, A. V *et al.* Increasing p16INK4a expression decreases forebrain progenitors and neurogenesis during ageing. *Nature* **443**, 448–52 (2006).
225. Janzen, V. *et al.* Stem-cell ageing modified by the cyclin-dependent kinase inhibitor p16INK4a. *Nature* **443**, 421–6 (2006).
226. Sharpless, N. E. & DePinho, R. A. How stem cells age and why this makes us grow old. *Nature reviews. Molecular cell biology* **8**, 703–13 (2007).
227. Kucia, M. *et al.* A population of very small embryonic-like (VSEL) CXCR4(+)SSEA-1(+)Oct-4+ stem cells identified in adult bone marrow. *Leukemia : official journal of the Leukemia Society of America, Leukemia Research Fund, U.K* **20**, 857–69 (2006).
228. Ratajczak, M. Z., Shin, D.-M., Ratajczak, J., Kucia, M. & Bartke, A. A novel insight into aging: are there pluripotent very small embryonic-like stem cells (VSELs) in adult tissues overtime depleted in an Igf-1-dependent manner? *Aging* **2**, 875–83 (2010).
229. Taichman, R. S. *et al.* Prospective identification and skeletal localization of cells capable of multilineage differentiation in vivo. *Stem cells and development* **19**, 1557–70 (2010).
230. Dawn, B. *et al.* Transplantation of bone marrow-derived very small embryonic-like stem cells attenuates left ventricular dysfunction and remodeling after myocardial infarction. *Stem cells (Dayton, Ohio)* **26**, 1646–55 (2008).

-
231. Ludwig, T. *et al.* Mouse mutants lacking the type 2 IGF receptor (IGF2R) are rescued from perinatal lethality in *Igf2* and *Igf1r* null backgrounds. *Developmental biology* **177**, 517–35 (1996).
 232. Ratajczak, M. Z., Zuba-Surma, E. K., Shin, D.-M., Ratajczak, J. & Kucia, M. Very small embryonic-like (VSEL) stem cells in adult organs and their potential role in rejuvenation of tissues and longevity. *Experimental gerontology* **43**, 1009–17 (2008).
 233. Bokov, A. F., Lindsey, M. L., Khodr, C., Sabia, M. R. & Richardson, A. Long-lived ames dwarf mice are resistant to chemical stressors. *The journals of gerontology. Series A, Biological sciences and medical sciences* **64**, 819–27 (2009).
 234. Sharma, N., Castorena, C. M. & Cartee, G. D. Tissue-specific responses of IGF-1/insulin and mTOR signaling in calorie restricted rats. *PLoS one* **7**, e38835 (2012).
 235. Conboy, I. M. *et al.* Rejuvenation of aged progenitor cells by exposure to a young systemic environment. *Nature* **433**, 760–4 (2005).
 236. Ogg, S. *et al.* The Fork head transcription factor DAF-16 transduces insulin-like metabolic and longevity signals in *C. elegans*. *Nature* **389**, 994–9 (1997).
 237. Eiraku, M. *et al.* Self-organized formation of polarized cortical tissues from ESCs and its active manipulation by extrinsic signals. *Cell stem cell* **3**, 519–32 (2008).
 238. Feng, B., Ng, J.-H., Heng, J.-C. D. & Ng, H.-H. Molecules that promote or enhance reprogramming of somatic cells to induced pluripotent stem cells. *Cell stem cell* **4**, 301–12 (2009).
 239. Roccatagliata, L., Bonzano, L., Mancardi, G., Canepa, C. & Caponnetto, C. Detection of motor cortex thinning and corticospinal tract involvement by quantitative MRI in amyotrophic lateral sclerosis. *Amyotrophic lateral sclerosis : official publication of the World Federation of Neurology Research Group on Motor Neuron Diseases* **10**, 47–52 (2009).
 240. Piras, F., Salani, F., Bossù, P., Caltagirone, C. & Spalletta, G. High serum levels of transforming growth factor β 1 are associated with increased cortical thickness in cingulate and right frontal areas in healthy subjects. *Journal of neuroinflammation* **9**, 42 (2012).
 241. Kucia, M. J. *et al.* Evidence that very small embryonic-like stem cells are mobilized into peripheral blood. *Stem cells (Dayton, Ohio)* **26**, 2083–92 (2008).
 242. Evans, M. J. & Kaufman, M. H. Establishment in culture of pluripotential cells from mouse embryos. *Nature* **292**, 154–6 (1981).
 243. Martin, G. R. Isolation of a pluripotent cell line from early mouse embryos cultured in medium conditioned by teratocarcinoma stem cells. *Proceedings of the National Academy of Sciences of the United States of America* **78**, 7634–8 (1981).
 244. Vitale, A. M. *et al.* Variability in the Generation of Induced Pluripotent Stem Cells: Importance for Disease Modeling. *Stem Cells Translational Medicine* **1**, 641–650 (2012).
 245. Björklund, A. & Dunnett, S. B. Dopamine neuron systems in the brain: an update. *Trends in neurosciences* **30**, 194–202 (2007).
-

