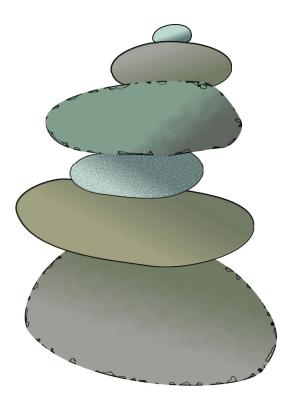
# NMDA Antagonism in the treatment of Major Depressive Disorder



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#### Abstract

With a lifetime prevalence of over 16% and spread over all social classes, major depressive disorder (MDD) is one of the leading causes for psychosocial disability. Although there is no established mechanism for depression most drugs used in pharmacotherapy focus on the monoamine-deficiency hypothesis and inhibit the reuptake or metabolism of serotonin or norepinephrine in order to raise their plasmalevels. With over 20% of patients being resistant to treatment, and very low remission and high relapse rates, new strategies and methods for this treatment-resistant form of depression are warranted. In recent years the use of N-methyl-D-aspartate (NMDA) antagonists, and in particular the well known dissociative drug ketamine, in the treatment of clinical depression has come under increasing interest. This administration is less rigorous than more invasive treatments like deep brain stimulation. Recently the mechanism of action of this acute response, at sub-psychotomimetic doses, has been linked to eukaryotic elongation factor 2 (eEF2) kinase inhibition and desupression of brain-derived neurotrophic factor (BDNF) translation. This supports other research implying synaptic plasticity as an important factor in treating MDD. Chronic administration of ketamine is however linked to impairments of verbal fluency, cognitive processing speed, and verbal learning and its other cognitive effects are associated with potential abuse. In this report we will outline the molecular mechanisms involved in NMDA antagonism, the effects on other neurotransmittersystems and glutamate signaling and how these relate to the positive effects on MDD. The major transduction pathways will be outlined and potential related targets highlighted.

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## Acronyms

4E-BP 4E binding protein.

ACC anterior cingulate cortex.
AD antidepressant.
Akt protein kinase B.
ALS amyotrophic lateral sclerosis.
AMPA α-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid.
AMPAR AMPA receptor.
Arc activity-regulated cytoskeleton-associated protein.

BBB blood-brain barrier.BDNF brain-derived neurotrophic factor.BPD bipolar disorder.

CaMKII Ca2+/calmodulin-dependent protein kinase.

**CBT** cognitive behavioral therapy.

**CNS** central nervous system.

CUS chronic unpredictable stress.

E-LTP early-LTP.

**eEF2** eukaryotic elongation factor 2.

**EPSC** excitatory postsynaptic current.

**EPSP** excitatory postsynaptic potential.

ERK extracellular signal-regulated kinase.

FST forced swim test.

GNDF glial-derived neurotrophic factor.GPCR G-protein coupled receptor.GSK-3 glycogen synthase kinase 3.

**ICV** intracerebralventricular.

**IPT** interpersonal psychotherapy.

KA kainate.

KO knockout.

L-LTP late-LTP.

LH learned helplessness.

LTD long-term depression.

LTP long-term potentiation.

MAOI monoamine oxidase inhibitor.MDD major depressive disorder.mGluR metabotropic glutamate receptor.mTOR mammalian target of rapamycin.

NBQX 2,3-dihydroxy-6-nitro-7-sulfamoyl-benzo[f]quinoxaline-2,3-dione.
NMDA *N*-methyl-D-aspartate.
NMDAR NMDA receptor.
NSFT novelty suppressed feeding test.

p70S6K p70S6 kinase.

PFC prefrontal fortex.

PP1 protein phosphatase 1.

PSD-95 postsynaptic density protein 95.

**SNRI** serotonin-norepinephrine reuptake inhibitor. **SSRI** selective serotonin reuptake inhibitor.

TCA tricyclic antidepressants.
TRD treatment-resistant depression.
TREK-1 TWIK-1-related K+ channel.
TrkB Tropomyosin receptor kinase B.
TST tail suspension test.

UCMS unpredictable chronic mild stress.

**VDCC** voltage-dependent Ca<sup>2+</sup> channel. **VEGF** vascular endothelial growth factor.

WKY Wistar-Kyoto. WT wild-type.

## 1 Major Depressive Dissorder

Clinical depression, or MDD, is a major cause of disability worldwide. The lifetime prevalence of this disorder varies widely, but in most countries the percentage of people that experience depression during their lives falls within 8-12%<sup>[1]</sup>. In western countries, particularly in North American, this number can rise up to nearly 17% and is spread throughout all demographics, with slightly elevated numbers for low-income backgrounds<sup>[2]</sup>. This indicates that MDD is relatively common and widely distributed among the population with a significant impact on the medical system.

#### 1.1 Symptoms and Treatment

MDD is characterized by a distinct change of mood, distinguished by sadness or irritability and associated with at least several psychophysiological changes such as disturbances in sleep, appetite, or reduced sex drive. Other symptoms may include constipation, general anhedonia (inability to experience pleasure in work or other activities), and suicidal thoughts<sup>[3]</sup>. These changes have to persist for several weeks and interfere substantially with work and social relations. Sometimes other related disorders are misdiagnosed such as dysthymia, which has milder symptoms, but is of a chronic variety. Some patients with MDD may encounter manic episodes that consist of additional symptoms such as hyperactivity, euphoria and an increase in pleasureseeking behavior. Patients experiencing these episodes are diagnosed with a distinct illness termed bipolar disorder (BPD)<sup>[4]</sup>.

The response to MDD treatment is generally inconsistent and there is no established mechanism<sup>[3]</sup>. The typical treatments for MDD are psychotherapy and pharmacotherapy. Both psychotherapy and medication have been shown to be viable treatments for MDD<sup>[5]</sup>. However where psychotherapy can be used more successfully in preventive follow-up care, pharmacotherapy is somewhat more successful in the treatment of dysthymia<sup>[6]</sup>. The combination of psychotherapy and pharmacotherapy is more effective than either of these therapies alone<sup>[7;8]</sup>.

Psychotherapeutic treatments usually employed are cognitive behavioral therapy (CBT), interpersonal psychotherapy (IPT), psychodynamic therapy, and upportive counselling, but will depend on the background and judgment of the treating psychotherapist. For mild depression psychotherapy is the first line of treatment<sup>[9]</sup>. Pharmacotherapeutic treatments typically focus on the monoamine-deficiency hypothesis where supposed deficiencies of endogenous mood-regulatory neurotransmitters such as serotonin, dopamine or norepinephrine are enhanced using selective serotonin reuptake inhibitors (SSRIs), serotonin-norepinephrine reuptake inhibitors (SNRIs) or monoamine oxidase inhibitors (MAOIs)<sup>[3]</sup>.

In some acute depressive treatments the classical tricyclic antidepressantss (TCAs) are used, although their current application is limited because of their general sideeffects and limited specificity<sup>[10;11]</sup>. Some melatonergic agonists, such as agomelatine, are reported to have antidepressant (AD) effects or may treat sleep disorders associated with MDD and are marketed as such<sup>[12]</sup>. MDD may occasionally be treated using lithium, although this is more common in BPD alongside an anticonvulsant or atypical antipsychotic. One of the major shortcomings of current antidepressant drugs is that several weeks or months of treatment are needed before improvements are observed.

#### 1.2 Animal-models and behavioral tests

In order to study the behavioral effects of treatments under controlled circumstances, a variation of translational animal-models are used to simulate the physical and psychological states associated with MDD. Most of these models focus on manipulating induced stress and behavioral despair and may use either mice or rats in their methodology<sup>[13;14]</sup>. These stress-based models can be categorized as either acute or chronic stress. Sucrose consumption is sometimes used as an inverse behavioral read-out for anhedonia, although its reproducibility has been criticized because it often depends on whether treatment with AD-medication is chronic or acute<sup>[14]</sup>.