-
246. Krencik, R., Weick, J. P., Liu, Y., Zhang, Z.-J. & Zhang, S.-C. Specification of transplantable astroglial subtypes from human pluripotent stem cells. *Nature biotechnology* **29**, 528–34 (2011).
247. Han, S. S. W., Williams, L. A. & Eggan, K. C. Constructing and deconstructing stem cell models of neurological disease. *Neuron* **70**, 626–44 (2011).
248. Yamanaka, S. Induced pluripotent stem cells: past, present, and future. *Cell stem cell* **10**, 678–84 (2012).
249. Takahashi, K. & Yamanaka, S. Induction of Pluripotent Stem Cells from Mouse Embryonic and Adult Fibroblast Cultures by Defined Factors. *Cell* **126**, 663–676 (2006).
250. Yamanaka, S. Elite and stochastic models for induced pluripotent stem cell generation. *Nature* **460**, 49–52 (2009).
251. Yamanaka, S. & Blau, H. M. Nuclear reprogramming to a pluripotent state by three approaches. *Nature* **465**, 704–12 (2010).
252. Loh, Y.-H. *et al.* Generation of induced pluripotent stem cells from human blood. *Blood* **113**, 5476–9 (2009).
253. Papapetrou, E. P. *et al.* Stoichiometric and temporal requirements of Oct4, Sox2, Klf4, and c-Myc expression for efficient human iPSC induction and differentiation. *Proceedings of the National Academy of Sciences of the United States of America* **106**, 12759–64 (2009).
254. Hu, B.-Y. *et al.* Neural differentiation of human induced pluripotent stem cells follows developmental principles but with variable potency. *Proceedings of the National Academy of Sciences of the United States of America* **107**, 4335–40 (2010).
255. Boulting, G. L. *et al.* A functionally characterized test set of human induced pluripotent stem cells. *Nature biotechnology* **29**, 279–86 (2011).
256. Patani, R. *et al.* Investigating the utility of human embryonic stem cell-derived neurons to model ageing and neurodegenerative disease using whole-genome gene expression and splicing analysis. *Journal of neurochemistry* **122**, 738–51 (2012).
257. Ghosh, Z. *et al.* Persistent donor cell gene expression among human induced pluripotent stem cells contributes to differences with human embryonic stem cells. *PloS one* **5**, e8975 (2010).
258. Maherali, N. *et al.* Directly reprogrammed fibroblasts show global epigenetic remodeling and widespread tissue contribution. *Cell stem cell* **1**, 55–70 (2007).
259. Dimos, J. T. *et al.* Generated from Patients with ALS Can Be Differentiated into Motor Neurons. 1218–1221 (2008).
260. Dimos, J. T. *et al.* Induced pluripotent stem cells generated from patients with ALS can be differentiated into motor neurons. *Science (New York, N.Y.)* **321**, 1218–21 (2008).
261. Karumbayaram, S. *et al.* Human embryonic stem cell-derived motor neurons expressing SOD1 mutants exhibit typical signs of motor neuron degeneration linked to ALS. *Disease models & mechanisms* **2**, 189–95
-