Acute stress-based tests - There are several acute tests employing behavioral despair that are used to indicate the efficacy of anti-depressant drugs and treatments. The most commonly used are the forced swim test (FST) and tail suspension test (TST). The performance in these tests has been used as a quantification of AD-effects for many decades and still counts as the 'gold standard' in AD research<sup>[15;16]</sup>. Other related tests are the learned helplessness (LH) and novelty suppressed feeding test (NSFT) in which the behavior of the animal is observed in either a conditioned environment where the animal has been taught to be helpless, or a completely new environment where the animal shows a delayed feeding response, respectively.

**Chronic stress-based models** - A wide variety of models have been developed using chronic stress to induce a semi-permanent state of depression in animal subjects. These stressors are typically randomly selected from a variety of methods such as water or food deprivation, 45 degrees cage tilt, exposure to rat faeces, cage overcrowding, wet bedding, overnight illumination, dark exposure during normal light cycle, cold bedding, acoustic disturbance (120 dB), strobe lights, or cagemate rotation and may last for an arbitrary length of 1 up to 12 hours<sup>[17]</sup>. These methods are also referred to as unpredictable chronic mild stress (UCMS) or chronic unpredictable stress (CUS). Some models for dysfunctional parenting, such as maternal deprivation during early postnatal life, may also be used as a chronic stress model.

**Genetic models for depression** - A number of genetic animal models have been developed that show endophenotypes related to depression. Wistar-Kyoto (WKY) rats were developed using selective inbreeding based on poor FST performance and show enhanced susceptibility to depressed states and resistance to a number of antidepressants, thus possibly providing a genetic base for treatment-resistent forms of depression<sup>[18;19]</sup>. The TWIK-1-related K+ channel (TREK-1) knockout mouse-model on the other hand shows increased resilience to depression in FST and TST, implicating an ion channel in depression-like animal behavior<sup>[20]</sup>.

## 2 Synaptic Plasticity in Depression

Moving beyond the monoamine hypothesis of depression, the past 10 years we have seen many molecular and cellular studies of stress, depression and antidepressants demonstrating opposing actions on the expression of particular neurotrophic factors in limbic brain regions that regulate mood and cognition. These neurotrophic proteins are growth factors that are neurodevelopmentally expressed and regulate neuroplasticity and cellular resistance in the brain<sup>[21]</sup>. During homeostasis in the differentiated adult brain, the contacts between neurons are continuously being replaced and renewed. The presynaptic site may undergo formation of new synapses due to enhanced axonal growth, likewise terminal degeneration may eliminate existing ones. The size on the dendritic tree, or its spine density, and changes in the organization of glial cells, which maintain homeostasis in the brain, may cause the number of postsynaptic sites to increase or decrease. Depression has been associated with hypocampal neuronal atrophy and synaptic loss<sup>[22]</sup>. After chronic treatment with AD drugs however, an increase in neurotrophic factor expression and enhanced synaptic plasticity is observed<sup>[23;24]</sup>.

Two forms of synaptic plasticity that are well understood are long-term potentiation (LTP) and long-term depression (LTD), which are enhancement and reduction of signal transmission between neurons respectively, thus LTP and LTD are an indication of synaptic strength and dendritic spine growth and retraction respectively<sup>[25]</sup>. Changes in synaptic strength are established through both pre- and postsynaptic mechanisms such as synthesis and movement of synaptic receptors and other proteins. Rapid changes, during early-LTP (E-LTP) for instance, depend on the movement of pre-existing proteins<sup>[26]</sup>, while RNA translation and changes in gene expression are a factor in late-LTP (L-LTP)<sup>[27]</sup>. LTD similarly involves, likely oppositional, adaptations to gene expression and protein metabolism.

During synaptic plasticity many signaling systems are involved in unison between neurons. Some intracellular signal transduction pathways are crucial in this regard and central to synaptic plasticity initiation is calcium influx through several ion channels. This is particularly mediated by a number of glutamate related receptors and release mechanisms that are thus deeply involved in brain plasticity.

#### 2.1 Glutamate and Neural Plasticity

Glutamate is the most abundant and important excitatory neurotransmitter in the brain. It is released by exocytosis and is present at very high concentrations in synaptic vesicles<sup>[28]</sup>. It is a non-essential amino acid that can be synthesized via a number of routes and as such plays an important role in many metabolic pathways<sup>[29]</sup>. Abnormalities in its metabolism have long been understood to play an important role in degenerative neurological disorders<sup>[30;31]</sup>.

Glutamatergic receptors in the central nervous system (CNS) consist of two major subtypes, ionotropic and metabotropic. The G-protein coupled receptors (GPCRs) of the metabotropic glutamate receptor (mGluR) subtype consist of eight identified proteins that can be divided in to three subgroups, which are based on their signaling pathways. The ionotropic subtype of glutamatergic receptors consists of three ligand-gated ion channels that each are formed from multiple, and varying, subunits. These are the  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA), kainate (KA) and NMDA receptors respectively, which may be found pre-, post-and extra-synaptically. Post-synaptically these receptors are anchored in a scaffolding complex with postsynaptic density protein 95 (PSD-95), which is a potent regulator of synaptic strength through its control over AMPA receptor (AMPAR) concentrations<sup>[32]</sup>.

These receptors are known to play an important role in synaptic plasticity. Expression of AMPARs is one of the best studied molecular correlates of synaptic strength at excitatory synapses and increases in this expression under LTP is related to dendritic spine growth and spine-head size<sup>[33]</sup>. During LTP, AMPARs containing the long-tailed subunits GluR1, GluR4 and GluR2L are added, while during the constitutive cycling of LTD the short-tailed GluR2, GluR3 and GluR4c subunits participate in AMPAR removal<sup>[34]</sup>. Similarly, following LTP and LTD, NMDA receptors (NMDARs) undergo trafficking and changes to GluN2 subunit ratios and it has been found that NMDAR synaptic response consistently decreases during LTD<sup>[35;36]</sup>.

There is increasing evidence that alterations in the glutamatergic ion-channels AM-PAR and NMDAR are involved in mood disorders such as MDD and BPD as their expression levels seem to be decreased in such patients<sup>[37]</sup>. Most AMPA receptors are heterotetrameric and primarily consist of the GluR2 subunit coupled as a 'dimer of dimers' with GluR1, Glur3 or Glur4<sup>[38]</sup>. Sodium and potassium are the principal ions for which AMPARs are permeable. This in contrast to NMDARs which primarily gate sodium and calcium ions, although potassium may be transferred out of the cell in a voltage dependent manner<sup>[39]</sup>. NMDARs form heterotetramers consisting of two GluN1 and two GluN2 subunits. Where AMPARs are activated at four glutamate binding sites, one at each subunit, the NMDARs have a glutumate and a glycine binding site and require co-activation.

After glutamate binds to postsynaptic AMPARs and NMDARs, Na<sup>+</sup> flows in to the postsynaptic cell, which results in excitatory postsynaptic potential (EPSP), or depolarization of the membrane. At resting membrane potential however, NMDAR channels are blocked by Mg<sup>2+</sup> and can only open when AMPAR activation has led to depolarization, causing repulsion of Mg<sup>2+</sup> cations. This then allows the NMDARs to permeate Ca<sup>2+</sup> in to the cell which subsequently triggers AMPAR upregulation and a further increase in EPSP size, an indicator of LTP. The Ca<sup>2+</sup> influx also results in phosphorylated Ca<sup>2+</sup>/calmodulin-dependent protein kinase (CaMKII) which phosphorylates AMPARs and enhances their conductance<sup>[40]</sup>. CaMKII is known to be an important requirement for LTP initiation<sup>[41]</sup>.

#### 2.2 Glutamatergic Drugs

A significant number of patients do not respond to the normal treatments for MDD and develop treatment-resistant depression (TRD). It is estimated that nearly 50% of patients will not reach full remission with their first AD treatment<sup>[42]</sup> while nearly half of that (20-30%) will be completely non-responsive, even after subsequent adaptations to their treatment<sup>[43;44]</sup>. The medium and long term outcomes for patients that develop TRD are very serious as they are associated with a high rate of relapse, significant disability and mortality and residual symptoms leading to this high relapse and long term disability<sup>[45;46]</sup>. TRD patients may be more likely to suffer from comorbid physical

and mental disorders<sup>[47;48]</sup>. In general, patients with TRD have a significantly higher disease burden and cost of burden than other patients<sup>[43;48]</sup>. Therefore new treatments and medications for this treatment-resistant form of depression are needed. Ever since NMDA-receptors and the glutamatergic system were implied in the mechanics of depression and anti-depressants in the 1990s their potential role as a target in the treatment of depression has increased<sup>[49–51]</sup>. This has resulted in a number of compounds that modulate the glutamate system, particularly ketamine which is a well-known dissociative anesthetic.

**Ketamine** - After many animal studies the first placebo-controlled, double-blinded trial with single dose ketamine, an NMDA antagonist, was published in 2000<sup>[52]</sup>. This showed the feasibility of NMDA-receptor modulation as a possible treatment for patients with MDD. Since then a number of randomized and open label trials have been conducted that support the claims of feasibility and show that the effects may last for days if not weeks in patients with TRD<sup>[53–57]</sup>. Treatment with ketamine also seems to reduce suicidality in this patient population<sup>[58]</sup>. Due to its long history of medicinal use and efficacy in TRD treatment, ketamine currently has a high interest as a potent medication for MDD.

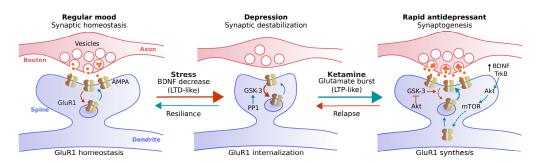
**Other glutamate modulators** - Among the variety of glutamatergic compounds the glutamate uptake enhancer and amyotrophic lateral sclerosis (ALS) medication Riluzole was explored in an open-label trial for TRD<sup>[59]</sup>. GLYX-13, a NMDAR glycine-site partial agonist, has been described to induce AD-like effects without the psychoactive side effects of ketamine<sup>[60]</sup>. Likewise the low-trapping and non-selective NMDA channel blocker AZD6765 (Lanicemine) has shown rapid, albeit short-lived, AD effects in patients with TRD<sup>[61]</sup>. The NR2B subunit selective NMDA antagonist CP-101,606 (Traxoprodil) has also shown efficacy in MDD treatment, without producing any of the dissociative reactions associated with most NMDA antagonists<sup>[62]</sup>.

**Proposed** - A number of NMDA antagonists and functional analogs of ketamine have been hypothesized to elicit similar AD effects and may be explored in future clinical research. These include the well known antitussive dextromethorphan<sup>[63]</sup> and the relatively new structural analogue of ketamine, methoxetamine<sup>[64]</sup>. It may or may not be warranted that these compounds are explored in future clinical studies.

## 3 Ketamine and NMDA Antagonism

Because NMDARs need agonism on both the glutamate and glycine binding site to open the ion channel, there are several methods by which they can be prevented from activation. There are competitive antagonists that inhibit glutamate binding; glycine antagonists which act at the glycine binding-site; uncompetitive channel blockers that prevent ions from passing the receptor; and finally there are non-competitive antagonists that bind to an allosteric site. Ketamine belongs to this final group of antagonists and although NMDARs may have several subunit compositions, ketamine does not appear to have a particular selectivity for these in its binding<sup>[65]</sup>.

Although post-synaptic NMDARs are primarily involved in gating sodium and calcium ions, presynaptic NMDARs have been shown to modulate glutamate release<sup>[66]</sup>. Low dose NMDA antagonism by ketamine in particular increases glutamate outflow



**Figure 1:** Schematic representation of the synaptogenic model of depression and subsequent ketamine induced recovery. Stress induces LTD-like synapse morphology to neuronal spines with decreased BDNF activity and AMPAR internalization. This results in neuronal atrophy in its depressed state. Ketamine then induces a glutamate burst that stimulates BDNF exocytosis and subsequent mammalian target of rapamycin (mTOR) activation which restores AMPAR signaling functionality reminiscent of LTP.

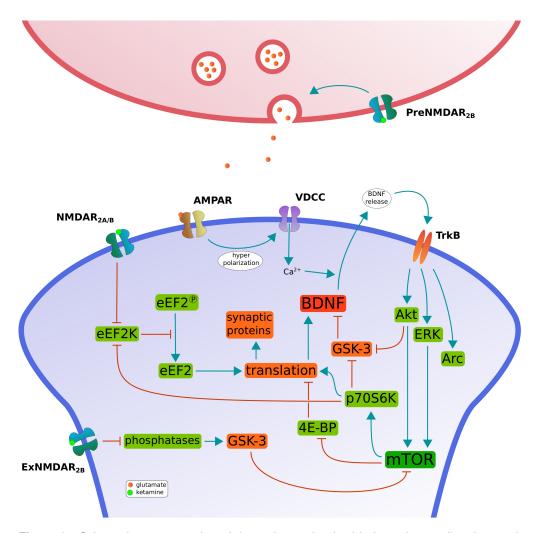
in the prefrontal fortex (PFC), which suggests that ketamine may thus increase glutamatergic neurotransmission at non-NMDA glutamate receptors<sup>[67]</sup>. While normally NMDAR activation directly leads to an influx of Ca<sup>2+</sup> ions and subsequent CaMKII phosphorylation, this increase of glutamatergic neurotransmission may cause Ca<sup>2+</sup> influx through hyperpolarization caused by AMPAR activation and resulting voltagedependent Ca<sup>2+</sup> channel (VDCC) opening.

Extra-synaptic NMDARs which, like the pre-synaptic receptors<sup>[68]</sup>, contain NR2B subunits<sup>[69]</sup>, are involved in a number of processes. ExNMDARs activation is particularly implicated in excitotoxicity and neuroprotection as part of LTD<sup>[70;71]</sup>. When extra-synaptic NR2B-containing NMDARs are excited under normal conditions they activate protein phosphatase 1 (PP1) and induce a feedback loop between glycogen synthase kinase 3 (GSK-3) and PP1<sup>[72]</sup>. Inhibition of these receptors conversely results in lower PP1 and GSK-3 deactivation, which in turn increases NR2B subunit levels<sup>[71]</sup>. This subunit increase has been observed in the antagonism by GLYX-13 and ketamine, which both led to increases in NR2B and GluR1 protein levels and a persistent enhancement of LTP<sup>[60]</sup>.

#### 3.1 Signaling Pathways

A number of signaling pathways have emerged that are modulated by NMDA antagonism and support the synaptogenic hypothesis of its fast acting and long lasting antidepressant effects. The AD effects of ketamine at least seem to be dependent on AMPAR throughput, as these effects are attenuated by the AMPAR antagonist 2,3-dihydroxy-6nitro-7-sulfamoyl-benzo[f]quinoxaline-2,3-dione (NBQX)<sup>[73;74]</sup>. Likewise, the long-lasting AD-effects of both ketamine and GLYX-13 treatment are partially abolished by NBQX pretreatment in either LH, TST or FST<sup>[60;74]</sup>. Further more, chronic treatment of WKY rats with low dose ketamine induced marked changes in the hippocampal AMPA/NMDA receptor ratio in favor of AMPAR, showing the importance of AMPAR in the long-lasting AD-effects of ketamine<sup>[75]</sup>.

The two major pathways currently under investigation are mediated by BDNF and



**Figure 2:** Schematic representation of the pathways involved in ketamine mediated synaptic plasticity. Antagonism of pre-synaptic NMDARs leads to glutamate release while post-synaptic suppression of NMDARs results in inhibition of eEF2 kinase and dephosphorylation of eEF2. This augments BDNF translation while the aforementioned glutamate release activates AMPARs which hyperpolarize the membrane and activate VDCCs. Calcium subsequently promotes BDNF release which may then activate Tropomyosin receptor kinase B (TrkB) receptors resulting in a signaling cascade that in turn activates protein kinase B (Akt) and extracellular signal-regulated kinase (ERK). Akt and ERK then activate mTOR which enables further translation of synaptic proteins, including BDNF, GluR1 (AMPA) and PSD-95, by activating p70S6 kinase (p70S6K) and inhibiting 4E binding proteins (4E-BPs). GSK-3, which inhibits BDNF expression, may be suppressed through phosphorylation by Akt or p70S6K. Likewise, stimulation of GSK-3 is suppressed by deregulation of phosphatases from extra-synaptic NMDAR antagonism, which may also result in mTOR mediation. Finally, eEF2 kinase is also suppressed by p70S6K, feeding in to the previously mentioned BDNF translation.

mTOR. In the following parts the role of these proteins and their relation in the overall pathway of synaptogenesis in NMDA antagonism will be highlighted. In fig. 1 a general overview of synaptic homeostasis, degerenation and genesis is presented culminating in the signaling pathway presented in fig. 2.