262. Brunet, A., Datta, S. R. & Greenberg, M. E. Transcription-dependent and -independent control of neuronal survival by the PI3K-Akt signaling pathway. *Current opinion in neurobiology* **11**, 297–305 (2001).
263. Bilican, B. *et al.* Mutant induced pluripotent stem cell lines recapitulate aspects of TDP-43 proteinopathies and reveal cell-specific vulnerability. *Proceedings of the National Academy of Sciences of the United States of America* **109**, 5803–8 (2012).
264. Egawa, N. *et al.* Drug screening for ALS using patient-specific induced pluripotent stem cells. *Science translational medicine* **4**, 145ra104 (2012).
265. Tsuda, H. *et al.* The amyotrophic lateral sclerosis 8 protein VAPB is cleaved, secreted, and acts as a ligand for Eph receptors. *Cell* **133**, 963–77 (2008).
266. Mitne-Neto, M. *et al.* Downregulation of VAPB expression in motor neurons derived from induced pluripotent stem cells of ALS8 patients. *Human molecular genetics* **20**, 3642–52 (2011).
267. Rowland, L. P. Ameliorating amyotrophic lateral sclerosis. *The New England journal of medicine* **362**, 953–4 (2010).
268. Di Giorgio, F. P., Carrasco, M. A., Siao, M. C., Maniatis, T. & Eggan, K. Non-cell autonomous effect of glia on motor neurons in an embryonic stem cell-based ALS model. *Nature neuroscience* **10**, 608–14 (2007).
269. Marchetto, M. C. N. *et al.* Non-cell-autonomous effect of human SOD1 G37R astrocytes on motor neurons derived from human embryonic stem cells. *Cell stem cell* **3**, 649–57 (2008).
270. Egawa, N. *et al.* Drug screening for ALS using patient-specific induced pluripotent stem cells. *Science translational medicine* **4**, 145ra104 (2012).
271. Wang, H. & Doering, L. C. Induced pluripotent stem cells to model and treat neurogenetic disorders. *Neural plasticity* **2012**, 346053 (2012).
272. Hargus, G. *et al.* Differentiated Parkinson patient-derived induced pluripotent stem cells grow in the adult rodent brain and reduce motor asymmetry in Parkinsonian rats. *Proceedings of the National Academy of Sciences of the United States of America* **107**, 15921–6 (2010).
273. Wernig, M. *et al.* Neurons derived from reprogrammed fibroblasts functionally integrate into the fetal brain and improve symptoms of rats with Parkinson’s disease. *Proceedings of the National Academy of Sciences of the United States of America* **105**, 5856–61 (2008).
274. Abeliovich, A. & Doege, C. A. Reprogramming therapeutics: iPS cell prospects for neurodegenerative disease. *Neuron* **61**, 337–9 (2009).
275. Page, R. L. *et al.* Induction of stem cell gene expression in adult human fibroblasts without transgenes. *Cloning and stem cells* **11**, 417–26 (2009).
276. Li, J.-Y. *et al.* Lewy bodies in grafted neurons in subjects with Parkinson’s disease suggest host-to-graft disease propagation. *Nature medicine* **14**, 501–3 (2008).

277. Koch, P. *et al.* Excitation-induced ataxin-3 aggregation in neurons from patients with Machado-Joseph disease. *Nature* **480**, 543–6 (2011).
278. Di Giorgio, F. P., Boulting, G. L., Bobrowicz, S. & Eggan, K. C. Human embryonic stem cell-derived motor neurons are sensitive to the toxic effect of glial cells carrying an ALS-causing mutation. *Cell stem cell* **3**, 637–48 (2008).
279. Sum-Yee Yung, J., Kwong-Hang Tam, P. & Sau-Wai Ngan, E. Pluripotent stem cell for modeling neurological diseases. *Experimental cell research* **null**, (2012).
280. Inoue, H. & Yamanaka, S. The use of induced pluripotent stem cells in drug development. *Clinical pharmacology and therapeutics* **89**, 655–61 (2011).
281. Ash, P. E. A. *et al.* Neurotoxic effects of TDP-43 overexpression in *C. elegans*. *Human molecular genetics* **19**, 3206–18 (2010).
282. Watson, M. R., Lagow, R. D., Xu, K., Zhang, B. & Bonini, N. M. A drosophila model for amyotrophic lateral sclerosis reveals motor neuron damage by human SOD1. *The Journal of biological chemistry* **283**, 24972–81 (2008).
283. Parkes, T. L. *et al.* Extension of *Drosophila* lifespan by overexpression of human SOD1 in motoneurons. *Nature genetics* **19**, 171–4 (1998).
284. Kabashi, E. *et al.* Gain and loss of function of ALS-related mutations of TARDBP (TDP-43) cause motor deficits in vivo. *Human molecular genetics* **19**, 671–83 (2010).
285. Gendron, T. F. & Petrucelli, L. Rodent models of TDP-43 proteinopathy: investigating the mechanisms of TDP-43-mediated neurodegeneration. *Journal of molecular neuroscience : MN* **45**, 486–99 (2011).
286. Shan, X., Chiang, P.-M., Price, D. L. & Wong, P. C. Altered distributions of Gemini of coiled bodies and mitochondria in motor neurons of TDP-43 transgenic mice. *Proceedings of the National Academy of Sciences of the United States of America* **107**, 16325–30 (2010).

 SUPPLEMENTARY FIGURES

Table 1: Pathological changes in ALS (Table adapted from *Ferraiuolo, L. et al. (2011)*)⁵⁹.

Gross pathological changes	Microscopic changes
Atrophy of the precentral gyrus	Depletion of $\geq 50\%$ of the spinal cord motor neurons
Shrinkage, sclerosis and pallor of the corticospinal tracts	Diffuse astrocytic gliosis of spinal gray matter
Thinning of the spinal ventral roots and hypoglossal nerves	Atrophic and basophilic changes in surviving motor neurons
Atrophy of the somatic and bulbar muscles	Presence in surviving lower motor neurons of ubiquitinated inclusion bodies with
	thread-like, skein-like or compact morphology
	Variable depletion of giant pyramidal neurons (Betz cells) in the motor cortex
	Variable astrocytic gliosis in the gray matter of the motor cortex and underlying subcortical white matter
	Evidence of microglial activation in pathologically affected areas
	Cytoplasmic aggregate inclusions within glial cell

Table 2: Aspects which make motor neurons more vulnerable for degeneration compared to other cell types (Tabel adapted from *Ferrainolo, L. et al. (2011)*)⁵⁹.

Motor neuron vulnerability
Large cell size and long, large-volume axonal compartment
High reliance on a robust cytoskeleton and axonal transport mechanisms
High metabolic demands
High reliance on optimal mitochondrial function
Increased generation of reactive oxygen species (ROS) and a tendency towards oxidative stress Possible motor neuron-specific and/or spinal cord-specific mitochondrial properties
High physiological reliance on signaling through ROS and reactive nitrogen species
High intrinsic oxidative stress
Vulnerability to excitotoxicity and dysregulation of intracellular calcium homeostasis
High expression of calcium-permeable α-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA) receptors lacking the GluR2 subunit
Low expression of calcium-buffering proteins
High reliance on efficient synaptic glutamate reuptake transport mechanisms
Reduced capacity for heat shock response and chaperone activity
Possibly reduced proteasome function
High physiological expression of particular proteins, such as superoxide dismutase 1 (SOD1)
Greater vulnerability to toxicity in the presence of mutant SOD1

Table 3: ALS models which has previously been used to investigate ALS mechanisms and pathology (Table adapted from Gama Sosa, M., et al (2012))¹⁶.