#### 3.1.1 mammalian target of rapamycin

mTOR, a serine/threonine protein kinase, is a regulatory protein involved in cell growth, proliferation, motility, survival, and protein synthesis<sup>[76]</sup>. The regulation of translation initiation by mTOR is exerted through a number of downstream proteins: p70S6K, eIF4E and eIF4B. Administration of rapamycin, the principal inhibitor of mTOR, disrupts the mTOR signaling pathway leading to reduced L-LTP<sup>[26]</sup>. It has been shown that patients with MDD have a marked deficit of mTOR and its downstream components which could contribute to the observed molecular pathology of this disorder<sup>[77]</sup>.

Rats exposed to CUS show these same deficits in synaptic proteins, as shown by immunoblotting of their phosphorylated activation states<sup>[77–80]</sup>. Exposure to CUS for a period of 3 weeks decreased spine density in layer V pyramidal neurons and decreased excitatory postsynaptic currents (EPSCs) and thus EPSPs<sup>[79]</sup>. Subsequently, administration of NMDA-antagonists (ketamine and Ro 25-6981) in these rats rapidly decreased the observed anhedonic and anxiogenic behavior and resulted in increased levels of these signaling proteins and increased numbers of new spine synapses in the PFC<sup>[78;79]</sup>. Other proteins influenced by mTOR are PSD-95, GluR1 and synapsin I, underlining its role in synaptogenic translation. These effects were completely blocked by pretreatment of the mTOR-inhibitor rapamycin, further supporting the role of mTOR in the synaptogenic pathway of ketamine administration.

Interestingly pretreatment with the selective AMPA inhibitor NBQX, 10 minutes before ketamine administration, resulted in complete blockade of induction of phosphorylated 4E-BP1, p70S6K and mTOR in the PFC<sup>[78]</sup>. These results underline the important role of AMPAR transmission in synaptogenesis, which is emphasized by the translation of the AMPA subunit GluR1 via the mTOR pathway.

#### 3.1.2 ketamine mediated BDNF release

BDNF, which is critical for axonal growth and neuronal survival, has been shown to be a significant biomarker in stress models and depressed patients<sup>[81]</sup>. Hippocampal levels of BDNF are decreased in animal models using immobilization<sup>[82–84]</sup>, social isolation<sup>[85]</sup>, maternal deprivation<sup>[86]</sup>, and swim stress<sup>[87]</sup>. Postmortem hippocampal levels of depressed suicide victims have also been found to be decreased<sup>[88–90]</sup>. And where serum levels of BDNF in depressed patients are lowered<sup>[90–92]</sup> these same levels increase during AD treatment<sup>[88;92–94]</sup>, showing that ADs influence this same neurotrophic mechanism.

Although no changes in BDNF could be detected in plasma levels of patients during the initial and rapid AD response of ketamine administration<sup>[95]</sup>, BDNF serum levels were in fact increased in chronic ketamine users<sup>[96]</sup>. Furthermore, mice with inducible BDNF knockout (KO) do not show the fast-acting, 30 minutes, AD-response of ketamine as seen in wild-types (WTs)<sup>[17]</sup>. This suggests that this AD-response relies on fast BDNF expression. Finally there are clear signs that BDNF-release is involved both pre-and post-LTP<sup>[97;98]</sup>

BDNF primarily acts by binding to TrkB which is its main receptor. This results in phosphorylation, and thus activation, of Akt and ERK which are both involved in the regulation of synaptic protein synthesis<sup>[99]</sup>. It has also been found that TrkB KOs result in ketamine-insensitive fenotypes in both FST and NSFT<sup>[17]</sup>. One striking discovery is that ketamine NMDAR antagonism can lead to inhibition of eEF2 kinase, which results in dephosphorylation of eEF2 and desupression of BDNF translation<sup>[17]</sup>. The aforementioned AMPAR and VDCC regulated calcium influx may finally result in BDNF exocytosis where it can activate TrkB<sup>[100]</sup>. The glutamate modulator riluzole likewise enhances BDNF, together with other signaling proteins like vascular endothelial growth factor (VEGF) and glial-derived neurotrophic factor (GNDF)<sup>[101]</sup>.

#### 3.1.3 related targets

**GSK-3** - As previously mentioned, GSK-3 has an inhibitory effect on NR2B expression, however it also mediates synaptic stimulation of mTOR<sup>[102]</sup> and is required in the phosphorylation of PSD-95 and its mobilization in LTD<sup>[103]</sup>. Inhibition of GSK-3 also seems to be necessary for the rapid antidepressant effect of ketamine in the LH-paradigm in mice<sup>[104]</sup>, although this regulation of its activity was not detected in subsequent efforts<sup>[105]</sup>. Thus, GSK-3 inhibition may be a viable target for future anti-depressant research.

**Arc** - This activity-regulated cytoskeleton-associated protein (Arc) has involvement in both LTP and LTD<sup>[106]</sup> and is an important regulator of AMPAR endocytosis and traffick-ing<sup>[107]</sup>. Ketamine induced the expression of Arc in the early stages of its downstream effects, with a peak after 1 to 2 hours, in a transient time course<sup>[78]</sup>. Potentially this identifies it as a component of E-LTP in ketamine's mechanism of action, mediating its initial and fast-acting effects.

**ERK and Akt** - As discussed earlier, these proteins are activated by TrkB stimulation and are involved with synaptic protein synthesis. Rats exposed to CUS show reduced phosphorylation of these proteins<sup>[80]</sup>. They are consequently observed to increase in concentration after ketamine administration, but their levels are normalized with pre-administration of NBQX emphasizing the role of AMPA activation<sup>[78]</sup>. Further more, intracerebralventricular (ICV) administration of selective ERK (U0126) or Akt (LY294002) inhibitors blocked the AD actions of ketamine in FST and NSFT<sup>[78]</sup>. Both are also identified in mTOR activation, providing another key route in the overall pathway<sup>[108;109]</sup>.

## 4 Discussion

We have seen that there are several pathways to consider in the short and long-term anti-depressant effects of ketamine and NMDA-antagonists. There is clear evidence that glutamatergic dysfunction plays an important role in the pathophysiology of depression and a number of signaling pathways can be considered as major targets.

Within the observed pathways we can see an interdependence emerging between mTOR and BDNF. Although BDNF clearly activates mTOR and its GluR1 expression<sup>[110]</sup> and the initial findings that rapamycin pre-treatment blocks the acute effects of ketamine could not be replicated<sup>[17]</sup>, which may have come from particular technical differences in the study<sup>[111]</sup>, there may still be some viable angles to approach mTOR as a key target.

Considering that the NR2B selective NMDA antagonist Traxoprodil is still effective in MDD treatment we may conclude that these routes in the pathway, mediated by this receptor-subtype, are most relevant to the observed outcomes. The equally rapid response to Traxoprodil, as compared to Ketamine, suggests that the post-synaptic NR2A pathway, which has been identified to disable eEF2 kinase, is not required for this result. However, mTOR activated p70S6K regulates eEF2 kinase as well, providing a different route for its inhibition<sup>[112;113]</sup>. Likewise we have seen that GSK-3 is a potential requisite in the observed activities, which may also be attenuated through NR2B receptors alone. The robust and long-term effects of both ketamine an GLYX-13 may also be a strong indication that they promote enhancement of 'metaplasticity'<sup>[60]</sup>, showing changes in plasticity that last much longer than their initial triggering mechanism, where again we see that NR2B containing NMDARs play a critical role<sup>[114]</sup>.