Animal model	Strategy	Pathology	Reference
Caenorhabditis elegans	Pan neuronal expression of wt human TDP-43	Motor neuron dysfunction, abnormal motor neuron synapses, and uncoordinated phenotype	²⁸¹
Fruit Fly <i>Drosophila melanogaster</i>	Motor neuron and photoreceptor GAL4-UAS driven expression of wt and mutant hSOD1	Neither loss of motor neurons nor retinal degeneration was observed, although motor neuron expression of either wt or mutant hSOD1 led to progressive motor dysfunction. Earlier attempts at expressing hSOD1 in motor neurons increased longevity and could rescue the shortened lifespan of a dSod null mutant	^{282 283 125} , ,
ZebraWsh <i>Danio rerio</i>	In vitro transcribed RNAs of TDP-43 (TARDBP wt and mutants G348C, A382T, A315T) were injected into 1–4 cell stage blastulae	Preferential vulnerability of motor neurons to toxicity of TDP43 mutants; shorter motor neurons axons, premature and excessive branching, and swimming deficits	²⁸⁴
Mouse <i>Mus musculus</i>	Pan-neuronal constitutive overexpression of TDP-43 or mutant TDP-43 (A315T) via pronuclear injection of transgenic cassettes using PrP and Thy1.2 promoters	Expression of TDP-43 wt and mutant proteins results in variable pathological phenotypes depending on the animal model	^{285 286} ,
	Forebrain-targeted Tet-OV conditional expression of TDP-43. Bigenic mice were created with cassettes harboring the Camk2a promoter driving the expression of Tet-OV tTA and the tetracycline-responsive promoter to drive hTDP-43 (tetO-hTDP). hTDP-43 was preferentially induced (in the absence of doxycycline) in forebrain neurons	Selective degeneration of hippocampal dentate gyrus and neocortical neurons along with dysregulation and loss of endogenous TDP-43 and presence of rare phosphorylated and ubiquitinated TDP-43 inclusions in affected neurons. Transgenic mice develop a motor phenotype associated with spasticity	
	Pronuclear injection of a human SOD1 transgene containing the G93A substitution associated with familial ALS	Hemizygous transgenic mice develop motor neuron disease associated with progressive limb paralysis and death by 5–6 months of age	
	CD11b-TKmt-30 mice contain a transgenic cassette in which the	After ganciclovir treatment, selective ablation of proliferating microglial cells occurs. Ablation of	

DISEASE MECHANISM AND MODELS FOR ALS

	<p>HSV-1 TKmt-30 gene is expressed under the control of the CD11b promoter, which directs high levels of expression in macrophages. Proliferating microglial cells in the brain can be selectively eliminated by ganciclovir treatment</p>	<p>proliferating microglia does not affect motor neuron degeneration in a mouse model of ALS but causes increased amyloid deposition in a mouse model of AD, suggesting that microglia play a critical role in restricting AD associated senile plaque formation</p>
Rat <i>Rattus norvegicus</i>	<p>Transgenesis with a 22-kb normal and retroviral mutant (M337 V) human TDP-43 gene extracted from a BAC</p>	<p>Constitutive expression of the mutant TDP-43 causes early postnatal death in transgenic rats. Founders develop weakness in the limbs and do not survive past 29 days</p>
	<p>Tet-OV inducible system for TDP-43 expression. tTA protein expression controlled by a CMV enhancer fused to the chicken actin promoter (CAG). Normal and mutant TDP-43 cDNAs were expressed under the control of a tTA-dependent promoter constructed by the juxtaposition of multiple tetracycline-responsive elements (TRE, tet-O) upstream from the minimal CMV promoter</p>	<p>Overexpression of mutant TDP-43 but not wt protein induce progressive widespread neurodegeneration affecting predominantly the motor system with denervation atrophy of skeletal muscles</p>