However, observing some of the potential long-term effects of mTOR modulation, we may consider its potential role in Alzheimers progression<sup>[115–117]</sup>. Although repeated ketamine use may cause impairment to cognitive processing speed and verbal fluency, no such specific effects have become apparent as of yet<sup>[118]</sup>. Concerns have previously been raised about ketamine's well known cognitive side-effects which occur during typical IV administration<sup>[119]</sup>. These concerns may be mediated by the fact that ketamine's robust AD-effects are equally effective at sub-psychotomimetic doses. Similarly the route of administration may be augmented to include oral delivery, which still has moderate bioavailability<sup>[120]</sup>. Making the drug more suitable for outpatient and nonprocedural psychiatric care.

Although many other molecules which act at some of the proposed targets show AD-like properties, the antagonist action on the NMDA receptor is currently the only clinically validated target<sup>[51]</sup>. However the current developments around this subject are continuously evolving with many clinical studies being conducted with related compounds. Primarily the elaboration of these mechanisms may result in novel directions for the development of fast-acting antidepressants and potential involvement of other neuropsychiatric disorders.

## References

- [1] Andrade, L., Caraveo-Anduaga, J. J., Berglund, P., Bijl, R. V., Graaf, R. D., Vollebergh, W., Dragomirecka, E., Kohn, R., Keller, M., Kessler, R. C., Kawakami, N., Kiliç, C., Offord, D., Ustun, T. B., and Wittchen, H.-U. (2003) The epidemiology of major depressive episodes: results from the International Consortium of Psychiatric Epidemiology (ICPE) Surveys. Int J Methods Psychiatr Res 12, 3–21.
- [2] Kessler, R. C., Berglund, P., Demler, O., Jin, R., Koretz, D., Merikangas, K. R., Rush, A. J., Walters, E. E., and Wang, P. S. (2003) The epidemiology of major depressive disorder: results from the National Comorbidity Survey Replication (NCS-R). *JAMA 289*, 3095– 105.
- [3] Belmaker, R. H., and Agam, G. (2008) Major depressive disorder. N. Engl. J. Med. 358, 55–68.
- [4] Belmaker, R. H. Encyclopedia of Psychopharmacology; Springer, 2010.
- [5] de Maat, S. M., Dekker, J., Schoevers, R. A., and de Jonghe, F. (2007) Relative efficacy of psychotherapy and combined therapy in the treatment of depression: a meta-analysis. *Eur. Psychiatry 22*, 1–8.
- [6] Imel, Z. E., Malterer, M. B., McKay, K. M., and Wampold, B. E. (2008) A meta-analysis of psychotherapy and medication in unipolar depression and dysthymia. J Affect Disord 110, 197–206.
- [7] Cuijpers, P., van Straten, A., Warmerdam, L., and Andersson, G. (2009) Psychotherapy versus the combination of psychotherapy and pharmacotherapy in the treatment of depression: a meta-analysis. *Depress Anxiety 26*, 279–88.
- [8] Cuijpers, P., Dekker, J., Hollon, S. D., and Andersson, G. (2009) Adding psychotherapy to pharmacotherapy in the treatment of depressive disorders in adults: a meta-analysis. *J Clin Psychiatry 70*, 1219– 29.
- [9] WHO, (2012) World Health Organization - Depression factsheet. http://www.who.int/mediacentre/factsheets/fs369/
- [10] Furukawa, T., McGuire, H., and Barbui, C. (2003) Low dosage tricyclic antidepressants for depression. *Cochrane Database Syst Rev* CD003197.
- [11] MacGillivray, S., Arroll, B., Hatcher, S., Ogston, S., Reid, I., Sullivan, F., Williams, B., and Crombie, I. (2003) Efficacy and tolerability of selective serotonin reuptake inhibitors compared with tricyclic antidepressants in depression treated in primary care: systematic review and meta-analysis. *BMJ 326*, 1014.
- [12] Norman, T. R. (2013) Agomelatine, melatonin and depressive disorder. *Expert Opin Investig Drugs 22*, 407–10.
- [13] Cryan, J. F., and Slattery, D. A. (2007) Animal models of mood disorders: Recent developments. *Curr Opin Psychiatry 20*, 1–7.
- [14] O'Leary, O. F., and Cryan, J. F. (2013) Towards translational rodent models of depression. *Cell and Tissue Research* 1–13.
- [15] Porsolt, R. D., Le Pichon, M., and Jalfre, M. (1977) Depression: a new animal model sensitive to antidepressant treatments. *Nature 266*, 730–732.
- [16] Petit-Demouliere, B., Chenu, F., and Bourin, M. (2005) Forced swimming test in mice: a review of antidepressant activity. *Psychopharmacology (Berl)* 177, 245–255.
- [17] Autry, A. E., Adachi, M., Nosyreva, E., Na, E. S.,

Los, M. F., fei Cheng, P., Kavalali, E. T., and Monteggia, L. M. (2011) NMDA receptor blockade at rest triggers rapid behavioural antidepressant responses. *Nature* 475, 91–5.

- [18] Lahmame, A., del Arco, C., Pazos, A., Yritia, M., and Armario, A. (1997) Are Wistar-Kyoto rats a genetic animal model of depression resistant to antidepressants? *Eur. J. Pharmacol.* 337, 115–23.
- [19] Will, C. C., Aird, F., and Redei, E. E. (2003) Selectively bred Wistar-Kyoto rats: an animal model of depression and hyper-responsiveness to antidepressants. *Mol. Psychiatry* 8, 925–32.
- [20] Heurteaux, C., Lucas, G., Guy, N., Yacoubi, M. E., Thümmler, S., Peng, X.-D., Noble, F., Blondeau, N., Widmann, C., Borsotto, M., Gobbi, G., Vaugeois, J.-M., Debonnel, G., and Lazdunski, M. (2006) Deletion of the background potassium channel TREK-1 results in a depression-resistant phenotype. *Nat. Neurosci. 9*, 1134–41.
- [21] Duman, R. S., and Monteggia, L. M. (2006) A neurotrophic model for stress-related mood disorders. *Biol. Psychiatry 59*, 1116–27.
- [22] Cole, J., Costafreda, S. G., McGuffin, P., and Fu, C. H. (2011) Hippocampal atrophy in first episode depression: a meta-analysis of magnetic resonance imaging studies. *Journal of affective disorders* 134, 483–487.
- [23] Bath, K. G., Jing, D. Q., Dincheva, I., Neeb, C. C., Pattwell, S. S., Chao, M. V., Lee, F. S., and Ninan, I. (2012) BDNF Val66Met impairs fluoxetine-induced enhancement of adult hippocampus plasticity. *Neuropsychopharmacology* 37, 1297–1304.
- [24] Vetencourt, J. F. M., Sale, A., Viegi, A., Baroncelli, L., De Pasquale, R., O'Leary, O. F., Castrén, E., and Maffei, L. (2008) The antidepressant fluoxetine restores plasticity in the adult visual cortex. *Science 320*, 385– 388.
- [25] Markram, H., Gerstner, W., and Sjöström, P. J. (2011) A history of spike-timing-dependent plasticity. Frontiers in synaptic neuroscience 3.
- [26] Tang, S. J., Reis, G., Kang, H., Gingras, A.-C., Sonenberg, N., and Schuman, E. M. (2002) A rapamycinsensitive signaling pathway contributes to long-term synaptic plasticity in the hippocampus. *Proceedings* of the National Academy of Sciences 99, 467–472.
- [27] Wu, H., Zhou, Y., and Xiong, Z.-Q. (2007) Transducer of regulated CREB and late phase long-term synaptic potentiation. *FEBS J.* 274, 3218–23.
- [28] Orrego, F., and Villanueva, S. (1993) The chemical nature of the main central excitatory transmitter: a critical appraisal based upon release studies and synaptic vesicle localization. *Neuroscience* 56, 539– 55.
- [29] Newsholme, P., Procopio, J., Lima, M. M. R., Pithon-Curi, T. C., and Curi, R. (2003) Glutamine and glutamate-their central role in cell metabolism and function. *Cell Biochem. Funct.* 21, 1–9.
- [30] Plaitakis, A., Berl, S., and Yahr, M. D. (1982) Abnormal glutamate metabolism in an adult-onset degenerative neurological disorder. *Science 216*, 193–6.
- [31] Plaitakis, A., and Caroscio, J. T. (1987) Abnormal glutamate metabolism in amyotrophic lateral sclerosis. *Ann. Neurol.* 22, 575–9.
- [32] Chen, X., Nelson, C. D., Li, X., Winters, C. A., Azzam, R., Sousa, A. A., Leapman, R. D., Gainer, H., Sheng, M., and Reese, T. S. (2011) PSD-95 is required to sustain the molecular organization of the postsynaptic density. *J. Neurosci.* 31, 6329–38.
- [33] Kasai, H., Fukuda, M., Watanabe, S., Hayashi-

Takagi, A., and Noguchi, J. (2010) Structural dynamics of dendritic spines in memory and cognition. *Trends in neurosciences 33*, 121–129.

- [34] Kessels, H. W., and Malinow, R. (2009) Synaptic AMPA receptor plasticity and behavior. *Neuron 61*, 340–350.
- [35] Peng, Y., Zhao, J., Gu, Q.-H., Chen, R.-Q., Xu, Z., Yan, J.-Z., Wang, S.-H., Liu, S.-Y., Chen, Z., and Lu, W. (2010) Distinct trafficking and expression mechanisms underlie LTP and LTD of NMDA receptor-mediated synaptic responses. *Hippocampus* 20, 646–658.
- [36] Morishita, W., and Malenka, R. C. (2008) Mechanisms underlying dedepression of synaptic NMDA receptors in the hippocampus. *Journal of neurophysiol*ogy 99, 254–263.
- [37] Sanacora, G., Zarate, C. A., Krystal, J. H., and Manji, H. K. (2008) Targeting the glutamatergic system to develop novel, improved therapeutics for mood disorders. *Nat Rev Drug Discov* 7, 426–37.
- [38] Greger, I. H., Ziff, E. B., and Penn, A. C. (2007) Molecular determinants of AMPA receptor subunit assembly. *Trends in neurosciences 30*, 407–416.
- [39] Liu, Y., and Zhang, J. (2000) Recent development in NMDA receptors. *Chinese medical journal* 113, 948– 956.
- [40] Lisman, J., Schulman, H., and Cline, H. (2002) The molecular basis of CaMKII function in synaptic and behavioural memory. *Nature Reviews Neuroscience* 3, 175–190.
- [41] Lisman, J., Yasuda, R., and Raghavachari, S. (2012) Mechanisms of CaMKII action in long-term potentiation. *Nature reviews neuroscience* 13, 169–182.
- [42] Souery, D., and der Auwera, K. V. (2004) The multiple facets of treatment-resistant depression. CNS Spectr 9, 803–7.
- [43] Greden, J. F. (2001) The burden of disease for treatment-resistant depression. J Clin Psychiatry 62 Suppl 16, 26–31.
- [44] Souery, D., Papakostas, G. I., and Trivedi, M. H. (2006) Treatment-resistant depression. J Clin Psychiatry 67 Suppl 6, 16–22.
- [45] Fekadu, A., Wooderson, S. C., Markopoulo, K., Donaldson, C., Papadopoulos, A., and Cleare, A. J. (2009) What happens to patients with treatmentresistant depression? A systematic review of medium to long term outcome studies. *J Affect Disord* 116, 4– 11.
- [46] Pfeiffer, P. N., Kim, H. M., Ganoczy, D., Zivin, K., and Valenstein, M. (2013) Treatment-Resistant Depression and Risk of Suicide. *Suicide Life Threat Behav*
- [47] Amital, D., Fostick, L., Silberman, A., Calati, R., Spindelegger, C., Serretti, A., Juven-Wetzler, A., Souery, D., Mendlewicz, J., Montgomery, S., Kasper, S., and Zohar, J. (2012) Physical comorbidity among treatment resistant vs. treatment responsive patients with major depressive disorder. *Eur Neuropsychopharmacol*
- [48] Ivanova, J. I., Birnbaum, H. G., Kidolezi, Y., Subramanian, G., Khan, S. A., and Stensland, M. D. (2010) Direct and indirect costs of employees with treatmentresistant and non-treatment-resistant major depressive disorder. *Curr Med Res Opin 26*, 2475–84.
- [49] Trullas, R., and Skolnick, P. (1990) Functional antagonists at the {NMDA} receptor complex exhibit antidepressant actions. *European Journal of Pharmacology* 185, 1–10.
- [50] Skolnick, P., Layer, R. T., Popik, P., Nowak, G.,

Paul, I. A., and Trullas, R. (1996) Adaptation of Nmethyl-D-aspartate (NMDA) receptors following antidepressant treatment: implications for the pharmacotherapy of depression. *Pharmacopsychiatry 29*, 23–6.

- [51] Pilc, A., Wierońska, J. M., and Skolnick, P. (2013) Glutamate-based antidepressants: preclinical psychopharmacology. *Biol. Psychiatry* 73, 1125–32.
- [52] Berman, R. M., Cappiello, A., Anand, A., Oren, D. A., Heninger, G. R., Charney, D. S., and Krystal, J. H. (2000) Antidepressant effects of ketamine in depressed patients. *Biological Psychiatry* 47, 351–354.
- [53] Zarate, C. A., Singh, J. B., Carlson, P. J., Brutsche, N. E., Ameli, R., Luckenbaugh, D. A., Charney, D. S., and Manji, H. K. (2006) A randomized trial of an N-methyl-D-aspartate antagonist in treatmentresistant major depression. *Arch. Gen. Psychiatry* 63, 856–64.
- [54] Diazgranados, N., Ibrahim, L., Brutsche, N. E., Newberg, A., Kronstein, P., Khalife, S., Kammerer, W. A., Quezado, Z., Luckenbaugh, D. A., Salvadore, G., Machado-Vieira, R., Manji, H. K., and Zarate, C. A. (2010) A randomized add-on trial of an N-methyl-D-aspartate antagonist in treatment-resistant bipolar depression. *Arch. Gen. Psychiatry* 67, 793–802.
- [55] aan het Rot, M., Collins, K. A., Murrough, J. W., Perez, A. M., Reich, D. L., Charney, D. S., and Mathew, S. J. (2010) Safety and efficacy of repeateddose intravenous ketamine for treatment-resistant depression. *Biol. Psychiatry* 67, 139–45.
- [56] Murrough, J. W., Perez, A. M., Pillemer, S., Stern, J., Parides, M. K., Rot, M. A. H., Collins, K. A., Mathew, S. J., Charney, D. S., and Iosifescu, D. V. (2012) Rapid and Longer-Term Antidepressant Effects of Repeated Ketamine Infusions in Treatment-Resistant Major Depression. *Biol. Psychiatry*
- [57] Rot, M. A. H., Zarate, C. A., Charney, D. S., and Mathew, S. J. (2012) Ketamine for depression: where do we go from here? *Biol. Psychiatry* 72, 537–47.
- [58] Price, R. B., Nock, M. K., Charney, D. S., and Mathew, S. J. (2009) Effects of intravenous ketamine on explicit and implicit measures of suicidality in treatment-resistant depression. *Biological psychiatry* 66, 522–526.
- [59] Zarate, C. A., Payne, J. L., Quiroz, J., Sporn, J., Denicoff, K. K., Luckenbaugh, D., Charney, D. S., and Manji, H. K. (2004) An open-label trial of riluzole in patients with treatment-resistant major depression. *Am J Psychiatry* 161, 171–4.
- [60] Burgdorf, J., lei Zhang, X., Nicholson, K. L., Balster, R. L., Leander, J. D., Stanton, P. K., Gross, A. L., Kroes, R. A., and Moskal, J. R. (2013) GLYX-13, a NMDA receptor glycine-site functional partial agonist, induces antidepressant-like effects without ketamine-like side effects. *Neuropsychopharmacol*ogy 38, 729–42.
- [61] Zarate, C. A., Jr., Mathews, D., Ibrahim, L., Chaves, J. F., Marquardt, C., Ukoh, I., Jolkovsky, L., Brutsche, N. E., Smith, M. A., and Luckenbaugh, D. A. (2012) A Randomized Trial of a Low-Trapping Nonselective N-Methyl-D-Aspartate Channel Blocker in Major Depression. *Biological Psychiatry* –.
- [62] Preskorn, S. H., Baker, B., Kolluri, S., Menniti, F. S., Krams, M., and Landen, J. W. (2008) An innovative design to establish proof of concept of the antidepressant effects of the NR2B subunit selective N-methyl-D-aspartate antagonist, CP-101,606, in patients with treatment-refractory major depressive disorder. J Clin Psychopharmacol 28, 631–7.