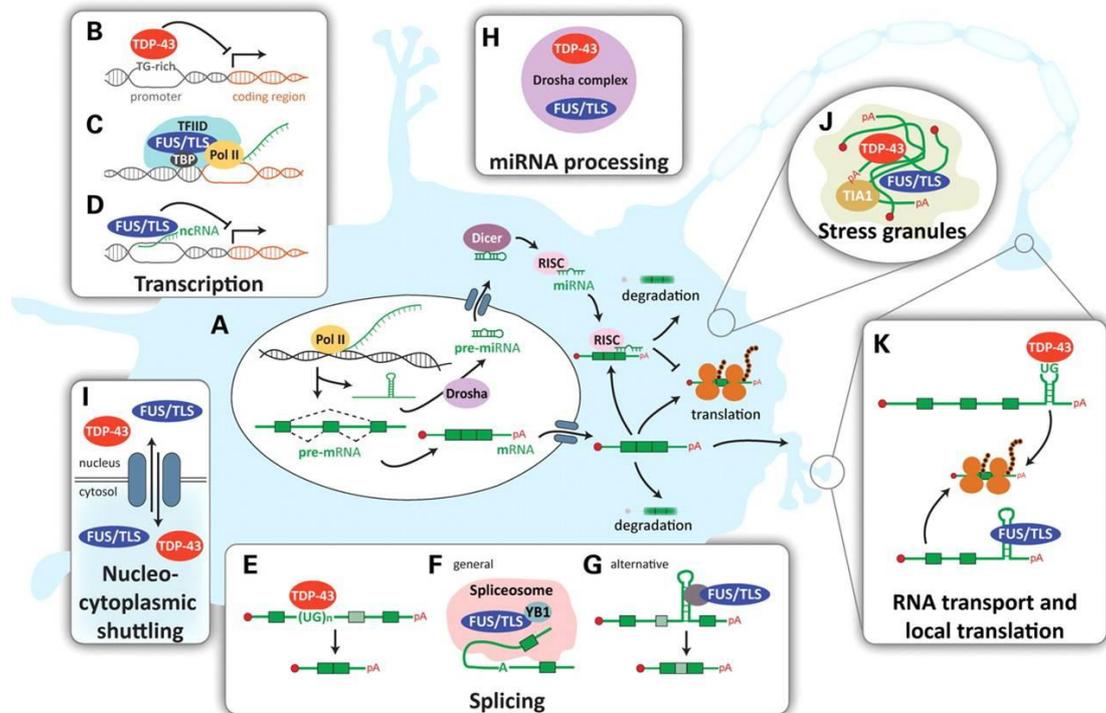
Figure 1: Processes in which TDP-43/FUS seems to be involved

Fig.1: Proposed roles of TDP-43 and FUS/TLS. (A) Overview of crucial steps of RNA processing from transcription to translation or degradation. (B) TDP-43 binds single-stranded TG-rich elements in the promoter regions. This causes transcription of the downstream gene to stop. (C) FUS has been associated with general transcriptional machinery processes. (D) In response to DNA damage, FUS is recruited in the promoter region of cyclin D1 (CCND1) through sense and antisense non-coding RNAs (ncRNAs). This represses CCND1 transcription. (E) TDP-43 binds towards UG area's in intronic regions prior to alternatively spliced exons. This enhances their exclusion. (F) FUS is identified to bind the spliceosome, and (G) was shown to promote exon inclusion in H-ras mRNA by binding to regulatory elements positioned on the downstream intron (H) Both proteins are associated with miRNA processing and (I) both shuttle between the nucleus and the cytosol. (J) In the cytosol they form complexes with mRNAs and other RNA binding proteins. (K) TDP-43 and FUS are both involved in the axonal transport of mRNAs to dendritic spines and/or the axonal terminal where they might facilitate local translation. (Image adapted from Lagier-Tourenne, C., et al. 2010)²⁶⁴.