- [63] Lauterbach, E. C. (2011) Dextromethorphan as a potential rapid-acting antidepressant. *Med. Hypotheses* 76, 717–9.
- [64] Coppola, M., and Mondola, R. (2012) Methoxetamine: from drug of abuse to rapid-acting antidepressant. *Med. Hypotheses 79*, 504–7.
- [65] Paoletti, P., and Neyton, J. (2007) NMDA receptor subunits: function and pharmacology. *Current opinion in pharmacology* 7, 39–47.
- [66] Bardoni, R., Torsney, C., Tong, C.-K., Prandini, M., and MacDermott, A. B. (2004) Presynaptic NMDA receptors modulate glutamate release from primary sensory neurons in rat spinal cord dorsal horn. *The Journal of neuroscience* 24, 2774–2781.
- [67] Moghaddam, B., Adams, B., Verma, A., and Daly, D. (1997) Activation of glutamatergic neurotransmission by ketamine: a novel step in the pathway from NMDA receptor blockade to dopaminergic and cognitive disruptions associated with the prefrontal cortex. *The Journal of neuroscience* 17, 2921–2927.
- [68] Brasier, D. J., and Feldman, D. E. (2008) Synapsespecific expression of functional presynaptic NMDA receptors in rat somatosensory cortex. *The Journal* of *Neuroscience 28*, 2199–2211.
- [69] Tovar, K. R., and Westbrook, G. L. (1999) The incorporation of NMDA receptors with a distinct subunit composition at nascent hippocampal synapses in vitro. *The Journal of neuroscience* 19, 4180–4188.
- [70] Hardingham, G. E., Fukunaga, Y., and Bading, H. (2002) Extrasynaptic NMDARs oppose synaptic NM-DARs by triggering CREB shut-off and cell death pathways. *Nature neuroscience* 5, 405–414.
- [71] Farinelli, M., Heitz, F. D., Grewe, B. F., Tyagarajan, S. K., Helmchen, F., and Mansuy, I. M. (2012) Selective Regulation of NR2B by Protein Phosphatase-1 for the Control of the NMDA Receptor in Neuroprotection. *PloS one* 7, e34047.
- [72] Szatmari, E., Habas, A., Yang, P., Zheng, J.-J., Hagg, T., and Hetman, M. (2005) A positive feedback loop between glycogen synthase kinase 3β and protein phosphatase 1 after stimulation of NR2B NMDA receptors in forebrain neurons. *Journal of Biological Chemistry 280*, 37526–37535.
- [73] Maeng, S., Zarate, C. A., Du, J., Schloesser, R. J., McCammon, J., Chen, G., and Manji, H. K. (2008) Cellular mechanisms underlying the antidepressant effects of ketamine: role of alpha-amino-3-hydroxy-5methylisoxazole-4-propionic acid receptors. *Biol. Psychiatry* 63, 349–52.
- [74] Koike, H., Iijima, M., and Chaki, S. (2011) Involvement of AMPA receptor in both the rapid and sustained antidepressant-like effects of ketamine in animal models of depression. *Behavioural brain research* 224, 107–111.
- [75] Tizabi, Y., Bhatti, B., Manaye, K., Das, J., and Akinfiresoye, L. (2012) Antidepressant-like effects of low ketamine dose is associated with increased hippocampal AMPA/NMDA receptor density ratio in female Wistar–Kyoto rats. *Neuroscience 213*, 72–80.
- [76] Hay, N., and Sonenberg, N. Upstream and downstream of mTOR.
- [77] Jernigan, C. S., Goswami, D. B., Austin, M. C., Iyo, A. H., Chandran, A., Stockmeier, C. A., and Karolewicz, B. (2011) The mTOR signaling pathway in the prefrontal cortex is compromised in major depressive disorder. *Prog. Neuropsychopharmacol. Biol. Psychiatry 35*, 1774–9.
- [78] Li, N., Lee, B., Liu, R.-J., Banasr, M., Dwyer, J. M.,

Iwata, M., Li, X.-Y., Aghajanian, G., and Duman, R. S. (2010) mTOR-dependent synapse formation underlies the rapid antidepressant effects of NMDA antagonists. *Science* 329, 959–64.

- [79] Li, N., Liu, R.-J., Dwyer, J. M., Banasr, M., Lee, B., Son, H., Li, X.-Y., Aghajanian, G., and Duman, R. S. (2011) Glutamate N-methyl-D-aspartate receptor antagonists rapidly reverse behavioral and synaptic deficits caused by chronic stress exposure. *Biol. Psychiatry* 69, 754–61.
- [80] Chandran, A., Iyo, A. H., Jernigan, C. S., Legutko, B., Austin, M. C., and Karolewicz, B. (2013) Reduced phosphorylation of the mTOR signaling pathway components in the amygdala of rats exposed to chronic stress. *Progress in Neuro-Psychopharmacology and Biological Psychiatry* 40, 240–245.
- [81] Mathewb, J. W. M. S. (2010) Overcoming Antidepressant Treatment Resistance: Focus on Glutamate. *Depression: From Psychopathology to Pharmacother*apy 27, 89.
- [82] Smith, M. A., Makino, S., Kvetnansky, R., and Post, R. M. (1995) Stress and glucocorticoids affect the expression of brain-derived neurotrophic factor and neurotrophin-3 mRNAs in the hippocampus. *The Journal of Neuroscience* 15, 1768–1777.
- [83] Nibuya, M., Morinobu, S., and Duman, R. S. (1995) Regulation of BDNF and trkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatments. *The Journal of Neuroscience* 15, 7539–7547.
- [84] Ueyama, T., Kawai, Y., Nemoto, K., Sekimoto, M., Toné, S., and Senba, E. (1997) Immobilization stress reduced the expression of neurotrophins and their receptors in the rat brain. *Neuroscience research 28*, 103–110.
- [85] Barrientos, R., Sprunger, D., Campeau, S., Higgins, E., Watkins, L., Rudy, J., and Maier, S. (2003) Brain-derived neurotrophic factor mRNA downregulation produced by social isolation is blocked by intrahippocampal interleukin-1 receptor antagonist. *Neuroscience* 121, 847–853.
- [86] Roceri, M., Hendriks, W., Racagni, G., Ellenbroek, B., and Riva, M. (2001) Early maternal deprivation reduces the expression of BDNF and NMDA receptor subunits in rat hippocampus. *Molecular psychiatry* 7, 609–616.
- [87] Roceri, M., Cirulli, F., Pessina, C., Peretto, P., Racagni, G., and Riva, M. A. (2004) Postnatal repeated maternal deprivation produces agedependent changes of brain-derived neurotrophic factor expression in selected rat brain regions. *Biological Psychiatry 55*, 708–714.
- [88] Chen, B., Dowlatshahi, D., MacQueen, G. M., Wang, J.-F., and Young, L. T. (2001) Increased hippocampal BDNF immunoreactivity in subjects treated with antidepressant medication. *Biological psychiatry* 50, 260–265.
- [89] Dwivedi, Y., Mondal, A. C., Rizavi, H. S., and Conley, R. R. (2005) Suicide Brain Is Associated with Decreased Expression of Neurotrophins . *Biological Psychiatry 58*, 315–324.
- [90] Karege, F., Vaudan, G., Schwald, M., Perroud, N., and Harpe, R. L. (2005) Neurotrophin levels in postmortem brains of suicide victims and the effects of antemortem diagnosis and psychotropic drugs. *Molecular Brain Research* 136, 29–37.
- [91] Karege, F., Perret, G., Bondolfi, G., Schwald, M., Bertschy, G., and Aubry, J.-M. (2002) Decreased serum brain-derived neurotrophic factor levels in ma-

jor depressed patients . *Psychiatry Research 109*, 143–148.

- [92] Shimizu, E., Hashimoto, K., Okamura, N., Koike, K., Komatsu, N., Kumakiri, C., Nakazato, M., Watanabe, H., Shinoda, N., ichi Okada, S., and Iyo, M. (2003) Alterations of serum levels of brain-derived neurotrophic factor (BDNF) in depressed patients with or without antidepressants . *Biological Psychiatry 54*, 70–75.
- [93] Gervasoni, N., Aubry, J.-M., Bondolfi, G., Osiek, C., Schwald, M., Bertschy, G., and Karege, F. (2005) Partial normalization of serum brain-derived neurotrophic factor in remitted patients after a major depressive episode. *Neuropsychobiology* 51, 234–238.
- [94] Aydemir, O., Deveci, A., and Taneli, F. (2005) The effect of chronic antidepressant treatment on serum brain-derived neurotrophic factor levels in depressed patients: a preliminary study . *Progress in Neuro-Psychopharmacology and Biological Psychiatry 29*, 261–265.
- [95] Machado-Vieira, R., Yuan, P., Brutsche, N., Diaz-Granados, N., Luckenbaugh, D., Manji, H. K., and Zarate, C. A. (2009) Brain-derived neurotrophic factor and initial antidepressant response to an N-methyl-Daspartate antagonist. *J Clin Psychiatry 70*, 1662–6.
- [96] Ricci, V., Martinotti, G., Gelfo, F., Tonioni, F., Caltagirone, C., Bria, P., and Angelucci, F. (2011) Chronic ketamine use increases serum levels of brain-derived neurotrophic factor. *Psychopharmacology (Berl.)* 215, 143–8.
- [97] Kovalchuk, Y., Hanse, E., Kafitz, K. W., and Konnerth, A. (2002) Postsynaptic Induction of BDNF-Mediated Long-Term Potentiation. *Science* 215, 1729–1733.
- [98] Dragunow, M., Beilharz, E., Mason, B., Lawlor, P., Abraham, W., and Gluckman, P. (1993) Brain-derived neurotrophic factor expression after long-term potentiation. *Neuroscience letters* 160, 232–236.
- [99] Patapoutian, A., and Reichardt, L. F. (2001) Trk receptors: mediators of neurotrophin action. *Current opin*ion in neurobiology 11, 272–280.
- [100] Kolarow, R., Brigadski, T., and Lessmann, V. (2007) Postsynaptic secretion of BDNF and NT-3 from hippocampal neurons depends on calcium-calmodulin kinase II signaling and proceeds via delayed fusion pore opening. *The Journal of neuroscience 27*, 10350–10364.
- [101] Katoh-Semba, R., Asano, T., Ueda, H., Morishita, R., Takeuchi, I. K., Inaguma, Y., and Kato, K. (2002) Riluzole enhances expression of brain-derived neurotrophic factor with consequent proliferation of granule precursor cells in the rat hippocampus. *FASEB J. 16*, 1328–30.
- [102] Ma, T., Tzavaras, N., Tsokas, P., Landau, E. M., and Blitzer, R. D. (2011) Synaptic stimulation of mTOR is mediated by Wnt signaling and regulation of glycogen synthetase kinase-3. *The Journal of Neuroscience* 31, 17537–17546.
- [103] Nelson, C. D., Kim, M. J., Hsin, H., Chen, Y., and Sheng, M. (2013) Phosphorylation of Threonine-19 of PSD-95 by GSK-3β is Required for PSD-95 Mobilization and Long-Term Depression. J. Neurosci. 33, 12122–35.
- [104] Beurel, E., Song, L., and Jope, R. (2011) Inhibition of glycogen synthase kinase-3 is necessary for the rapid antidepressant effect of ketamine in mice. *Molecular psychiatry* 16, 1068–1070.
- [105] Müller, H. K., Wegener, G., Liebenberg, N.,

Zarate, C. A., Jr., Popoli, M., and Elfving, B. (2013) Ketamine regulates the presynaptic release machinery in the hippocampus. *Journal of Psychiatric Research* 47, 892–899.

- [106] Bramham, C. R., Worley, P. F., Moore, M. J., and Guzowski, J. F. (2008) The immediate early gene arc/arg3.1: regulation, mechanisms, and function. *J. Neurosci.* 28, 11760–7.
- [107] Chowdhury, S., Shepherd, J. D., Okuno, H., Lyford, G., Petralia, R. S., Plath, N., Kuhl, D., Huganir, R. L., and Worley, P. F. (2006) Arc/Arg3.1 interacts with the endocytic machinery to regulate AMPA receptor trafficking. *Neuron 52*, 445–59.
- [108] Hahn-Windgassen, A., Nogueira, V., Chen, C.-C., Skeen, J. E., Sonenberg, N., and Hay, N. (2005) Akt activates the mammalian target of rapamycin by regulating cellular ATP level and AMPK activity. *Journal* of *Biological Chemistry* 280, 32081–32089.
- [109] You, J. S., Frey, J. W., and Hornberger, T. A. (2012) Mechanical Stimulation Induces mTOR Signaling via an ERK-Independent Mechanism: Implications for a Direct Activation of mTOR by Phosphatidic Acid. *PloS* one 7, e47258.
- [110] Slipczuk, L., Bekinschtein, P., Katche, C., Cammarota, M., Izquierdo, I., and Medina, J. H. (2009) BDNF activates mTOR to regulate GluR1 expression required for memory formation. *PLoS One* 4, e6007.
- [111] Duman, R. S., Li, N., Liu, R.-J., Duric, V., and Aghajanian, G. (2012) Signaling pathways underlying the rapid antidepressant actions of ketamine. *Neuropharmacology* 62, 35–41.
- [112] Wang, X., Li, W., Williams, M., Terada, N., Alessi, D. R., and Proud, C. G. (2001) Regulation of elongation factor 2 kinase by p90RSK1 and p70 S6 kinase. *The EMBO journal 20*, 4370–4379.
- [113] Hoeffer, C. A., and Klann, E. (2010) mTOR signaling: at the crossroads of plasticity, memory and disease. *Trends Neurosci.* 33, 67–75.
- [114] Yang, Q., Liao, Z.-H., Xiao, Y.-X., Lin, Q.-S., Zhu, Y.-S., and Li, S.-T. (2011) Hippocampal synaptic metaplasticity requires the activation of NR2B-containing NMDA receptors. *Brain Res. Bull.* 84, 137–43.
- [115] Li, X., Alafuzoff, I., Soininen, H., Winblad, B., and Pei, J.-J. (2005) Levels of mTOR and its downstream targets 4E-BP1, eEF2, and eEF2 kinase in relationships with tau in Alzheimer's disease brain. *Febs Journal 272*, 4211–4220.
- [116] Chano, T., Okabe, H., and Hulette, C. M. (2007) RB1CC1 insufficiency causes neuronal atrophy through mTOR signaling alteration and involved in the pathology of Alzheimer's diseases. *Brain research 1168*, 97–105.
- [117] Rosner, M., Hanneder, M., Siegel, N., Valli, A., Fuchs, C., and Hengstschläger, M. (2008) The mTOR pathway and its role in human genetic diseases. *Mutation Research/Reviews in Mutation Research 659*, 284–292.
- [118] Chan, K. W. S., Lee, T. M. C., Siu, A. M. H., Wong, D. P. L., Kam, C.-M., Tsang, S. K. M., and Chan, C. C. H. (2013) Effects of chronic ketamine use on frontal and medial temporal cognition. *Addict Behav* 38, 2128–32.
- [119] Preskorn, S. H. (2012) Ketamine: the hopes and the hurdles. *Biol. Psychiatry 72*, 522–3.
- [120] Noppers, I., Niesters, M., Aarts, L., Smith, T., Sarton, E., and Dahan, A. (2010) Ketamine for the treatment of chronic non-cancer pain. *Expert opinion on pharmacotherapy* 11, 2417–2429.