Figure 2: Proposed mechanisms of SOD-1 mediated ALS

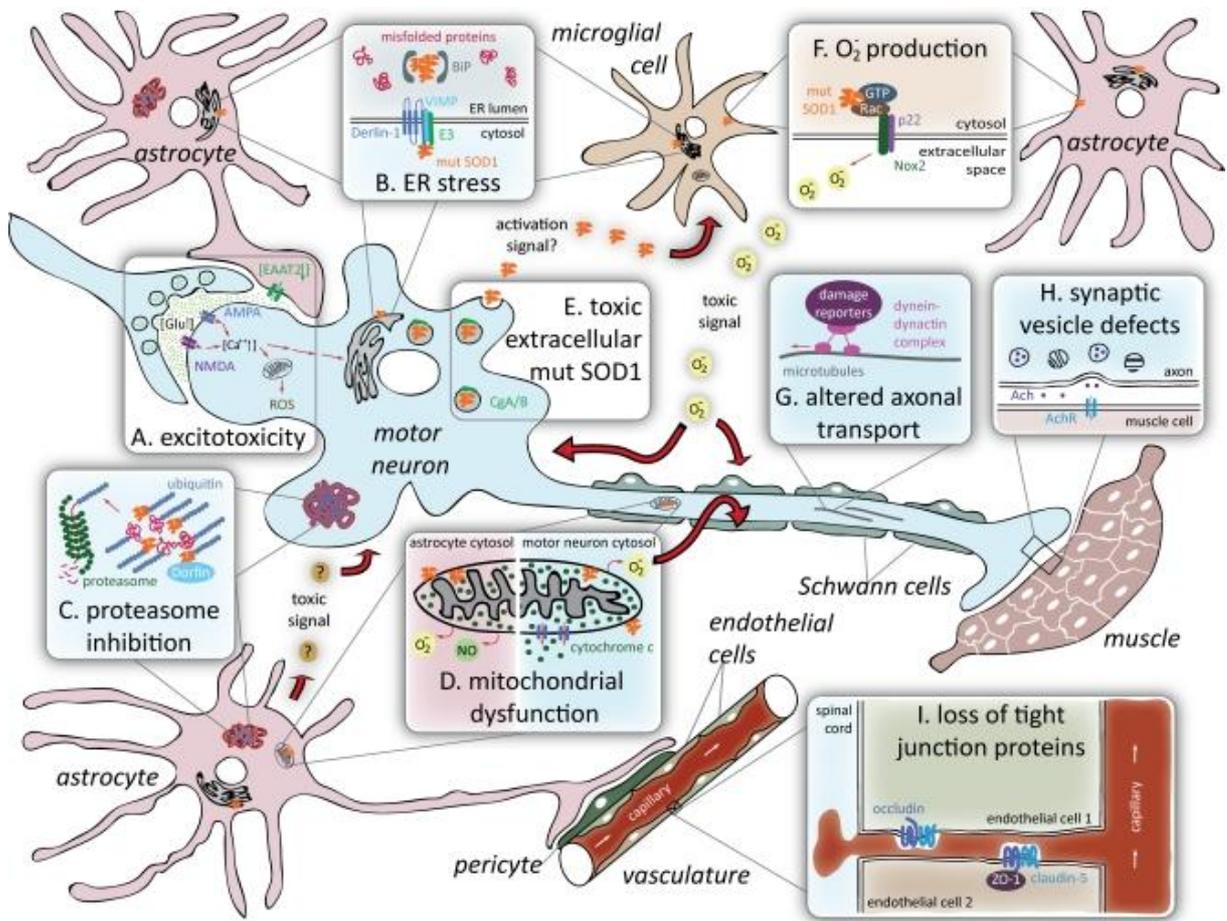


Fig.2: Proposed mechanisms of SOD1-mediated ALS. A. Excitotoxicity B. ER stress is induced by protein aggregations (see text for details). C. Proteasome dysfunction caused by “overload” of the proteasome degradation pathway. This is caused by protein aggregations and ubiquitinated misfolded protein and lead to damage astrocytes and motor neurons. D. Mitochondrial dysfunction. E Toxic secretion of mutant SOD-1 into the extracellular environment. F. Non-cell autonomous regulation from microglia or astrocytes can damage neighboring motor neurons. G. Altered axonal transport: Retrograde transported stress-related proteins were reported in mutant SOD1-expressing motor neurons. H. Synaptic vesicle defects. I. Loss of tight junction proteins within capillary endothelial cells lead to disruption of the BBB/BSCB. This may cause vascular ruptures. (Image is adapted from Ilieva, H., et al., 2009) ²⁶⁴.